



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 5 : C07K 15/00, C12Q 1/70 C12P 21/02, C12N 5/00	A1	(11) International Publication Number: WO 92/08738 (43) International Publication Date: 29 May 1992 (29.05.92)
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(22) International Filing Date: 7 November 1991 (07.11.91)		(81) Designated States: AT (European patent), AU, BE (European patent), CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FR (European patent), GB (European patent), GR (European patent), IT (European patent), JP, KR, LU (European patent), NL (European patent), SE (European patent).
(30) Priority data: 610,175 7 November 1990 (07.11.90) US 648,473 31 January 1991 (31.01.91) US 648,477 31 January 1991 (31.01.91) US		Published <i>With international search report.</i>
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(54) Title: MONOCLONAL ANTIBODIES TO HEPATITIS C VIRUS AND METHOD FOR USING SAME		
(57) Abstract		
Monoclonal antibodies which specifically bind to either Hepatitis C Virus C-100 protein, Hepatitis C Virus 33C protein and Hepatitis C Virus CORE protein, and hybridomas which produce these monoclonal antibodies. Also provided are methods for using these monoclonal antibodies and assay kits containing these antibodies.		

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MONOCLONAL ANTIBODIES TO HEPATITIS C VIRUS AND METHOD FOR USING SAME5 BACKGROUND OF THE INVENTION

This invention relates generally to antibodies which specifically bind to Hepatitis C Virus (HCV), and more specifically, relates to a panel of novel hybridoma cells lines which secrete monoclonal antibodies to HCV proteins C-100, 33C and CORE, and methods for using these monoclonal antibodies.

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Descriptions of Hepatitis diseases causing jaundice and icterus have been known to man since antiquity. Viral hepatitis is now known to include a group of viral agents with distinctive viral organization protein structure and mode of replication, causing hepatitis with different degrees of severity of hepatic damage through different routes of transmission. Acute viral hepatitis is clinically diagnosed by well-defined patient symptoms including jaundice, hepatic tenderness and an elevated level of liver transaminases such as Aspartate Transaminase and Alanine Transaminase.

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Serological assays currently are employed to further distinguish between Hepatitis-A and Hepatitis-B. Non-A Non-B Hepatitis (NANBH) is a term first used in 1975 that described cases of post-transfusion hepatitis not caused by either Hepatitis A Virus or Hepatitis B Virus. Feinstone et al., New Engl. J. Med. 292:454-457 (1975). The diagnosis of NANBH has been made primarily by means of exclusion on the basis of serological analysis for the presence of Hepatitis A and Hepatitis B. NANBH is responsible for about 90% of the cases of post-transfusion hepatitis. Hollinger et al. in N. R. Rose et al., eds., Manual of Clinical Immunology, American Society for Microbiology, Washington, D. C., 558-572 (1986).

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Attempts to identify the NANBH virus by virtue of genomic similarity to one of the known hepatitis viruses have failed thus far, suggesting that NANBH has a distinctive genomic organization and structure. Fowler et al., J. Med. Virol. 12:205-213 (1983), and Weiner et al., J. Med. Virol. 21:239-247 (1987). Progress in developing assays to detect antibodies specific for NANBH has been hampered by difficulties encountered in identifying antigens associated with the virus. Wards et al., U. S. Patent No. 4,870,076; Wards et al., Proc. Natl. Acad. Sci. 83:6608-6612 (1986); Ohori et al., J. Med. Virol. 12:161-178 (1983); Bradly et al., Proc. Natl. Acad. Sci. 84:6277-6281 (1987); Akatsuka et al., J. Med. Virol. 20:43-56 (1986).

In May of 1988, a collaborative effort of Chiron Corporation with the Centers for Disease Control resulted in the identification of a putative NANB agent, Hepatitis C Virus (HCV). M. Houghton et al. cloned and expressed in E. coli a NANB agent obtained from the infectious plasma of a chimp. Cuo et al., Science 244:359-361 (1989); Choo et al., Science 244:362-364 (1989). CDNA sequences from HCV were identified which encode antigens that react immunologically with antibodies present in a majority of the patients clinically diagnosed with NANBH. Based on the information available and on the molecular structure of HCV, the genetic makeup of the virus consists of single stranded linear RNA (positive strand) of molecular weight approximately 9.5 kb, and possessing one continuous translational open reading frame. J. A. Cuthbert, Amer. J. Med. Sci. 299:346-355 (1990). It is a small enveloped virus resembling the Flaviviruses. Investigators have made attempts to identify the NANB agent by ultrastructural changes in hepatocytes in infected individuals. H. Gupta, Liver 8:111-115 (1988); D.W. Bradley J. Virol. Methods 10:307-319 (1985). Similar ultrastructural changes in hepatocytes as well as PCR amplified HCV RNA sequences have been detected in NANBH patients as well as in chimps experimentally infected with infectious HCV plasma. T. Shimizu et al., Proc. Natl. Acad. Sci. 87:6441-6444 (1990).

Considerable serological evidence has been found to implicate HCV as the etiological agent for post-transfusion NANBH. H. Alter et al., N. Eng. J. Med. 321:1494-1500 (1989); Estaben et al., The Lancet: Aug. 5:294-296 (1989); C. Van Der Poel et al., The Lancet Aug. 5:297-298 (1989); G. Sbolli, J. Med. Virol. 30:230-232 (1990); M. Makris et al., The Lancet 335:1117-1119 (1990). Although the detection of HCV antibodies eliminates 70 to 80% of NANBH infected blood from the blood supply system, the antibodies apparently are readily detected during the chronic state of the disease, while only 60% of the samples from the acute NANBH stage are HCV antibody positive. H. Alter et al., New Eng. J. Med. 321:1494-1500 (1989). The prolonged interval between exposure to HCV and antibody detection, and the lack of adequate information regarding the profile of immune response to various structural and non-structural proteins raises questions regarding the infectious state of the patient in the latent and antibody negative phase during NANBH infection. Therefore, there is a need for the development of assay systems to identify acute infection and viremia which may be present. Tools are needed to distinguish between acute and persistent infection and to define the prognostic course of NANBH infection, in order to develop preventive strategies.

SUMMARY OF THE INVENTION

The present invention provides a panel of highly specific and novel monoclonal antibodies that can be employed for the detection of Hepatitis C viral proteins. The monoclonal antibodies specifically bind to either C-100, 33C or CORE antigens, and do not significantly bind to 33C and CORE, C-100 and CORE, and C-100 and 33C, respectively. The hybridomas which produce (secrete) these monoclonal antibodies are identified as follows: H81C17 (A.T.C.C. deposit No. HB 10588, producing monoclonal antibody H81C17), H35C54 (A.T.C.C. deposit No. HB 10592, producing monoclonal antibody H35C54), H28C110 (A.T.C.C. deposit No. HB 10587, producing monoclonal antibody H28C110), H4C20 (A.T.C.C. deposit No. HB 10593, producing monoclonal antibody H4C20), H11C130 (A.T.C.C. deposit No. HB 10589, producing monoclonal antibody H11C130), H1C46 (A.T.C.C. deposit No. HB 10594, producing monoclonal antibody H1C46), 13-975-157 (A.T.C.C. deposit No. HB 10608, producing monoclonal antibody 13-975-157), 14-153-234 (A.T.C.C. deposit No. 10604, producing monoclonal antibody 14-153-234), 14-1350-210 (A.T.C.C. deposit No. HB 10602, producing monoclonal antibody 14-1350-210), 6-296-534 (A.T.C.C. deposit No. HB 10607, secreting monoclonal antibody 6-296-534), and 6-914-518 (A.T.C.C. deposit No. HB 10600, secreting monoclonal antibody 6-914-518). The specificity of these monoclonal antibodies enables advantageous differentiation studies, as well as prognostic and diagnostic applications in the diagnosis and evaluation of NANBH.

In a preferred assay format, a test sample which may contain HCV antigens is contacted with a solid phase to which a polyclonal or a monoclonal anti-HCV antibody or a fragment thereof has been bound, to form a mixture. This mixture is incubated for a time and under conditions sufficient for antigen/antibody complexes to form. The so-formed complexes then are contacted with an indicator reagent comprising a monoclonal or polyclonal antibody or a fragment thereof, specific for the HCV antigen attached to a signal generating compound to form a second mixture. This second mixture is reacted for a time and under conditions sufficient to form antibody/antigen/antibody complexes. The presence of HCV antigen is determined by detecting the measurable signal generated. The amount of HCV present in the test sample, thus the amount of HCV antigen captured on the solid phase, is proportional to the amount of signal generated.

Alternatively, an indicator reagent comprising a monoclonal or polyclonal antibody, or fragment thereof, specific for HCV and a signal generating compound is added

to a polyclonal or monoclonal anti-HCV antibody or fragment thereof coated on a solid phase and the test sample to form a mixture. This mixture is incubated for a time and under conditions sufficient to form antibody/antigen/antibody complexes. The presence and amount of HCV present in the test sample, and thus the amount of HCV antigen captured on the solid phase, is determined by detecting the measurable signal. The amount of HCV present in the test sample is proportional to the amount of signal generated.

In another alternate assay format, one or a combination of more than one monoclonal antibody of the invention can be employed as a competitive probe for the detection of antibodies to HCV antigen. For example, HCV CORE antigens, either alone or in combination, can be coated on a solid phase. A test sample suspected of containing antibody to HCV CORE antigen then is incubated with an indicator reagent comprising a signal generating compound and a monoclonal antibody of the invention for a time and under conditions sufficient to form antigen/antibody complexes of either the test sample and indicator reagent to the solid phase or the indicator reagent to the solid phase. The reduction in binding of the monoclonal antibody to the solid phase can be quantitatively measured. A measurable reduction in the signal compared to the signal generated from a confirmed negative NANBH test sample would indicate the presence of anti-HCV CORE antibody in the test sample.

In yet another assay format, a test sample is contacted with a solid phase to which Hepatitis C Virus proteins are attached and an indicator reagent comprising a monoclonal antibody or fragment thereof specific for Hepatitis C Virus attached to a signal generating compound, to form a mixture. The mixture is incubated for a time and under conditions sufficient for antibody/antigen complexes to form. The presence of anti-Hepatitis C virus present in the test sample is determined by detecting the measurable signal generated, and comparing the signal to the measured signal generated from a known negative sample. A measurable reduction of signal of the test sample, compared to the known negative sample's signal, is indicative of the presence of anti-HCV antibodies. Competitive assays for the detection of anti-HCV antibody using antigens free in solution also can be performed.

The presence of Hepatitis C Virus can be detected in a tissue sample by contacting the tissue sample with an indicator reagent comprising a signal generating compound attached to a monoclonal antibody selected from the group consisting of an anti-HCV

C100 antibody or fragment thereof, a monoclonal anti-HCV 33C antibody or fragment thereof, and an anti-HCV CORE antibody or fragment thereof, to form a mixture. This mixture is incubated for a time and under conditions sufficient for antigen/antibody complex to form. The presence of Hepatitis C Virus present in the tissue sample is determined by detecting the signal generated.

Also provided are kits for using the monoclonal antibodies of the invention.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1A is a drawing of the location of the location of the recombinant proteins on the HCV genome, employed as immunogens for generation of the hybridomas of the invention as well as that of subfragments employed for epitope mapping of the monoclonal antibodies of the invention.

FIG. 1B is a map of the HCV GENOME representing the non-structural (NS) genes and the structural genes, core (C) and envelope (E).

FIG. 2A is a photograph of Western blot analysis illustrating the specific binding and epitope mapping of monoclonal antibody H11C130 wherein lane 1 is pHCV29 (a.a. 1192-1457), lane 2 is 33Ca·CKS (a.a. 1192-1331), lane 3 is 33Cb·CKS (a.a. 1330-1457), lane 4 is 33C (λ pL) (a.a. 1192-1457) and lane 5 is CKS;

FIG. 2B is a photograph of Western blot analysis illustrating the specific binding and epitope mapping of monoclonal antibody H1C46 wherein lane 1 is pHCV29 (a.a. 1192-1457), lane 2 is 33Ca·CKS (a.a. 1192-1331), lane 3 is 33Cb·CKS (a.a. 1330-1457), lane 4 is 33C (λ pL) (a.a. 1192-1457) and lane 5 is CKS;

FIG. 2C is a photograph of Western blot analysis illustrating the specific binding of monoclonal antibody H81C17 wherein lane 1 is pHCV34 (a.a. 1-150), lane 2 is HCVCORE (λ pL) and lane 3 is CKS;

FIG. 2D is a photograph of Western blot analysis illustrating the specific binding of monoclonal antibody H35C54 wherein lane 1 is pHCV34 (a.a. 1-150), lane 2 is HCVCORE (λ pL) and lane 3 is CKS;

FIG. 2E is a photograph of Western blot analysis illustrating specific binding and epitope mapping of monoclonal antibody H28C110 wherein lane 1 is C-100B·CKS (a.a. 1676-1790), lane 2 is C-100 C·CKS (a.a. 1789-1863), lane 3 is C-100D·CKS (a.a. 1861-1931), lane 4 is C-100 (λ pL; a.a. 1676-1790) and lane 5 is CKS;

FIG. 2F is a photograph of Western blot analysis illustrating specific binding and epitope mapping of monoclonal antibody H4C20 wherein lane 1 is C-100B·CKS (a.a.

1676-1790), lane 2 is C-100-C-CKS (a.a. 1789-1863), lane 3 is C-100D-CKS (a.a. 1861-1931), lane 4 is C-100 (λ pL, a.a. 1676-1790) and lane 5 is CKS.

FIG. 3 is a profile of PEPSCAN analysis with overlapping hexamer peptides (a.a. 5 1694-1735) of monoclonal antibody H28C110 illustrating the epitope specificity and amino acid sequence of the HCV genome recognized by monoclonal antibody H28C110.

FIG. 4 is a graph illustrating the efficiency of anti-HCV rabbit and human 10 polyclonal capture antibodies for the detection of HCV core protein in the antigen capture assay of the invention employing monoclonal antibody H81C17.

FIG. 5 is a graph of a negative sera and plasma population distribution tested by an antigen capture EIA wherein frequency is plotted against A492.

15 FIGS. 6 to 17 are photographs of Western blots showing the reactivity of the monoclonal antibodies of the invention, where

lanes 1 to 3 contain monoclonal antibodies against HCV 33C protein (6-296-534 in lane 1, 6-914-518 in lane 2 and 6-1070-110 in lane 3);

20 lanes 4-6 contain monoclonal antibodies against HCV CORE (13-975-157 in lane 4, 14-153-234 in lane 5 and 14-1350-210 in lane 6);

lanes 7 and 8 contain monoclonal antibodies against the putative HCV ENV region (16-407-209 in lane 7 and 16-803-174 in lane 8);

lanes 9-10 contain monoclonal antibodies against HCV C-100 (25-1518-105 in lane 9, 28-735-355 in lane 10);

25 lane 11 contains monoclonal antibody against CKS (29-121-236 in lane 11);

lane 12 contains a normal mouse serum control; and

lane 13 contains a negative control of antibody diluent.

FIG. 6 is an electroblot of these monoclonal antibodies run against CKS-CORE;

FIG. 7 is an electroblot of these monoclonal antibodies run against λ PL-CORE;

30 FIG. 8 is an electroblot of these monoclonal antibodies run against λ PL-33C-CORE;

FIG. 9 is an electroblot of these monoclonal antibodies run against CKS-33C;

FIG. 10 is an electroblot of these monoclonal antibodies run against CKS-33C-BCD;

35 FIG. 11 is an electroblot of these monoclonal antibodies run against CKS-BCD;

FIG. 12 is an electroblot of these monoclonal antibodies run against CKS-B;

FIG. 13 is an electroblot of these monoclonal antibodies run against CKS-E;
FIG. 14 is an electroblot of these monoclonal antibodies run against CKS;
FIG. 15 is an electroblot of these monoclonal antibodies run against SOD-100;
FIG. 16 is an electroblot of these monoclonal antibodies run against CKS-A'BCD;

5 and

FIG. 17 is an electroblot of these monoclonal antibodies run against CKS-A"BCD.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides novel monoclonal antibodies to HCV proteins C-
10 100, 33C and CORE, methods for using the monoclonal antibodies, and kits which contain
these monoclonal antibodies.

The monoclonal antibodies of the present invention can be employed in various
assay systems to determine the presence, if any, of any or all of HCV proteins C-100,
15 33C or CORE, or a combination thereof. Fragments of the monoclonal antibodies provided
also may be used. For example, in a first assay format, a polyclonal or monoclonal
anti-HCV-100, anti-33C or anti-CORE antibody or a fragment thereof, or a
combination thereof, which has been coated on a solid phase, is contacted with a test
sample which may contain any or all of these HCV proteins or a combination of them, to
20 form a mixture. This mixture is incubated for a time and under conditions sufficient to
form antigen/antibody complexes. Then, an indicator reagent comprising a monoclonal
or a polyclonal antibody or a fragment thereof, which specifically binds to either HCV C-
100 protein, HCV 33C protein or CORE protein, or a combination of these antibodies, to
which a signal generating compound has been attached, is contacted with the
25 antigen/antibody complexes to form a second mixture. This second mixture then is
incubated for a time and under conditions sufficient to form antibody/antigen/antibody
complexes. The presence of HCV protein present in the test sample and captured on the
solid phase, if any, is determined by detecting the measurable signal generated by the
signal generating compound. The amount of HCV protein present in the test sample is
30 proportional to the signal generated.

Alternatively, a polyclonal or monoclonal anti-HCV C-100, anti-HCV 33C or
anti-HCV CORE antibody or fragment thereof, or a combination of these antibodies which
is bound to a solid support, the test sample and an indicator reagent comprising a
35 monoclonal or polyclonal antibody or fragments thereof, which specifically binds to
either HCV C-100, HCV C33, or HCV CORE protein, or a combination of these antibodies

to which a signal generating compound is attached, are contacted to form a mixture. This mixture is incubated for a time and under conditions sufficient to form antibody/antigen/antibody complexes. The presence, if any, of HCV protein(s) present in the test sample and captured on the solid phase is determined by detecting the measurable signal generated by the signal generating compound. The amount of HCV protein(s) present in the test sample is proportional to the signal generated.

In another alternate assay format, one or a combination of more than one monoclonal antibody of the invention can be employed as a competitive probe for the detection of antibodies to HCV protein C-100, 33C or CORE. For example, HCV proteins, either alone or in combination, can be coated on a solid phase. A test sample suspected of containing antibody to Hepatitis C Virus then is incubated with an indicator reagent comprising a signal generating compound and a monoclonal antibody of the invention for a time and under conditions sufficient to form antigen/antibody complexes of either the test sample and indicator reagent to the solid phase or the indicator reagent to the solid phase. The reduction in binding of the monoclonal antibody to the solid phase can be quantitatively measured. A measurable reduction in the signal compared to the signal generated from a confirmed negative NANBH test sample would indicate the presence of anti-HCV antibody in the test sample.

In yet another detection method, each of the monoclonal antibodies of the present invention can be employed in the detection of HCV antigens in fixed tissue sections, as well as fixed cells by immunohistochemical analysis.

In addition, these monoclonal antibodies can be bound to matrices similar to CNBr-activated sepharose and used for the affinity purification of specific HCV proteins from cell cultures, or biological tissues such as blood and liver.

The monoclonal antibodies of the invention can also be used for the generation of chimeric antibodies for therapeutic use, or other similar applications.

The monoclonal antibodies or fragments thereof can be provided individually to detect either HCV C-100, HCV 33C or HCV CORE protein. Combinations of the monoclonal antibodies (and fragments thereof) provided herein also may be used together as components in a mixture or "cocktail" of anti-HCV protein antibodies, each having different binding specificities. Thus, this cocktail can include the monoclonal

antibodies of the invention directed to different antigenic determinants of the HCV genome, along with monoclonal antibodies directed to putative HCV ENV region.

The polyclonal antibody or fragment thereof which can be used in the assay formats should specifically bind to either HCV C-100 protein, HCV 33C protein or HCV CORE protein. The polyclonal antibody used preferably is of mammalian origin; human, goat, rabbit or sheep anti-HCV polyclonal antibody can be used. Most preferably, the polyclonal antibody is rabbit polyclonal anti-HCV antibody. The polyclonal antibodies used in the assays can be used either alone or as a cocktail of polyclonal antibodies. Since the cocktails used in the assay formats are comprised of either monoclonal antibodies or polyclonal antibodies having different HCV specificity, they would be useful for diagnosis, evaluation and prognosis of HCV infection, as well as for studying HCV protein differentiation and specificity.

Test samples which can be tested by the methods of the present invention described herein include human and animal body fluids such as whole blood, serum, plasma, cerebrospinal fluid, urine, biological fluids such as cell culture supernatants, fixed tissue specimens and fixed cell specimens. Solid supports are known to those in the art and include the walls of wells of a reaction tray, test tubes, polystyrene beads, magnetic beads, nitrocellulose strips, membranes, microparticles such as latex particles, and others.

The indicator reagent comprises a signal generating compound (label) which is capable of generating a measurable signal detectable by external means conjugated (attached) to a specific binding member for HCV. "Specific binding member" as used herein means a member of a specific binding pair. That is, two different molecules where one of the molecules through chemical or physical means specifically binds to the second molecule. In addition to being an antibody member of a specific binding pair for HCV, the indicator reagent also can be a member of any specific binding pair, including either hapten-anti-hapten systems such as biotin or anti-biotin, avidin or biotin, a carbohydrate or a lectin, a complementary nucleotide sequence, an effector or a receptor molecule, an enzyme cofactor and an enzyme, an enzyme inhibitor or an enzyme, and the like. An immunoreactive specific binding member can be an antibody, an antigen, or an antibody/antigen complex that is capable of binding either to HCV as in a sandwich assay, to the capture reagent as in a competitive assay, or to the ancillary specific binding member as in an indirect assay.

The various signal generating compounds (labels) contemplated include chromogens, catalysts such as enzymes, luminescent compounds such as fluorescein and rhodamine, chemiluminescent compounds, radioactive elements, and direct visual labels.

5 Examples of enzymes include alkaline phosphatase, horseradish peroxidase, beta-galactosidase, and the like. The selection of a particular label is not critical, but it will be capable of producing a signal either by itself or in conjunction with one or more additional substances.

10 It is contemplated that the reagent employed for the assay can be provided in the form of a kit with one or more containers such as vials or bottles, with each container containing a separate reagent such as a monoclonal antibody, or a cocktail of monoclonal antibodies, employed in the assay.

15 The following examples demonstrate the advantages and utility of this invention for serodiagnosis of Hepatitis C virus by describing methods for the development, characterization, epitope mapping and clinical utility of these monoclonal antibodies. The methods used for monoclonal antibody development follow procedures known in the art and detailed in Kohler and Milstein, *Nature* 256:494 (1975), and reviewed in

20 J.G.R. Hurrel, ed., *Monoclonal Hybridoma Antibodies: Techniques and Applications*, CRC Press, Inc., Boca Raton, FL (1982). Another method of monoclonal antibody development which is based on the Kohler and Milstein method is that of L.T. Mimms et al., *Virology* 176:604-619 (1990), which is incorporated herein by reference. These examples are meant to illustrate, but not to limit, the spirit and scope of the invention.

25 Examples 1-8 exemplify the production and uses of cell lines 81C17, H35C54, H28C110, H4C20, H11C130 and H1C46. Examples 9-13 exemplify the production and uses of cell lines 13-975-257, 14-153-234 and 14-1350-210. Examples 14-17 exemplify the production and uses of cell lines 6-296-534, 6-914-518 and 6-1070-
30 110.

EXAMPLES

EXAMPLE 1

Immunization of Mice

35 *E. coli* derived recombinant antigens encoded by HCV sequences, designated as pHCV23 (HCV C-100, a.a. 1677-1931), pHCV29 (HCV 33C, a.a. 1192-1457) and

pHCV34 (HCV-CORE, a.a. 1-150) were employed as immunogens for the generation of murine monoclonal antibodies specific for HCV C-100, 33C and CORE, respectively. Detailed information on the synthesis, cloning and expression of these recombinant proteins is disclosed in U.S. Patent Application Serial No. 07/572,822, which enjoys common ownership and is incorporated herein by reference. These proteins were prepared for immunization with appropriate adjuvants after purification with protein purification methods known to those skilled in the art. FIG. 1A shows the location of recombinant HCV proteins and their subfragments on the genome.

10

Immunization with pHCV23

On day one, BALB/c mice received 15 μ g of purified pHCV23 in 200 μ l of Freund's complete adjuvant injected intraperitoneally (i.p.). A second immunization was done after 14 days with 15 μ g of pHCV23 in incomplete Freund's adjuvant. Mice were bled on day 21 and the immune response to pHCV was assessed by enzyme linked immunoassay (EIA) and Western blot analysis. Fusion was performed after allowing the mice to rest for at least eight weeks.

15

Immunization with pHCV29

On day one, BALB/c mice received 15 μ g of purified pHCV29 in 100 μ l of Freund's complete adjuvant injected i.p. Subsequent immunizations were done after 14 and 28 days with 15 μ g of pHCV23 in incomplete Freund's adjuvant. Mice were bled on day 21 and the immune response to pHCV23 was assessed as described above.

20

Immunization with pHCV34

BALB/c mice were immunized following a schedule similar to that of pHCV29 immunization described above, using RIBI adjuvant system (RIBI Immunochem Research, Hamilton, Montana). On day one, mice received 15 μ g of purified pHCV34 with 15 μ g each of Trehalose dimycolate (TDM) and M. phlei in a buffer emulsion prepared according to the manufacturer's instructions. Subsequent immunizations were performed on day 14, 28 and 42. Mice were bled on days 21 and 49 and the immune response was assessed as described hereinbelow.

25

Enzyme-Linked Immunoassay (EIA)

The immune response to the immunizing antigen was assessed by microtiter EIA and Western blot analysis. Wells of microtiter plates were coated with 100 μ l of purified antigen in 0.1M bicarbonate buffer at pH 9.5. After washing with Phosphate

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Buffered Saline (PBS) which also contained 0.01% sodium dodecyl sulfate (SDS) and 0.05% Tween-20® (available from Biorad Laboratories, Richmond, CA), free sites were overcoated with 1% BSA in bicarbonate buffer at pH 9.5. Plates were stored at 4°C following a final wash. Sera from native or immunized mice were serially diluted in

- 5 100 µl of dilution buffer which contained 20 mM sodium phosphate, pH 7.4, 0.15M NaCl, 20% normal goat serum, 10% fetal calf serum, 5 mM EDTA, 10 mM EGTA, 50 mM Tris, 0.2% Tween-20®, with sodium azide as a preservative (at pH 6.8). The diluted sera were reacted with the antigen for three (3) hours at 37°C. The plates were washed and 100 µl of appropriately diluted goat anti-mouse IgG (heavy (H) and light (L) chain)
- 10 Horseradish Peroxidase (HRPO)-conjugated antibody (Jackson Immunochemicals, West Grove, PA) was added. The plates were incubated at 37°C for two (2) hours. After a final wash, 100 µl of o-phenylenediamine:2HCl (OPD) color reagent was added. The reaction was carried out at room temperature for 10 to 30 minutes, and then stopped by the addition of 1 ml of 1N H₂SO₄. The absorbance at 492/600 nm was recorded, which
- 15 was found to be directly proportional to the amount of specific antibody bound to the respective antigen.

Western Blot Analysis

- Approximately 300 µg of purified rHCV protein were treated with SDS and 2-20 mercaptoethanol at 95°C, and electrophoresed in a 12% polyacrylamide-SDS gel (Laemmli et al., *Nature* 227:680-685 (1970)). Proteins were transferred overnight from the gel to nitrocellulose by electrophoresis at 100 mamp, or transferred in 1-2 hours at 1.0 amp, in a standard transfer buffer which comprised 25 mM Tris [(Hydroxymethyl) Aminomethane], 192 mM glycine, and 2.0% methanol, pH 8.3.
- 25 (Towbin et al., *Proc. Natl. Acad. Sci.* 73:4350-4354 [1979]). After transferring the proteins and blocking the nitrocellulose with 5% dry milk in PBS, the nitrocellulose was cut into strips (each strip containing approximately 5 µg of recombinant protein) which then were used to determine the presence of anti-HCV antibody in test sera (or other samples). Reaction mixtures consisted of a nitrocellulose strip incubated with an appropriate amount of test sample in 2.0 ml of buffer (20 mM Tris, 1 mM EDTA, 0.2 M NaCl, 0.3% Triton X-100® and 2 mg/ml bovine serum albumin (BSA), pH 7.5, 5% *E. coli* lysate and 3% CKS lysate overnight at 4°C. The strips were washed with buffered detergent (10 mM phosphate buffered saline (PBS) pH 7.5, containing 0.1% SDS and 0.5% Triton X-100®), followed by addition of goat anti-mouse IgG antibody conjugated to HRPO. The strips were incubated for one to two hours at room temperature, followed by washing with buffered detergent. Finally, antibody bound to the protein was

visualized by addition of freshly prepared HRP color reagent (Biorad Laboratories, Richmond, CA) (120 mg dissolved in 40 ml ice-cold methanol, then diluted into 200 ml Tris buffered saline, pH 7.8, containing 120 µl of 30% hydrogen peroxide). This assay demonstrated the presence of antibody to the respective proteins with which the mice had been immunized.

EXAMPLE 2

Cell Fusion

Upon demonstration of specific anti-HCV antibody present at reasonable titers in sera of immunized mice, the mice were allowed to rest for at least eight weeks prior to a pre-fusion boost of antigen. The pre-fusion antigen boost then was performed by intravenous (IV) tail vein injection of approximately 40 µg of respective purified recombinant HCV protein. Three days later the mice were sacrificed, and their spleens which contained anti-HCV antibody-producing cells were disrupted into single cells. These single cell suspensions were treated with 0.83% NH₄Cl to remove red blood cells, and then these suspensions were mixed with SP2/0 cells at a 10:1 (SP2/0:spleen cells) ratio. The mixed cells were centrifuged, washed once with serum-free medium, and again centrifuged. The fusogen polyethylene glycol (PEG) was used to form hybrids of the immune donor spleen cells with the myeloma cell line SP2/0 (HPRT neg.) Kohler and Milstein, Nature 256:494 (1975), and reviewed in J.G.R. Hurrel, ed., Monoclonal Hybridoma Antibodies: Techniques and Applications, CRC Press, Inc., Boca Raton, FL (1982). Briefly, fusion of the spleen and SP2/0 cells was accomplished by exposing the pellet to 40% PEG (ATTC, MW 1300-1600) in serum-free Isco's Modified Dulbecco's Medium (IMDM) for two minutes. The PEG and cell suspension was diluted slowly by the addition of 20 ml of serum-free IMDM over a period of five minutes, followed by collection of the cells by centrifugation. The supernatant was decanted and replaced with 30 ml IMDI containing 20% fetal bovine serum (FBS) (Hyclone Laboratoreis, Logan, Utah) with HAT (hypoxanthene, aminopterin and thymidine) media in order to select for hybridomas. Spleen cells from one non-immune BALB/c mouse also were added as a feeder layer. The cells were plated at 0.1 ml/well in three 96-well tissue culture plates. An additional 0.1 ml of HAT media was added to each well three days later. At weekly intervals thereafter, one-half the media was replaced with IMDM containing 20% FBS with HT (hypoxanthene and thymidine), and hybrids were allowed to grow for an additional seven to fourteen days.

It was found that some of the hybrids were composed of spleen cells making antibody to HCV fused with SP2/0 cells. Briefly, the fusogen promoted fusion of spleen cell and SP2/0 cell membranes, which formed a heterokaryon containing nuclei of both cells. Eventually, the dissimilar nuclei fuse produced a single nucleus capable of 5 synchronous mitosis. As the fused cells divided, the hybrid stabilized by losing chromosomes of each nucleus. The fused cells were plated into multiple 96-well plates at 10^5 to 10^6 cells per well. The hybrid cells formed from SP2/0:spleen cell fusions were selectively propagated by culturing in HAT medium. All unused SP2/0 or SP2/0:SP2/0 fused cells were prevented from growing by aminopterin, and unfused 10 spleen cells or spleen:spleen fused cells died off in culture. Only SP2/0:spleen cell hybrids grew in the HAT selection medium.

EXAMPLE 3

Screening and Cloning of Monoclonal Antibodies

15 After 10 to 14 days, culture fluids from wells containing hybridoma cell growth were screened for the presence of a monospecific antibody as follows. Each of the hybridoma culture fluids was tested on a plate coated with the immunogen as well as on a plate coated with CKS protein (fusion partner used for HCV proteins) by the EIA procedure described in Example 1. Hybridoma culture fluids reacting specifically to the 20 immunogen, i. e., HCV protein, and not the CKS fusion partner were selected for further analysis by Western blot analysis. EIA-positive hybridoma culture fluids were tested for their reactivity to the respective HCV proteins as well as CKS by Western blot analysis as described in Example 1. Hybrid samples reacting specifically with the HCV protein but not with the CKS protein by both EIA and Western blot were identified, and 25 selected for cloning by the limiting dilution method, using the guidelines outlined by J. W. Goding, Monoclonal Antibodies: Principles and Practices, Academic Press, New York, (1983). Culture supernatant of cloned samples were tested again by EIA with the immunogen and the CKS protein as described above in Example 1, for the confirmation of monospecific reactivity to HCV protein sequence. Clones with strongest reactivity to the 30 protein of choice were selected for expansion and further analysis.

EXAMPLE 4

Amplification of Antibody Yields by Ascites Method

In order to obtain greater amounts of monoclonal antibodies, 10 to 20 million 35 cloned cells of the desired hybridoma cell line were inoculated into a BALB/c mouse previously treated i.p. with 0.5 ml pristane (2,6,10,14-tetramethylpentadecane) by

the method outlined in J. G. R. Hurrell, ed., Monoclonal Hybridoma Antibodies: Techniques and Application, CRC Press, Boca Raton, FL (1982). Pristane treatment enhanced growth of mouse myeloma hybrids within the peritoneum of the mouse, and the ascites fluids which formed were rich in the monoclonal antibody secreted by the hybrid cells. After formation of adequate ascites fluid (approximately seven days), the mice were sacrificed and the ascites were withdrawn from the peritoneum, clarified by centrifugation and stored at -20°C. Monoclonal antibodies from ascites fluid were purified using protein-A sepharose (according to J.G.R. Hurrell et al, *supra*). All characterization procedures described herein were performed with either culture supernatants, ascites fluids or protein-A purified IgG.

EXAMPLE 5

Characterization of Monoclonal Antibodies

EIA

15 Enzyme-linked immunoassay as described in Example 1 was used to determine the specificity of each of the monoclonal antibodies. Briefly, clarified ascites fluids or protein-A purified IgG were reacted in serial dilutions in microtiter plates coated with either a) the immunogen (i.e., pHCV23, or pHCV39 or pHCV34), b) CKS protein (fusion partners used for cloning and expression of all three immunogens), and c) respective proteins (i.e. C-100, 33C or CORE) expressed in E. coli under the control of bacteriophage λpL promoter (i.e. HCV proteins expressed without CKS fusion partner). Specificity of each of the monoclonal antibodies for the respective protein was confirmed by specific activity of the monoclonal antibody to the immunogen as well as the HCV protein expressed in E. coli λpL but not with CKS protein. TABLE 1 illustrates the 20 representative data for monoclonal antibodies of the invention to C-100, 33C and CORE proteins of HCV.

25

Western blot analysis

The general protocol for Western blot analysis is as described in Example 1, except that CKS lysate was excluded from the dilution buffer. Briefly, approximately 30 300 µg of either a) the immunogen (i.e. either pHCV 23, pHCV 29 or pHCV 34), or b) CKS protein (fusion partner for immunogen) or c) the respective proteins expressed in E. coli under the control of λpL promoter, were electrophoresed and transferred to the nitrocellulose. After blocking free sites on nitrocellulose, 2 mm side strips were cut. 35 Each of the monoclonal antibodies was tested for reactivity against all three antigens (i.e., the immunogens, CKS and the respective HCV proteins expressed in E. coli λpL).

Specificity of each of the monoclonal antibodies was confirmed as described for EIA analysis. Representative data are illustrated in FIGS. 2A to 2F. Referring to FIGS. 2A to 2F, the photographs show mono-specific binding of each monoclonal antibody of the invention to its specific protein.

5

Isotype

The isotype of each of the monoclonal antibodies was determined by using an isotyping kit (Amersham, Arlington Heights, IL) and following the instructions included with it. Briefly, the tissue culture supernatant of each monoclonal antibody and appropriate controls were reacted at a 1:5 dilution with strips coated with specific anti-isotype antibody, provided in the kit described above. Assay protocol was followed exactly according to the manufacturer's instructions. The isotype of each monoclonal antibody of the invention is provided in TABLE 1.

10

Competition With Immune Human Sera

In order to establish whether each of the monoclonal antibodies recognized an epitope that is immunogenic in humans, a competition assay was performed as follows. Each of the monoclonal antibodies was tested in an assay where the monoclonal antibody competed with a human sera seropositive for antibody to C-100, 33C and CORE for the binding to the respective antigen. Briefly, a human serum from an individual infected with NANBH and strongly seropositive for antibodies to C-100, 33C and CORE proteins of HCV was included in the reaction mixture with each of the monoclonal antibodies at a final concentration of 10%. Microtiter EIA was carried out as described in Example 1. A greater than 50% inhibition in the binding of the monoclonal antibody to the respective protein by the immune human sera was considered as competitive (data presented in TABLE 1).

EXAMPLE 6

Epitope Mapping

15

Monoclonal antibodies to HCV proteins C-100, 33C and CORE were mapped to the specific region of the protein by (a) Western blot reactivity of each of the monoclonal antibodies with subfragments of the respective HCV proteins and (b) reactivity with several synthetic peptides selected for respective protein sequences, by microtiterEIA. In addition to these two methods, monoclonal antibodies to C-100 also were mapped by PEPSCAN analysis, in order to further define the epitope recognized by these antibodies.

20

Specific additional details for mapping will be detailed where applicable for an individual monoclonal antibody.

Reactivity of Monoclonals to Various Subfragments of Recombinant HCV proteins

5 Briefly, several individual oligonucleotides representing a.a. 1676-1931 of HCV genome were ligated and cloned as three separate EcoRI-BamHI subfragments into the CKS fusion vector pJ0200. These three subfragments were designated as CKS-B (a.a. 1676-1790), CKS-C (a.a. 1789-1863) and CKS-D (a.a. 1861-1931) as described in FIG.1.. The Western blots of selected monoclonal antibodies of the invention are shown in 10 FIGS. 2-2F. The detailed methods for cloning and expression of the CKS-fusion proteins are as disclosed in U. S. Patent Application Serial No. 07/572,822, which enjoys common ownership and is incorporated herein by reference. Cell lysates of these clones 15 were used as antigens on Western blot analysis for preliminary epitope mapping of anti-C-100 monoclonal antibodies. Similarly, two subfragments from HCV 22C regions designated as 33C A-CKS (a.a. 1192-1331) and 33CB-CKS (a.a. 1330-1457) also were cloned and expressed in E. coli as described above for FIG. 1. These lysates were used as antigens for epitope mapping of anti-33C monoclonal antibodies.

20 Western blot analysis of each of the monoclonal antibodies with appropriate sets of subfragments of control proteins (full length proteins as well as CKS fusion partner) was carried out as described in Example 1, except that CKS lysate was excluded from the dilution buffer. Data for epitope mapping with these recombinant 25 subfragments are illustrated in FIGS. 2A to 2F. Monoclonal antibody H28C110 showed reactivity with C-100B, pHCV23 and C-100 (λ pL construct) but did not react with C-100C, C-100D or CKS protein, which indicated that H28C110 specifically recognizes or binds an epitope between a.a. 1676-1790 of the HCV genome. Similarly, monoclonal antibody H4C20 recognizes (specifically binds) an epitope between a.a. 1861-1931. Based on these data, monoclonal antibody H11C130 recognizes an epitope between a.a. 1192-1331, and monoclonal antibody H1C46 recognizes (specifically binds) an epitope 30 between a.a. 1330-1457.

Reactivity with Synthetic Peptides

Several amino acid sequences were selected from different regions of HCV proteins C-100, 33C and CORE. A list of the peptides used for the epitope mapping of 35 these monoclonal antibodies is listed below in TABLE 2.

TABLE 2
Epitope Mapping With Synthetic Peptides

	REGION OF HCV GENOME	MONOCLONAL TESTED	PEPTIDE a.a.	REACTIVITY OF EACH WITH PEPTIDE
5	CORE	H81C17	sp 1-75	None
		H35C54	sp 35-75	None
10	33C	H1C46	sp 1192-1240	None
			sp 1223-1240	None
		H11C130	sp 1357-1407	None
15	C-100	H28C110	sp 1418-1457	None
			PEPSCAN analysis (a.a. 1694-1735)	sp 1694-1750 and
			sp 1694-1750	sp 1684-1735;
			sp 1684-1735	a.a. 1702-1709;
			sp 1696-1708	(PEPSCAN)
20		H4C20	sp 1866-1930	sp 1899-1930
			sp 1899-1930	sp 1899-1930

Each of these peptides were assembled on a resin support by a stepwise solid phase synthesis, starting with the carboxy terminal residue. A procedure was employed similar to that described in E. Gross and T. Heinehofer, eds., Barany and Merrifield, The Peptides 2:1284, Academic Press, New York, New York (1980), using a reaction vessel of an Applied Biosystems Synthesizer Model 430A. After cleavage of the peptide from the resin, the peptide was washed with diethyl ether and extracted in 40% acetic acid solution. Crude peptide obtained after lyophilization of the aqueous solution was employed as the antigen target for epitope mapping experiments. Briefly, each of the peptides tested was coated on microtiter wells at a concentration of 10 μ g/ml in bicarbonate buffer at pH 9.5. EIA was performed in the manner described in Example 1. Monoclonal antibody showing reactivity four times the negative control was considered positive.

In addition, monoclonal antibodies to HCV C-100 were also mapped with PEPSCAN analysis. Synthetic peptides were synthesized on polypropylene pins following the manufacturer's instructions (Cambridge Research Bioscience, Valley Stream, New York). An EIA was performed with each of the monoclonal antibodies to HCV C-100 by the procedure outlined by the manufacturer with 67 overlapping hexamer peptides

encompassing a.a. 1684-1750 of the HCV genome. Representative data are illustrated in FIG. 4. Monoclonal antibody H28C110 specifically reacted with peptide sequence Tyr-Arg-Glu-Phe-Asp-Glu-Met-Glu (a.a. 1702-1709 of the HCV genome). Monoclonal antibody H4C20 did not show reactivity in PEPSCAN but did react in EIA with a larger peptide 1899-1930 a.a. This could be due to the fact that H4C20 requires a longer peptide to recognize a linear epitope.

EXAMPLE 7

EIA for the Detection of HCV Proteins in Biological Samples

For the purposes of simplification, detailed methods are described for the CORE proteins of HCV. Methods described for the preparation of rabbit polyclonal antibodies to HCV proteins 33C and C-100 were identical to that detailed hereinbelow for HCV CORE protein.

Preparation of Rabbit Polyclonal Antibodies Against HCV CORE Protein

Young rabbits (3-4 months old and weighing 2-3 kg) were obtained from Hazelton Labs, Denver, PA. Primary immunization was carried out with 100-150 µg of highly purified HCV CORE protein (expressed in *E. coli* under λpL promoter) in Freund's complete adjuvant by intra-muscular (I.M.) injection at four different sites.

Subsequently, two immunizations were carried out at two week intervals in similar fashion in Freund's incomplete adjuvant. Immune response of the rabbits was monitored by EIA and Western blot analysis as described in Example 1. Rabbits were bled when acceptable immune response to the protein was achieved. IgG from the immune rabbit sera was purified by Protein-A sepharose affinity chromatography, by methods known to those in the art.

Bead Coating

In the presently most preferred assay format, rabbit IgG prepared as hereinabove was coated on polystyrene beads as the solid support for capture of CORE antigens in the test sample. The polystyrene beads were washed with distilled water and incubated at 40°C for two (2) hours with 5-10 µg/ml of purified HCV CORE rabbit IgG in a buffer solution (0.1M Tris, 0.5M NaCl, 0.0022% Triton X-100®, pH 8.5). The beads were washed once with PBS and then soaked in 0.1% Triton X-100® in PBS for approximately one (1) hour at 40°C. After washing twice with PBS, the beads were overcoated with 3% bovine serum albumin (BSA) in PBS for approximately one (1) hour at 40°C. Finally, the beads were overcoated with a 5% sucrose solution in PBS and

dried under nitrogen. Anti-HCV human polyclonal IgG, purified from sera of individuals seropositive for HCV antibodies to C-100, 33C and CORE also was coated in similar fashion.

5 EIA

Several monoclonal antibodies specific for either HCV C-100, HCV 33C or HCV CORE were screened for use as the probe for detection of HCV proteins in a test sample by EIA. Briefly, each of the monoclonal antibodies was incubated with the respective antigen in the presence of polystyrene beads coated with anti-HCV rabbit polyclonal IgG.

10 The detailed protocol for EIA was similar to that described hereinbelow. Data illustrating the reactivity of the monoclonal antibodies which showed the best reactivity as a probe for the antigen detection assay are shown in TABLE 3.

TABLE 3

15

Monoclonal Antibody	Antigen	Absorbance at A492 at antigen conc.	
		NEG. CONTROL 0 ng/ml	POS. CONTROL 100 ng/ml
20 H81C17	HCV CORE	0.246	>6.62
H11C130	HCV 33C	0.194	0.832
H28C110	HCV C-100	0.623	2.53

25 In the EIA procedure, 200 µl of specimen which was suspected of containing antigen to HCV CORE protein was incubated in a reaction tray with 50 µl of monoclonal antibody H81C17 (at a final protein concentration of 5-10 µg/ml diluted in a buffer containing 20 mM Tris, 0.1 mM NaCl, 1 mM EDTA, 3.0% BSA, 0.3% Tween-20® and 10% FBS at pH 7.5), and a bead coated with HCV rabbit IgG (prepared as described hereinabove). After overnight incubation at ambient room temperature, the beads were washed with distilled water and 200 µl of appropriately diluted Horseradish peroxidase labeled goat anti-mouse IgG (H + L) (Jackson Immunoresearch, West Grove, PA) was added. Incubation with the labeled probe was carried out at about 40°C for approximately two (2) hours. Beads were washed and transferred to reaction tubes containing 300 µl of O-phenylenediamine:2HCl (OPD) color reagent. The reaction was carried out at ambient room temperature in the dark for 30 minutes, and then it was stopped by the addition of 1 ml of 1N H₂SO₄. Absorbance was recorded at 492/600 nm. A negative control which was previously screened and confirmed to be negative for

NANBH infection was included in the experiment. The positive control consisted of a solution of recombinant HCV CORE protein (pHCV 34) in the buffer solution described hereinabove. Triplicates of both positive and negative control were included with each set of experiments.

5

In order to determine the efficiency of the antigen capture assay for the detection of HCV CORE protein in a sample, various concentrations of recombinant HCV CORE protein (expressed in *E. coli* under λ pL promoter), ranging from 100 ng protein/ml to 100 pg protein/ml were diluted in the buffer mentioned above. The EIA procedure was performed with each of diluted the panel members following the procedure described herein. For the purpose of comparison, each of the panel members was tested with (a) anti-HCV rabbit polyclonal antibody on the solid phase and (b) anti-HCV human polyclonal antibody on the solid phase. As illustrated in FIG. 4 and TABLE 4, the antigen capture assay of the present invention detected as low as 200 pg HCV CORE protein in 200 μ l of the test sample. Although there was no significant difference in the sensitivity of detection with either rabbit or human capture antibody, anti-HCV rabbit antibody showed overall superior performance in the assay and was chosen as the preferred polyclonal antibody for the antigen capture.

20

TABLE 4
Detection of HCV CORE Protein by Monoclonal Antibody H81C17

	CONC. OF ANTIGEN	REACTIVITY IN EIA	
		A492 Rabbit Bead	A492 Human Bead
	pL CORE* ng/ml		
25	100	6.39	4.45
	50	4.06	2.19
30	25	2.19	1.25
	12.5	1.02	0.887
	6.25	0.339	0.382
	3.125	0.230	0.121
	1.56	0.084	0.043
	0.78	0.050	0.032
35	0.39	0.038	0.014
	0.2	0.028	0.015
	0.1	0.020	0.014
	0	0.014	0.014

40 * Recombinant HCV CORE expressed in *E. coli* under λ pL promoter system used as the antigen. Concentrations represent ng of protein/ml based on A280 values. The assay

system has 200 µl/well for testing, so the actual amount of antigen per well is one-fifth of the value given here.

EXAMPLE 8

5 Testing of Negative Serum and Plasma Samples

Sixty sera and sixty plasma samples negative for NANBH were tested by the EIA procedure detailed in Example 7. The results of these assays are presented in FIG. 5. It can be seen by the data that the majority of negative plasma and sera fell within a tight range of O.D. (Optical Density) values close to the negative control value. One plasma 10 sample with an O.D. of 0.1725 was found to be repeat reactive. Confirmation of the presence of antigen could not be performed on this sample due to the insufficient volume of the sample..

EXAMPLE 9

15 Production and Use of Cell Lines 1 3-975-157, 14-153-234 and 14-1350-210

A. Production of recombinant HCV antigens and immunogens

Synthetic peptides corresponding to regions within the putative CORE domains of the HCV genome were made by automated peptide synthesizer. The following peptides 20 were constructed utilizing methods known in the art:

CORE 1-75

35-75

35-61

These peptides are described in pending United States patent application Serial 25 No. 07/610,180, entitled HEPATITIS C ASSAY, which enjoys common ownership and is incorporated herein by reference.

Recombinant antigens were made as fusion proteins with CMP-XDO synthetase in E. coli according to methods known in the art, or as non fused protein with λPL promoter 30 system. The following proteins were cloned and purified:

λPL-CORE (1-150)

CKS-CORE (1-150)

CKS-33c (1191-1457)

λPL-33c-CORE (fusion of 1191-1457 and 1-150)

35 CKS-BCD (156-1930)

CKS-E (1931-2189) (NS4/NS5 reaction)

CKS-B (1676-1790)

See FIG. 1B for map of HCV genome and approximate locations of HCV regions.

Recombinant protein C-100 (1569-1930) was obtained from Chiron as a fusion protein with superoxide dismutase (SOD). All recombinant proteins were greater than 90% pure by SDS-PAGE.

B. Immunization of Mice

BALB/c mice (Charles River Laboratories, Charles River, NY), 6-8 weeks old, were initially immunized subcutaneously and intraperitoneally with 50 µg of λPL-CORE in 100 µl of Freund's complete adjuvant (Difco, Detroit, MI). On day 15, 50 µg of the immunogen was diluted into 100 µl of phosphate buffered saline (PBS), pH 7.2, and injected intravenously into the tail vein (J. Goding, Monoclonal Antibodies: Principles and Practice [New York; Academic Press, 1986]). Sera titers were not evaluated.

C. Fusion

On day 18, mice were sacrificed and splenocytes were fused in a 1:1 ratio with the SP2/0 myeloma line according to known conventional methods (G. Kohler and C. Milstein, Nature (1975) 256:495-497; J. Goding, supra). The cell fusion pellet was dispersed with 1 ml 50% polyethylene glycol (PEG) (American Type Culture Collection, MW 1450) and centrifuged in Iscove's Modified Dulbecco's Medium (IMDM) (Gibco, Grand Island, NY). The cells were resuspended in HAT (hypoxanthine-aminopterin-thymidine)-selective IMDM with 10% fetal bovine serum (FBS) (Hyclone Laboratories, Logan, UT) and plated at 3×10^5 cells per 96-well tissue culture plates. Growth promoters included in the HAT media were 0.5% STM (RIBI Immunochem Research, Inc., Hamilton, MT) and 1% Origen Hybridoma Cloning Factor (Igen, Rockville, MD). Growth medium was replaced in culture wells post-fusion on day 5 and 7 using HT (hypoxanthine-thymidine) supplemented IMDM with 10% FBS.

D. Enzyme Immunoassay (EIA)

Culture supernates were EIA screened 10 days post-fusion against the immunizing antigen to detect hybrids secreting HCV specific antibody and a non-specific protein to eliminate any false positives (Langone & Van Yunakis, eds., Methods in Enzymology, 92:168-174, Academic Press [1983]). Polystyrene 96-well microtiter plates were coated overnight at room temperature with 50 µl per well of a 1 µg/ml HCV antigen solution in PBS. Any remaining binding sites on the polystyrene wells were

blocked with 3% bovine serum albumin (BSA) (Intergen, Purchase, NY) in PBS for 30 minutes at room temperature. Plates were washed three times with distilled water. Fifty microliters of hybridoma tissue culture supernatants were incubated for 1 hour at room temperature in the wells, and the wells were washed three times with distilled water. Antibody binding to antigen was detected using goat anti-mouse IgG+M-horseradish peroxidase (HRPO) (Kirkegaard-Perry Laboratories [KPL], Gaithersburg, MD) diluted at a concentration of 1:1000 in the block solution and incubated 30 minutes at room temperature. The plates were washed with distilled water and o-phenylenediamine substrate (OPD; Abbott Laboratories, Abbott Park, IL) was used as the chromogen. Plates were read at 492 nm. Hybrid cultures were regarded as potential HCV antibody-positive when the optical density (OD) was 3 times the negative control (NC) and significant preferential to the HCV antigen plate was observed compared to antibody binding of the irrelevant antigen coated plate, i.e.: >0.2 OD difference and <0.2 OD signal on the latter.

E. Western Blot

Hybrid antibody specificity was confirmed with Western blot analysis (Towbin & Gordon, J. Immunol. Methods, 72:313-340 [1984]). HCV recombinant proteins and irrelevant proteins were electrophoresed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and then transferred to nitrocellulose, according to the manufacturer's instructions (Schleicher & Schuell, Keene, NH; Bio-Rad, Richmond, CA). The nitrocellulose strips were blocked with 1% bovine hemoglobin (Sigma Chemical Co., St. Louis, MO) and 0.5% Tween-20 (Fisher Scientific, Pittsburgh, PA) in PBS for 30 minutes at room temperature, then the strips were incubated with hybrid tissue culture supernatant. The strips were then washed in PBS and goat anti-mouse IgG+M-HRPO (KPL) added for 30 minutes. Antibody binding to the HCV antigen was visualized with 4-chloro-1-naphthol (Sigma) as the chromogenic substrate. Hybrid cultures were cloned and placed in cryostorage if HCV antibody specificity was demonstrated.

E. Establishment of Clones

HCV specific hybrids were cloned by limiting dilution (Goding, Monoclonal Antibodies: Principles and Practices, 2nd ed, Academic Press, New York [1986]). Modifications included plating of the cultures in \log_{10} dilution series and selecting positive clones for expansion from plates which exhibit <20% growth per 96 well tissue culture plate. Culture supernates were tested after 10 days using the EIA and

Western blot procedures described above. The selected clones were expanded for further evaluation and cryostoraged in 80% IMDM with 10% FBS and 10% DMSO (Sigma).

5 G. Monoclonal Antibody Isotype

Monoclonal antibody isotype was determined with the SBA Clonotyping System-III kit (Southern Biotechnology Associates, Inc., Birmingham, AL) with modifications. EIA 96-well microtiter plates were coated overnight at room temperature with 100 µl/well of a 1:1000 dilution of goat anti-mouse IgG+M (H+L) (KPL). Plates were 10 blocked for 30 minutes with 3% BSA in PBS and washed with water. Culture samples were added to the wells, incubated for 1 hour, and washed with water. The kit's goat 15 anti-mouse subtype specific conjugates were added for a 30 minute incubation period. Following a water wash, color was identified with OPD substrate. The goat anti-mouse isotype specific conjugate that bound to the mouse immunoglobulin and displayed a >0.1 OD at 492 nm signaled the subtype.

H. Monoclonal Antibody Production

Clones selected for further evaluation were scaled up in tissue culture T-flasks and 10⁶ cells were injected into the peritoneal cavity of pre-pristaned BALB/c mice 20 (Charles River Biotechnical Services, Inc., Wilmington, MA) (see Hurrell, supra). The resulting ascites fluid was harvested 7-10 days after injection, centrifuged, and stored at -20°C. The IgG antibody was affinity purified on Protein A (Pharmacia-LKB Biotechnologies, Piscataway, NJ) utilizing the automated OROS purification system Model 100 (see Goding, supra, for basic principles). The IgM antibodies were purified 25 by molecular sizing on a S-300 column (Pharmacia-LKB).

All the following characterization information was performed with purified monoclonal antibody.

30 I. Isoelectric Focusing (IEF)

A cell line quality control to ensure consistency of frozen lots included measuring the antibody pI point on an IEF gel apparatus (Bio-Rad) which separates proteins based on net charge. Briefly, a bis-acrylamide-riboflavin solution was applied to an acrylamide gel, exposed to fluorescent lighting for 1 hour, then stored overnight at 4°C. 35 A 1 µg sample of monoclonal antibody and standards were laid upon the gel and electrophoresed over a 1-2 hour period. Following a series of fixatives and washes, the

gel was silver stained (Bio-Rad). The pI value of the monoclonal antibody was calculated by migratory distance through the gel and was directly compared to the protein standards' migratory distance of known pI values. The distinctive finger print banding pattern reflected the pI microheterogeneity between independently produced lots of antibody (Hamilton, R.G., Reimer, C.B., Rodkey, L.S. (1987) Quality control of murine monoclonal antibodies using isoelectric focusing affinity immunoblot analysis. Hybridoma 6:205-217).

J. EIA and Western Blot Specificity of Monoclonal Antibodies

All monoclonal antibodies noted herein were screened on an assortment of available recombinant HCV antigens as disclosed in U. S. Patent Application Serial No. 07/572,822 entitled HEPATITIS C ASSAY UTILIZING RECOMBINANT PROTEINS, which enjoys common ownership and is incorporated herein by reference. The procedures are outlined above. The multiple antigen screening technique confirmed the HCV specificity and to exclude the HCV non-specific CKS, λ PL, or linker-arm reactivity of the monoclonal antibodies.

K. EIA Epitope Competition Studies

To investigate specificity and antigen binding distinctions, epitope grouping experiments were performed utilizing biotin labeled and unlabeled monoclonal antibodies (Langone & Van Vunakis, Methods in Enzymology, 92:242-253, Academic Press [1983]). Briefly, the antibodies were labeled with NHS-LC-biotin (Pierce Chemical Co., Rockford, IL) according to the manufacturer's instructions. Microtiter wells were coated with the immunogen as previously described. First, \log_2 dilutions of the unlabeled antibody were pre-incubated in the wells for 15 minutes, followed by the addition of a fixed amount of biotinylated antibody (the dilution in a direct EIA of the biotinylated antibody alone which gave a value of 50% of the maximum absorbance value) and incubated for 20 minutes. Plates were washed three times with water. Diluted streptavidin-HRPO (Zymed, South San Francisco, CA) was added to the wells and incubated for 30 minutes. The plates were washed again and OPD color developed as previously described. The absorbance was read at 492 nm. Antibodies of the same or related epitope had signal blocked or inhibited by >50%. No inhibition was observed with antibodies of distinct specificity. This was performed reciprocally for antibodies produced within HCV core region.

L. RIA Reciprocal Competition

Beads coated with the appropriate antigen or peptide were incubated with 100 µl of unlabeled monoclonal antibody diluted into recalcified negative human plasma (NHP, testing negative for anti-HCV, anti-HIV and HBsAg) at monoclonal antibody

5 concentrations of 1-20 µg/ml. 100 µl of radiolabeled antibody at 1 to 4 µCi/ml diluted into HTLV I kit specimen diluent (containing detergent, animal sera, buffer) was incubated with the bead for 2 hours at 45°C or 18-20 hours at 20-25°C. Beads were washed and counted for radioactivity.

10 M. EIA Reactivity with Synthetic Peptides

Beads coated with 3 mg/ml were incubated with 100 ml of monoclonal antibody at concentrations of 0.02 -1 mg/ml for 1 hour. The beads were washed and goat anti-mouse IgG+M-HRPO (KPL) was added for 30 minutes. The beads were washed again and OPD (Abbott Labs) was used as the chromagen for O.D. readings at 492 nm.

15

N. HCV Antigen Assays

Beads coated with one or a cocktail of anti-HCV monoclonal antibody were incubated with 200 µl of specimen for 2 hours at 40-45°C or 18-20 hours at 20-25°C. Beads were washed with distilled water and then incubated with 200 µl of 20 radiolabeled anti-HCV monoclonal antibody (one or more) for 2 hours at 45°C. Beads were washed and counted in a gamma counter.

O. Characterization of Monoclonal Antibody

Monoclonal antibody against the HCV CORE domain (1-75) fell into two distinct 25 groups based on reciprocal competitive studies. All groups reacted with CKS-CORE (1-150) and IPL-core, IPL-33c-core and synthetic peptides corresponding to (1-75). Group 1 monoclonal antibody 13-975-157 strongly reacted with peptide (1-75), somewhat to (35-75), and not to (35-61). Group 2 monoclonal antibody 14-153-334 clearly reacted to peptides (1-75) and (35-75), but not to (35-61). Unlike the 30 other groups, monoclonal antibody 14-1350-210, reacted strongly to all 3 of the HCV core synthetic peptides. These data are shown in Tables 5 and 6. Referring to FIGS. 6 to 17, the reactivities summarized below in Table 5 are shown in lanes 4, 5 and 6. Lanes 1 to 3 contains monoclonal antibodies against HCV 33C protein (6-296-534 in lane 1; 6-914-518 in lane 2 and 6-1070-110 in lane 3); lanes 4-6 contain monoclonal 35 antibodies against HCV CORE (13-975-157 in lane 4, 14-153-234 in lane 5 and 14-1350-210 in lane 6); lanes 7 and 8 contain monoclonal antibodies against the putative

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TABLE 7
Competitive One step anti-CORE assay

		Label: 14-153-234					Label: 14-1350-210				
	Specimen	CPM	Avg	S/N	% Inhibition	Result	CPM	Avg	S/N	% Inhibition	Result
5	NC	48272	48542				36866	34521			
		47010					35655				
		50345					31041				
10	Elevated ALT										
	27	4845	5130	0.11	89.4	+	440	405	0.01	98.8	+
		5414					370				
	238-NC	40035	41964	0.86	13.6	-	28481	28294	0.82	18.0	-
		43892					28106				
15	135	620	558	0.01	98.9	+	114	117	0.00	99.7	+
		496					119				
	163	14540	15471	0.32	68.1	+	100	102	0.00	99.6	+
		16402					104				
	173	5344	5531	0.11	88.6	+	114	126	0.00	99.6	+
20		5717					137				
	220	1049	1012	0.02	97.6	+	124	130	0.00	99.6	+
		974					136				
	252	231	212	0.00	99.6	+	138	135	0.00	99.6	+
		192					132				
25	R13203	56075	55778	1.15	-14.9	-	3029	3479	0.10	89.9	+
		55480					3929				
	SAC190	472	417	0.01	99.1	+	102	194	0.01	99.4	+
		362					284				
	7088396	41692	41963	0.86	13.6	-	102	122	0.00	99.6	+
30		42233					142				
	EP10968	39418	40470	0.83	16.6	-	94	95	0.00	99.7	+
		41522					95				
	14-153	5580	5491	0.11	88.7	+	16216	14178	0.41	58.9	+
		5402					12140				
35	283-NC	45170	44219	0.91	8.9	-	24748	23908	0.69	30.7	
		43268					23067				

40

EXAMPLE 11Anti-HCV CORE Two-Step Blocking Assay

A two step blocking assay performed according to a competitive one-step assay described herein for "RIA Reciprocal Competition" for detection of anti-HCV CORE, using labeled group 3 monoclonal antibody (14-1350-210), is shown in Table 8. Sixteen specimens which were repeat reactive for anti-C100 (using the Ortho 1.0 gen kit, available from Ortho Diagnostics, Raritan, New Jersey) were tested. Six had been

shown to be false positive by an anti-C100 confirmatory assay and 10 were confirmed as positive. None of the six false positives gave greater than 20% inhibition in the assay compared to 48-99% inhibition observed for true anti-C100 positives in this anti-CORE assay.

5

Table 8

Two Step Blocking Anti-HCV-CORE Assay

Bead: 1.0 µg λPL-CORE; Label: 14-1350 in HTLV I diluent

10

	Specimen	CPM	AVG	S/N	% Inhibition	Result
	NC	23376	25969			
		26257				
		28275				
15	Sacramento Negative 1.0 Gen.					
	183	31956	1.23	-23.1		
	184	26525	1.02	-2.1		
	185	24714	0.95	4.8		
	192	20664	0.80	20.4		
20	193	20934	0.81	19.4		
	194	23321	0.90	10.2		
	Sacramento Positive 1.0 Gen.					
	188	13569	0.52	47.7	+	
	189	2243	0.09	91.4	+	
25	191	2361	0.09	90.9	+	
	195	284	0.01	98.9	+	
	196	170	0.01	99.3	+	
	197	487	0.02	98.1	+	
30	206	3639	0.14	86.0	+	
	207	2713	0.10	89.6	+	
	208	145	0.01	99.4	+	
	214	10398	0.40	60.0	+	

30

35

EXAMPLE 12HCV CORE Ag Assay

Results from one type of CORE Ag assay is shown in Table 9. This assay was performed as a 2 step assay according to the procedure outlined hereinabove in "HCV Antigen Assay" and as follows: The first incubation with specimen (200 µl) was 18 hours at room temperature, followed by 2 hr. incubation at 45°C with radiolabeled group 2 monoclonal antibody 14-153. Since the bead was coated with a group 3 monoclonal antibody 14-1350 which does not compete with group 2, a one step sandwich assay was also possible.

Specimens giving S/N values greater than 3.0 were considered reactive for CORE Ag. Sensitivity of the assay with recombinant λPL-CORE was about 100 ng/ml. Two of

13 specimens from patients with elevated ALT and anti-C-100 reactivity were considered reactive for CORE antigen.

Table 9
HCV CORE Ag Assay
(14-1350 bead/14-153 label)

	Specimen	CPM	Average	S/N	Result
5	NC	114	128		
10		137			
		133			
15	λPL-CORE	110 µg/ml	1846	1822	14.23 +
			1798		
	"	11 µg/ml	1488	1447	11.30 +
	"		1405		
20	"	1.1 µg/ml	1220	1246	9.73 +
	"		1271		
	"	110 ng/ml	514	472	3.69 +
			430		
25	"	11 ng/ml	182	175	1.36 -
			167		
	Elevated ALT	10	206	212	1.65 -
			217		
30	"	27	160	174	1.36 -
			187		
	"	54	710	628	4.91 +
			546		
35	"	77	454	458	3.57 +
			461		
40	"	135	160	189	1.48 -
			218		
	"	163	323	328	2.56 -
			332		
45	"	173	346	342	2.67 -
			337		
	"	220	274	275	2.14 -
			275		
	"	238	157	171	1.34 -
			185		
	"	252	227	216	1.68 -
			204		
	"	283	194	175	1.36 -
			155		
	"	28	258	226	1.77 -
			194		
	"	290	162	175	1.36 -
			187		

EXAMPLE 13HCV Antibody Test Employing Cocktails of Monoclonal Antibodies

Also performed was a similar CORE Antigen format using a cocktail of anti-CORE monoclonal antibody on the solid phase and in label (14-1350-210, 14-153-234, and 5 14-726). In twenty five anti-C-100 repeat reactive specimens, one specimen (SAC 161) was significantly elevated in reactivity. A frequency histogram of a negative anti-HCV population from the Interstate Blood Bank was prepared (not shown). A cutoff set at S/N=2.0 gave 5 standard deviations from the negative population mean. No negative specimen gave S/N > 1.6.

10

EXAMPLE 14Production and use of cell lines 16-296-534, 6-914-518 and 6-1070-110A. Production of recombinant HCV antigens and immunogens

Recombinant antigens were made as fusion proteins with CMP-XDO synthetase in 15 E. coli according to methods known to those in the art, or as non fused protein with λ PL promoter system. The following proteins were cloned and purified:

APL-CORE (1-150)
CKS-CORE (1-150)
CKS-33c (1191-1457)
20 λ PL-33c-CORE (fusion of 1191-1457 and 1-150)
CKS-BCD (156 -1930)
CKS-E (1931-2189) (NS4/NS5 reaction)
CKS-B (1676-1790)

25 See FIG. 1B for map of HCV genome and approximate locations of HCV regions.

Recombinant protein C-100 (1569-1930) was obtained from Chiron as a fusion protein with superoxide dismutase (SOD). All recombinant proteins were greater than 90% pure by SDS-PAGE.

30 B. Immunization of Mice

BALB/c mice (Charles River Laboratories, Charles River, NY), 6-8 weeks old, were initially immunized subcutaneously and intraperitoneally with 10-100 μ g of the HCV CKS-33C in 100 μ l of Freund's complete adjuvant (Difco, Detroit, MI). On day 14, a second similar boost was administered with the immunogen emulsified in incomplete 35 Freund's adjuvant (Difco). On day 25, 10-100 μ g of the immunogen was diluted into 100 μ l of phosphate buffered saline (PBS), pH 7.2, and injected intravenously into the

tail vein (J. Goding, Monoclonal Antibodies: Principles and Practice [New York; Academic Press, 1986]). Sera titers were not evaluated.

C. Fusion

5 On day 28, mice were sacrificed and splenocytes were fused in a 1:1 ratio with the SP2/0 myeloma line according to known conventional methods (G. Kohler and C. Milstein, Nature (1975) 256:495-497; J. Goding, *supra*). The cell fusion pellet was dispersed with 1 ml 50% polyethylene glycol (PEG) (American Type Culture Collection, MW 1450) and centrifuged in Iscove's Modified Dulbecco's Medium (IMDM) (Gibco, Grand Island, NY). The cells were resuspended in HAT (hypoxanthine-aminopterin-thymidine)-selective IMDM with 10% fetal bovine serum (FBS) (Hyclone Laboratories, Logan, UT) and plated at 3×10^5 cells per 96-well tissue culture plates. Growth promoters included in the HAT media were 0.5% STM (RIBI Immunochem Research, Inc., Hamilton, MT) and 1% Origen Hybridoma Cloning Factor (Igen, Rockville, MD). Growth medium was replaced in culture wells post-fusion on day 5 and 7 using HT (hypoxanthine-thymidine) supplemented IMDM with 10% FBS.

D. Enzyme Immunoassay (EIA)

Culture supernates were EIA screened 10 days post-fusion against the immunizing antigen to detect hybrids secreting HCV specific antibody and a non-specific protein to eliminate any false positives (Langone & Van Vunakis, eds., Methods in Enzymology, 92:168-174, Academic Press [1983]). Polystyrene 96-well microtiter plates were coated overnight at room temperature with 100 μ l per well of a 1 μ g/ml HCV antigen solution in PBS. Any remaining binding sites on the polystyrene wells were blocked with 3% bovine serum albumin (BSA) (Intergen, Purchase, NY) in PBS for 30 minutes at room temperature. Plates were washed three times with distilled water. Fifty microliters of hybridoma tissue culture supernatants were incubated for 1 hour at room temperature in the wells, and the wells were washed three times with distilled water. Antibody binding to antigen was detected using goat anti-mouse IgG+M-horseradish peroxidase (HRPO) (Kirkegaard-Perry Laboratories [KPL], Gaithersburg, MD) diluted at a concentration of 1:1000 in the block solution and incubated 30 minutes at room temperature. The plates were washed with distilled water and o-phenylenediamine substrate (OPD; Abbott Laboratories, Abbott Park, IL) was used as the chromogen. Plates were read at 492 nm. Hybrid cultures were regarded as potential HCV antibody-positive when the optical density (OD) was 3 times the negative control (NC) and significant preferential to the HCV antigen plate was observed.

compared to antibody binding of the irrelevant antigen coated plate, i.e.: >0.2 OD difference and <0.2 OD signal on the latter.

E. Western Blot

5 Hybrid antibody specificity was confirmed with Western blot analysis (Towbin & Gordon, J. Immunol. Methods, 72:313-340 [1984]). HCV recombinant proteins and irrelevant proteins were electrophoresed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and then transferred to nitrocellulose, according to the manufacturer's instructions (Schleicher & Schuell, Keene, NH; Bio-Rad, Richmond, CA). The nitrocellulose strips were blocked with 1% bovine hemoglobin (Sigma Chemical Co., St. Louis, MO) and 0.5% Tween-20 (Fisher Scientific, Pittsburgh, PA) in PBS for 30 minutes at room temperature, then the strips were incubated with hybrid tissue culture supernatant. The strips were then washed in PBS and goat anti-mouse IgG+M-HRPO (KPL) added for 30 minutes. Antibody binding to the HCV antigen was visualized with 4-chloro-1-naphthol (Sigma) as the chromogenic substrate. Hybrid cultures were cloned and placed in cryostorage if HCV antibody specificity was demonstrated.

F. Establishment of Clones

20 HCV specific hybrids were cloned by limiting dilution (Goding, Monoclonal Antibodies: Principles and Practices, 2nd ed, Academic Press, New York [1986]). Modifications included plating of the cultures in log₁₀ dilution series and selecting positive clones for expansion from plates which exhibit <20% growth per 96 well tissue culture plate. Culture supernates were tested after 10 days using the EIA and 25 Western blot procedures described above. The selected clones were expanded for further evaluation and cryostoraged in 80% IMDM with 10% FBS and 10% DMSO (Sigma).

G. Monoclonal Antibody Isotype

30 Monoclonal antibody isotype was determined with the SBA Clonotyping System III kit (Southern Biotechnology Associates, Inc., Birmingham, AL) with modifications. EIA 96-well microtiter plates were coated overnight at room temperature with 100 µl/well of a 1:1000 dilution of goat anti-mouse IgG+M (H+L) (KPL). Plates were blocked for 30 minutes with 3% BSA in PBS and washed with water. Culture samples 35 were added to the wells, incubated for 1 hour, and washed with water. The kit's goat anti-mouse subtype specific conjugates were added for a 30 minute incubation period.

Following a water wash, color was identified with OPD substrate. The goat anti-mouse isotype specific conjugate that bound to the mouse immunoglobin and displayed a >0.1 OD at 492 nm signaled the subtype.

5 H. Monoclonal Antibody Production

Clones selected for further evaluation were scaled up in tissue culture T-flasks and 10^6 cells were injected into the peritoneal cavity of pre-pristane BALB/c mice (Charles River Biotechnical Services, Inc., Wilmington, MA) (see Hurrell, supra). The resulting ascites fluid was harvested 7-10 days after injection, centrifuged, and stored at -20°C. The IgG antibody was affinity purified on Protein A (Pharmacia-LKB Biotechnologies, Piscataway, NJ) utilizing the automated OROS purification system Model 100 (see Goding, supra, for basic principles). The IgM antibodies were purified by molecular sizing on a S-300 column (Pharmacia-LKB).

15 All the following characterization information was performed with purified monoclonal antibody.

I. Isoelectric Focusing (IEF)

A cell line quality control to ensure consistency of frozen lots included measuring the antibody pI point on an IEF gel apparatus (Bio-Rad) which separates proteins based on net charge. Briefly, a bis-acrylamide-riboflavin solution was applied to an acrylamide gel, exposed to fluorescent lighting for 1 hour, then stored overnight at 4°C. A 1 µg sample of monoclonal antibody and standards were laid upon the gel and electrophoresed over a 1-2 hour period. Following a series of fixatives and washes, the gel was silver stained (Bio-Rad). The pI value of the monoclonal antibody was calculated by migratory distance through the gel and was directly compared to the protein standards' migratory distance of known pI values. The distinctive finger print banding pattern reflected the pI microheterogeneity between independently produced lots of antibody (Hamilton, R.G., Reimer, C.B., Rodkey, L.S. (1987) Quality control of murine monoclonal antibodies using isoelectric focusing affinity immunoblot analysis. Hybridoma 6:205-217).

J. EIA and Western Blot Specificity of Monoclonal Antibodies

All monoclonal antibodies noted herein were screened on an assortment of available recombinant HCV antigens as disclosed in U. S. Patent Application Serial No. 07/572,822 entitled HEPATITIS C ASSAY UTILIZING RECOMBINANT PROTEINS, which

enjoys common ownership and is incorporated herein by reference. The procedures are outlined above. The multiple antigen screening technique confirmed the HCV specificity and to exclude the HCV non-specific CKS, IPL, or linker-arm reactivity of the monoclonal antibodies.

5

K. EIA Epitope Competition Studies

To investigate specificity and antigen binding distinctions, epitope grouping experiments were performed utilizing biotin labeled and unlabeled monoclonal antibodies (Langone & Van Yunakis, Methods in Enzymology, 92:242-253, Academic Press [1983]). Briefly, the antibodies were labeled with NHS-LC-biotin (Pierce Chemical Co., Rockford, IL) according to the manufacturer's instructions. Microtiter wells were coated with the immunogen as previously described. First, \log_2 dilutions of the unlabeled antibody were pre-incubated in the wells for 15 minutes, followed by the addition of a fixed amount of biotinylated antibody (the dilution in a direct EIA of the biotinylated antibody alone which gave a value of 50% of the maximum absorbance value) and incubated for 20 minutes. Plates were washed three times with water. Diluted streptavidin-HRPO (Zymed, South San Francisco, CA) was added to the wells and incubated for 30 minutes. The plates were washed again and OPD color developed as previously described. The absorbance was read at 492 nm. Antibodies of the same or related epitope had signal blocked or inhibited by >50%. No inhibition was observed with antibodies of distinct specificity. This was performed reciprocally for antibodies produced within HCV core region.

L. RIA Reciprocal Competition

25 Beads coated with the appropriate antigen or peptide were incubated with 100 μ l of unlabeled monoclonal antibody diluted into recalcified negative human plasma (NHP, testing negative for anti-HCV, anti-HIV and HBsAg) at monoclonal antibody concentrations of 1-20 μ g/ml. 100 μ l of radiolabeled antibody at 1 to 4 μ Ci/ml diluted into HTLV I kit specimen diluent (containing detergent, animal sera, buffer, available from Abbott Laboratories, Abbott Park, IL) was incubated with the bead for 2 hours at 45°C or 18-20 hours at 20-25°C. Beads were washed and counted for radioactivity.

M. HCV Antigen Assays

35 Beads coated with one or a cocktail of anti-HCV monoclonal antibody were incubated with 200 μ l of specimen for 2 hours at 40-45°C or 18-20 hours at 20-25°C. Beads were washed with distilled water and then incubated with 200 μ l of

radiolabeled anti-HCV monoclonal antibody (one or more) for 2 hours at 45°C. Beads were washed and counted in a gamma counter.

N. Characterization of Monoclonal Antibody

5 Two monoclonal antibody competition groups against the HCV 33c region (1191-1457) are found in Table 10. The groups reacted with HCV CKS-33c, CKS-33c-CORE, and λ PL-33c-CORE fusion proteins. Referring to FIGS. 6 TO 17, the reactivities summarized below in Table 10 are shown in lanes 1 and 2. Lanes 1 to 3 contain 10 monoclonal antibodies against HCV 33C protein (6-296-534 in lane 1, 6-914-518 in lane 2 and 6-1070-110 in lane 3); lanes 4-6 contain monoclonal antibodies against HCV CORE (13-975-157 in lane 4, 14-153-234 in lane 5 and 14-1350-210 in lane 6); lanes 7 and 8 contain monoclonal antibodies against the putative HCV ENV region (16-407-209 in lane 7 and 16-803-174 in lane 8); lanes 9-11 contain monoclonal 15 antibodies against HCV C-100 (25-1518-105 in lane 9, 28-735-355 in lane 10; lane 11 contains CKS control monoclonal antibody (29-121-236); lane 12 contains a normal mouse serum control; and lane 13 contains a negative control.

TABLE 10
Anti-HCV-33c Monoclonal Antibodies
EIA and Western Blot Reactivity

25	Group #	Cell Line	pI	Recombinant HCV Antigen Reactivity							
				Isotype	λ PL-		CKS-		CKS-		CKS-
					CKS-	λ PL-	33c-	CKS-	33c-	CKS-	B
	1	6-296-534	6.0	IgG1 k	-	-	+	+	+	-	-
	2	6-914-518		IgG2b k	-	-	-	+	+	-	-

EXAMPLE 15

Anti-HCV-33c Competitive Assay

30 A one step competitive assay was developed using CKS-33c coated beads (0.1 μ g/ml) coating) and radiolabeled group 1 (6-296-534) and group 2 (6-914-518) monoclonal antibody for detection. Seven anti-C100 false positive sera samples and ten anti-C100 true positive sera samples obtained from the Interstate Blood Bank were 35 tested using the two anti-33c monoclonal antibodies of the invention (Table 2). The false positive specimens gave no more than 25% inhibition for any of the monoclonal antibodies. Binding of monoclonal antibody 6-296-534 was almost completely inhibited by the anti-C100 positive specimen (84-100%). Interestingly, these

specimens were not able to effectively compete group 2 monoclonal antibody for binding to 33c. (0-24% inhibition).

Subclones of Table 11 hybrids have been established demonstrating comparable reactivity by EIA and RIA competitive binding studies.

5

TABLE 11
Competitive anti-HCV 33c Assays

Label: 6-296 in HTLV I diluent						
	Specimen	CPM	Avg	S/N	% Inhib.	Result
10	NC	72473	70641			
		67993				
		71456				
	Sacramento negative specimens					
15	145	55325	0.78	21.7		
	146	65197	0.92	7.7		
	148	53705	0.76	24.0		
	149	57741	0.82	18.3		
	151	89301	1.26	-26.4		
20	152	88711	1.26	-25.6		
	154	67721	0.96	4.1		
	Sacramento positive specimens					
	147	5366	0.08	92.4	+	
	150	3351	0.05	95.3	+	
25	153	240	0.00	99.7	+	
	156	128	0.00	99.8	+	
	159	3045	0.04	95.7	+	
	160	47	0.00	99.9	+	
	161	11276	0.16	84.0	+	
30	163	42	0.00	99.9	+	
	165	1076	0.02	98.5	+	
	167	66	0.00	99.9	+	
Label: 6-914 in HTLV I diluent						
35	Specimen	CPM	Avg	S/N	% Inhib.	Result
	NC	32156	34010			
		34431				
		35444				
	Sacramento negative specimens					
40	145	38613	1.14		-13.5	-
	146	36134	1.06		-6.2	-
	148	35130	1.03		-3.3	-
	149	30137	0.89		11.4	-
	151	36794	1.08		-8.2	-
45	152	30860	0.91		9.3	-
	154	34493	1.01		-1.4	-
	Sacramento positive specimens					
	Specimen	CPM	Avg	S/N	% Inhib.	Result
	147	29573	0.87		13.0	-
50	150	34219	1.01		-0.6	-

39

	153	28002	0.82	17.7
	156	31866	0.94	6.3
	159	32633	0.96	4.0
	160	30984	0.91	8.9
5	161	34569	1.02	-1.6
	163	27479	0.81	19.2
	165	25865	0.76	23.9

10

EXAMPLE 16HCV-33c Antigen Assay

Results from a two step 33c antigen RIA assay are shown in Table 12. Assay specimens which gave S/N values greater than 4.0 were considered reactive. The assay was able to detect CKS-33c and IPL-33c-CORE at 300 ng/ml and 2.0 µg/ml respectively. Two of seven specimens from patients with elevated ALT and anti-G-100 were reactive for 33c antigen.

TABLE 12
HCV-33c Ag Assay

		(6-914-518 bead/6-296-534.6-1070-110 label)					
		Specimen	CPM	Average	S/N	Result	
		NC	327 287 322	312			
20		CKS-33c	36 µg/ml 3.6 µg/ml 360 ng/ml 36 ng/ml 3.6 ng/ml	17439 18810 5256 5010 1401 1279 408 415 305 336	18125 5133 1340 4.29	58.09 16.45	+ +
25							
30							
35		IPL-33c-CORE	248 µg/ml 24.8 µg/ml 2.48 g/ml 248 mg/ml 24.8 ng/ml	5832 5913 3607 3483 1261 1476 455 487 310 352	5873 3545 1369 4.39	18.82 11.36 + +	
40							
45		Elevated ALT	27 238	1999 717 315	6.41 1.65	+ -	

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recognize serum Ag both free and bound with human antibody. Furthermore, "cryptic" or hidden antigens or antigenic determinants may be uncovered by treatment of specimen with detergent or reducing agent or both. For example, CORE antigen may exist in a capsid form covered by the virus envelope. Stripping the envelope with detergent should

- 5 expose CORE antigen. Monoclonal antibodies may also offer pragmatic advantages over high titer polyclonal antibody in giving greater sensitivity in assay or allowing shorter incubation times.

Further, antibody immunoassays, one or two step competitive assays, were developed in which anti-HCV competed with labeled anti-HCV monoclonal antibody for binding to a limited number of antigenic sites. A more sensitive competitive assay may be developed in which human anti-HCV binds to HCV Ag in solution blocking or inhibiting the HCV Ag binding in HCV Ag sandwich assay. Competitive assays using monoclonal antibodies allow a more precise mapping of human antibody epitopes and may be useful for determining virus neutralizing antibody epitopes. Some monoclonal antibodies may have virus neutralizing activity. Finally, monoclonal antibodies should be useful in immunoaffinity purification of native viral and recombinant HCV antigens and proteins.

The hybridomas which produce the monoclonal antibodies of the invention are identified as hybridoma H81C17 producing monoclonal antibody H81C17, hybridoma H35C54 producing monoclonal antibody H35C54, hybridoma H28C110 producing monoclonal antibody H28C110, hybridoma H4C20 producing monoclonal antibody H4C20, hybridoma H11C130 producing monoclonal antibody H11C130 and hybridoma H1C46 producing monoclonal antibody H1C46. Hybridomas H28C110, H81C17 and H11C130 were deposited at the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852 as of October 30, 1990, and have been accorded the following deposit numbers: H28C110 was accorded A.T.C.C. deposit No. HB 10587; H81C17 was accorded A.T.C.C. deposit No. HB10588 and H11C130 was accorded A.T.C.C. deposit No. HB 10589. Hybridomas H35C54, H4C20 and H1C46 were deposited at the American Type Culture Collection, 10231 Parklawn Drive, Rockville, Maryland 20852 as of October 31, 1990, and were accorded the following deposit numbers: H35C54 was accorded A.T.C.C. deposit No. HB 10592; H4C20 was accorded A.T.C.C. deposit No. HB 10593 and H1C46 was accorded A.T.C.C. deposit No. HB 10594.

35 Also, the hybridoma cell lines which produce the monoclonal antibodies of the invention are identified as hybridoma cell line 13-975-157 (producing monoclonal

antibody 13-975-157), hybridoma cell line 14-153-234 (producing monoclonal antibody 14-153-234) and hybridoma cell line 14-1350-210 (producing monoclonal antibody 14-1350-210). These hybridoma cell lines were deposited at the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852 on 5 November 16, 1990 and were accorded the following deposit numbers: Hybridoma cell line 13-975-157 was accorded A.T.C.C. deposit No. HB 10608, hybridoma cell line 14-153-234 was accorded A.T.C.C. deposit No. HB 10604, and hybridoma cell line 14-1350-210 was accorded A.T.C.C. deposit No. HB 10602.

10 Further, the hybridoma cell lines which secrete the monoclonal antibodies of the invention are identified as hybridoma cell line 6-296-534 (secreting monoclonal antibody 6-296-534) and hybridoma cell line 6-914-518 (secreting hybridoma cell line 6-914-518). These hybridoma cell lines were deposited at the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852 on November 15 16, 1990 and were accorded the following deposit numbers: Hybridoma cell line 6-296-534 was accorded A.T.C.C. deposit No. HB 10607, and hybridoma cell line 6-914-518 was accorded A.T.C.C. deposit No. HB 10600.

Other variations of applications of the use of the unique monoclonal antibodies 20 provided herein include the detection of HCV antigen in immune complexes, or latent and/or cryptic antigens, and/or associated with viral nucleic acid for detection of the nucleic acid by PCR, LCR, or by direct hybridization. Still other variations and modifications of the specific embodiments of the invention as set forth herein will be apparent to those skilled in the art. Accordingly, the invention is intended to be limited 25 only in accordance with the appended claims.

TABLE 1
CHARACTERIZATION OF MONOCLONAL ANTIBODIES TO HCV PROTEINS

MAB ID	Immunogen	Western Blot Reactivity ¹	EIA Titer With Purified IgG ²	Isotype	Competition w/ Immune Human Sera ³	Epitope Specificity
		Cks λ PL fusion construct protein ng/ml	Cks λ PL fusion construct protein ng/ml			a.a. of HCV genome ⁴
H81C17	pHCV34 (HCV-CORE)	+++	+++	16	3.9	IgG1, k
H35C54	pHCV34	+++	++	31	7.8	IgG1, k
H28C110	pHCV23 (HCV C-100)	+++	+++	0.5	0.4	IgG1, k
H4C20	pHCV23 (HCV C-100)	+++	+++	125	30	IgG1, k
H11C130	pHCV29 (HCV 33C)	+++	+++	0.5	0.5	IgG1, k
H1C46	pHCV29 (HCV 33C)	+++	+	0.5	63	IgG1, k
						1-150

1 Data reflects the reactivity of one each monoclonal antibody with the respective immunogen expressed as (a) CKS fusion protein and (b) under λ PL promoter without any fusion protein in *E. coli*. ++ indicates strong reactivity; + indicates weak reactivity.

2 Data reflects the reactivity of each monoclonal with the immunogen as described in 1. The EIA Titer is defined as the concentration of the monoclonal IgG protein in ng/ml which will give the absorbance of 4 times the negative control at 492 nm.

3 +++ indicates strong competition (>80%); + indicates weak competition (approximately 50%).

4 Epitope specificity was determined based on several experiments as described in Examples 5 and 6.

WE CLAIM:

1. A monoclonal antibody which specifically binds to HCV protein C-100 and does not significantly bind to HCV proteins 33C and CORE.
2. The monoclonal antibody of claim 1 secreted by a hybridoma selected from the group consisting of A.T.C.C. deposit Nos. HB 10593 and HB 10587.
3. A monoclonal antibody which specifically binds to HCV protein 33C and does not significantly bind to HCV proteins C-100 and CORE.
4. The monoclonal antibody of claim 3 secreted by a hybridoma selected from the group consisting of A.T.C.C. deposit Nos. HB 10594, HB 10589, HB 10607 and HB 10600.
5. A monoclonal antibody which specifically binds to HCV protein CORE and does not significantly bind to HCV proteins C-100 and 33C.
6. The monoclonal antibody of claim 5 secreted by a hybridoma selected from the group consisting of A.T.C.C. deposit Nos. HB 10592, HB 10588, HB 10608, HB 10604 and HB 10602.
7. A hybridoma which produces a monoclonal antibody which specifically binds to HCV protein C-100 and does not significantly bind to HCV 33C and HCV CORE proteins.
8. The hybridoma of claim 7 selected from the group consisting of A.T.C.C. deposit Nos. HB 10593 and HB 10587.
9. A hybridoma which produces a monoclonal antibody which specifically binds to HCV protein 33C and does not significantly bind to HCV C-100 and HCV CORE proteins.
10. The hybridoma of claim 9 selected from the group consisting of A.T.C.C. deposit Nos. HB 10594, HB 10589, HB 10607 and HB 10600.

11. A hybridoma which produces a monoclonal antibody which specifically binds to HCV protein CORE and does not significantly bind to HCV C-100 and HCV 33C proteins.

12. The hybridoma of claim 11 selected from the group consisting of A.T.C.C. deposit Nos. HB 10592, HB 10588, HB 10608, HB 10604 and HB 10602.

13. A method for determining the presence of Hepatitis C Virus HCV C-100 protein, or HCV 33C protein or HCV CORE protein in a test sample which may contain the Hepatitis C Virus, comprising:

- a. contacting the test sample with an anti-HCV antibody which specifically binds to HCV C-100 protein, or HCV 33C protein or HCV CORE protein attached to a solid phase which antibody specifically binds to HCV protein, to form a mixture;
- b. incubating said mixture for a time and under conditions sufficient to form antigen/antibody complexes;
- c. contacting said complexes with an indicator reagent comprising a signal generating compound capable of generating a measurable detectable signal attached to an anti-HCV antibody which specifically binds to HCV C-100 protein, or HCV 33C protein or HCV CORE protein to form a second mixture;
- d. incubating said second mixture for a time and under conditions sufficient to form antibody/antigen/antibody complexes; and
- e. determining the presence of Hepatitis C Virus in the test sample by detecting the measurable signal generated, wherein the amount of HCV C-100 protein, or HCV 33C protein or HCV CORE protein present in the test sample is proportional to said measurable signal.

14. A competitive assay method for determining the presence and amount of HCV antibody which may be present in a test sample, comprising:

- a. contacting a test sample suspected of containing HCV antibodies with a solid phase coated with HCV 33C, C-100 or HCV CORE proteins and an indicator reagent comprising a signal generating compound and a monoclonal antibody which specifically binds to said proteins, for a time and under conditions sufficient to form antigen/antibody complexes of the test sample and solid phase and/or indicator reagent and solid phase;

b. determining the presence of HCV antibody present in the test sample by detecting the reduction in binding of the indicator reagent to the solid phase as compared to the signal generated from a negative test sample to indicate the presence of HCV antibody in the test sample

15. The method of claim 13 wherein step (a) and (c) are performed simultaneously.

16. The method of claim 13 wherein the anti-HCV antibody attached to the solid phase is a polyclonal antibody.

17. The method of claim 13 or 14 wherein the signal generating compound is selected from the group consisting of a luminescent compound, a chemiluminescent compound, an enzyme and a radioactive element.

18. The method of claim 13 wherein said anti-HCV antibody attached to the solid phase is a monoclonal antibody.

19. The method of claim 13 wherein the indicator reagent comprises a signal generating compound attached to a polyclonal antibody.

20. The method of claim 13 wherein said indicator reagent comprises a signal generating compound attached to a monoclonal antibody.

21. The method of either one of claims 14, 18 or 20 wherein said monoclonal antibody is the monoclonal antibody secreted by a hybridoma selected from the group consisting of A.T.C.C. deposit Nos. HB 10593, HB 10587, HB 10594, HB 10589, HB 10592, HB 10588, HB 10607, HB 10600, HB 10608, HB 10604 and HB 10602.

22. An assay kit for detecting the presence of HCV in a test sample comprising: a container containing a monoclonal antibody which specifically binds either HCV C-100 protein, HCV 33C protein or HCV CORE protein.

23. The assay kit of claim 22 wherein said monoclonal antibody is secreted by a hybridoma selected from the group consisting of A.T.C.C. deposit Nos. HB 10593, HB

10587, HB 10594, HB 10589, HB 10592, HB 10588, HB 10607, HB 10600, HB 10608, HB 10604 and HB 10602, or a combination thereof.

24. A hybridoma cell line selected from the group consisting of A.T.C.C. deposit Nos. HB 10593, HB 10587, HB 10594, HB 10589, HB 10592, HB 10588, HB 10607, HB 10600, HB 10608, HB 10604 and HB 10602.

25. A monoclonal antibody selected from the group consisting of A.T.C.C. deposit Nos. HB 10593, HB 10587, HB 10594, HB 10589, HB 10592, HB 10588, HB 10607, HB 10600, HB 10608, HB 10604 and HB 10602.

1/14

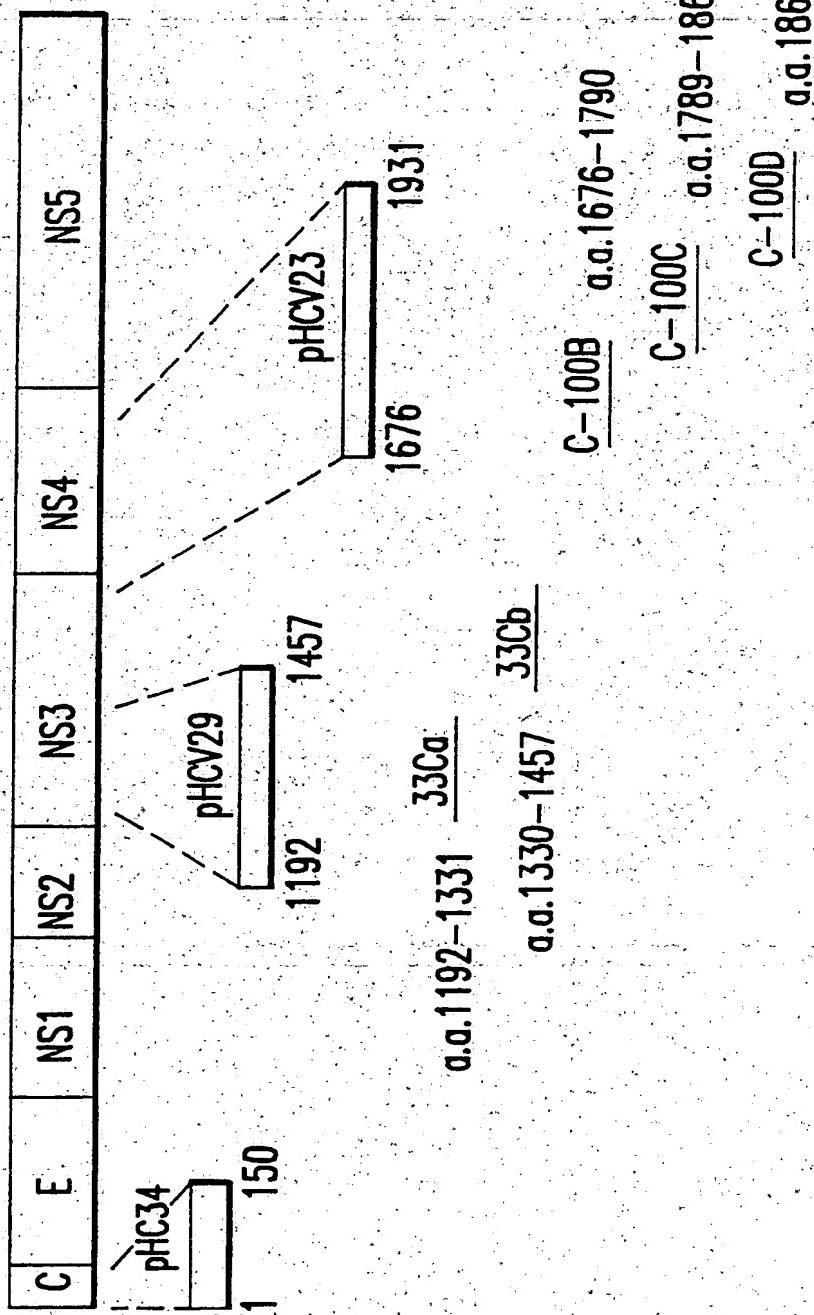
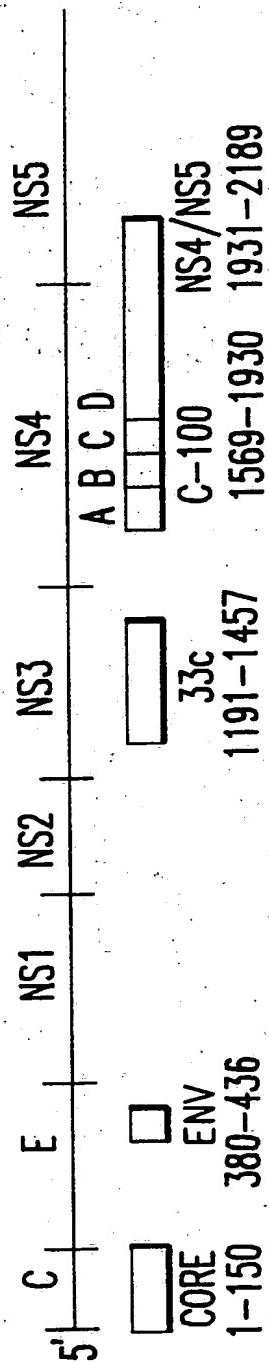


FIG. 1A

2/14

FIG. 1B



SUBSTITUTE SHEET

3/14

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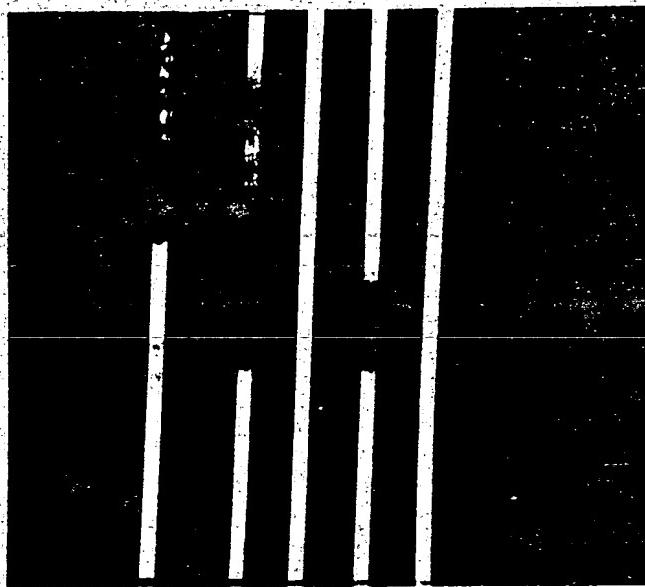


FIG. 2A

1 2 3 4 5

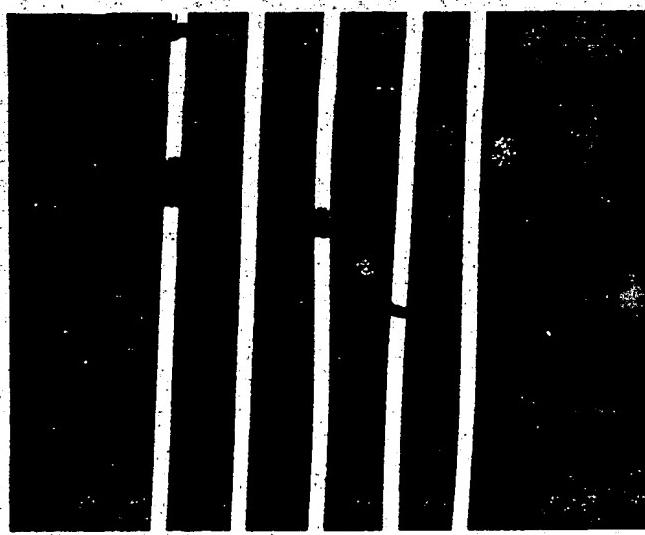


FIG. 2B

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4/14

1 2 3

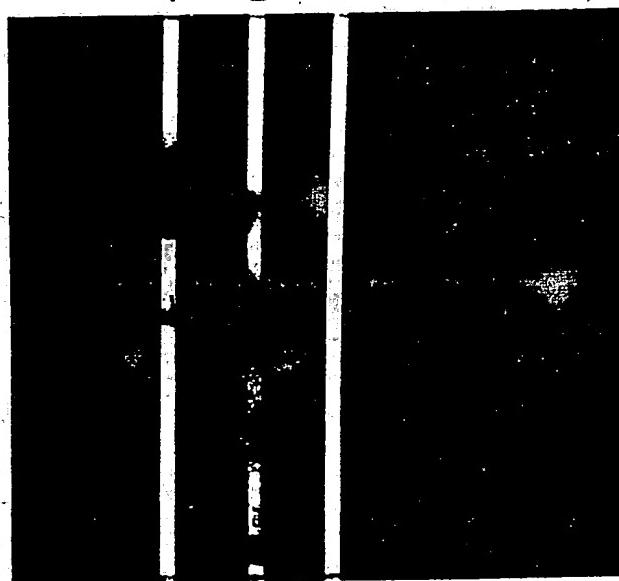


FIG. 2C

1 2 3

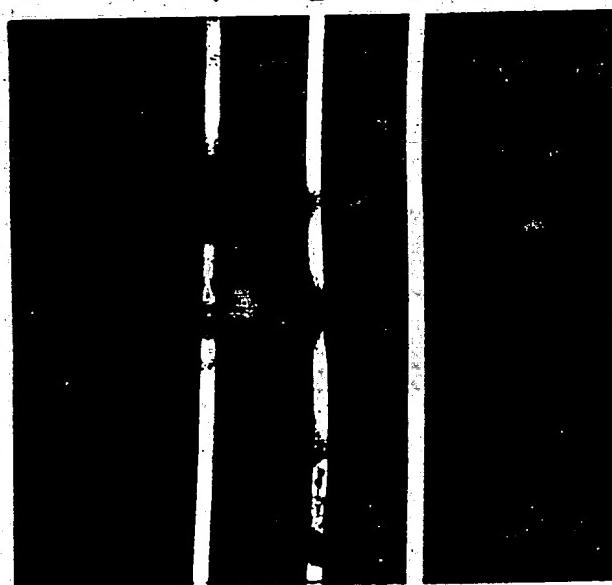


FIG. 2D

5/14

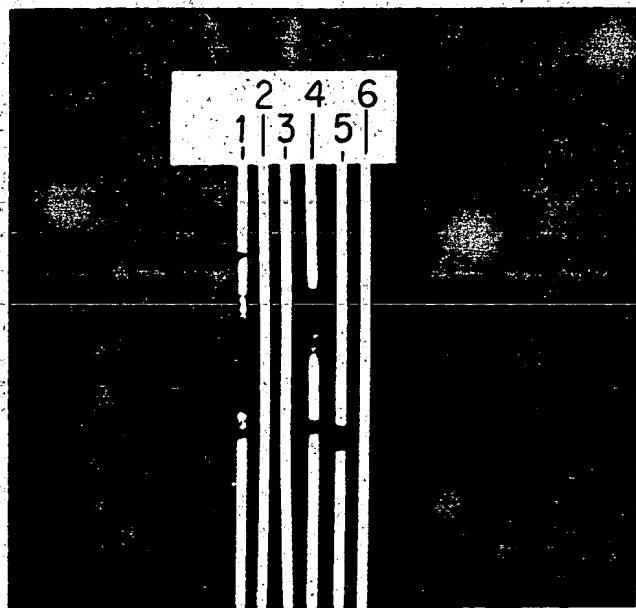


FIG. 2E

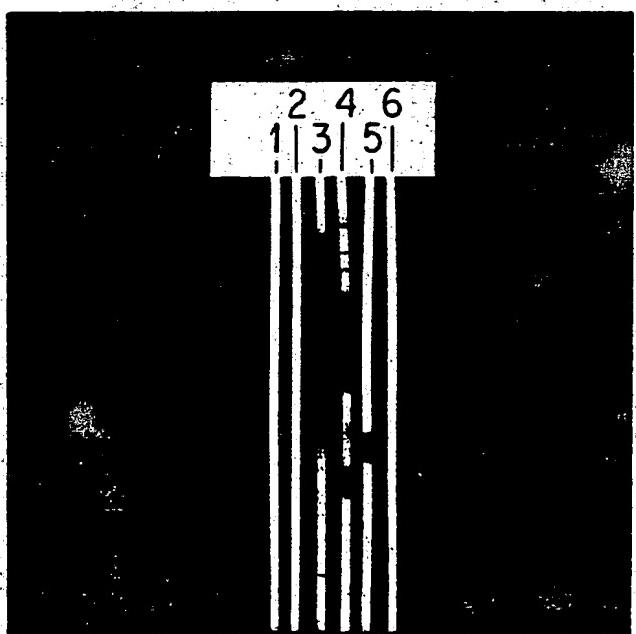
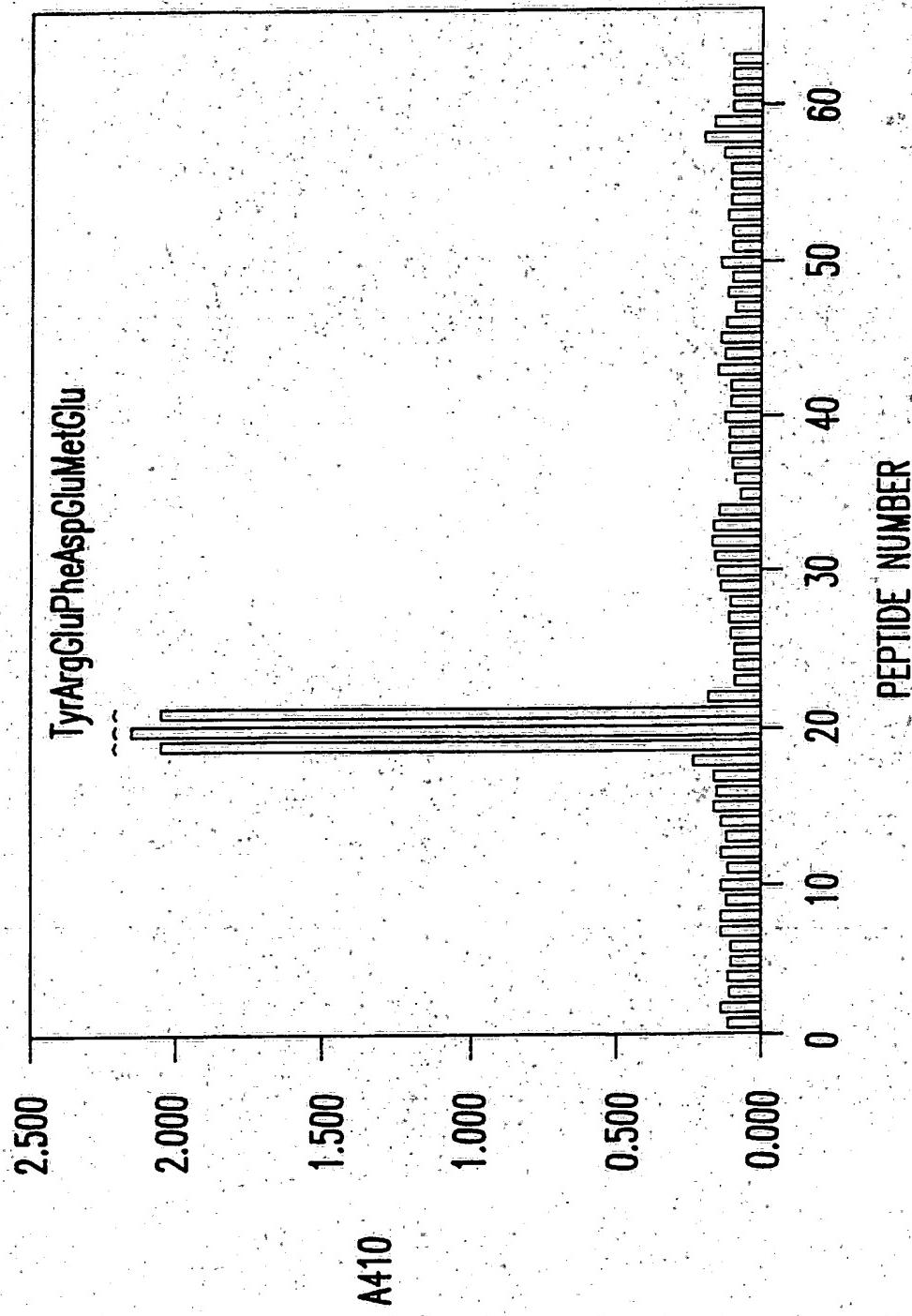


FIG. 2F

6/14

FIG. 3



7/14

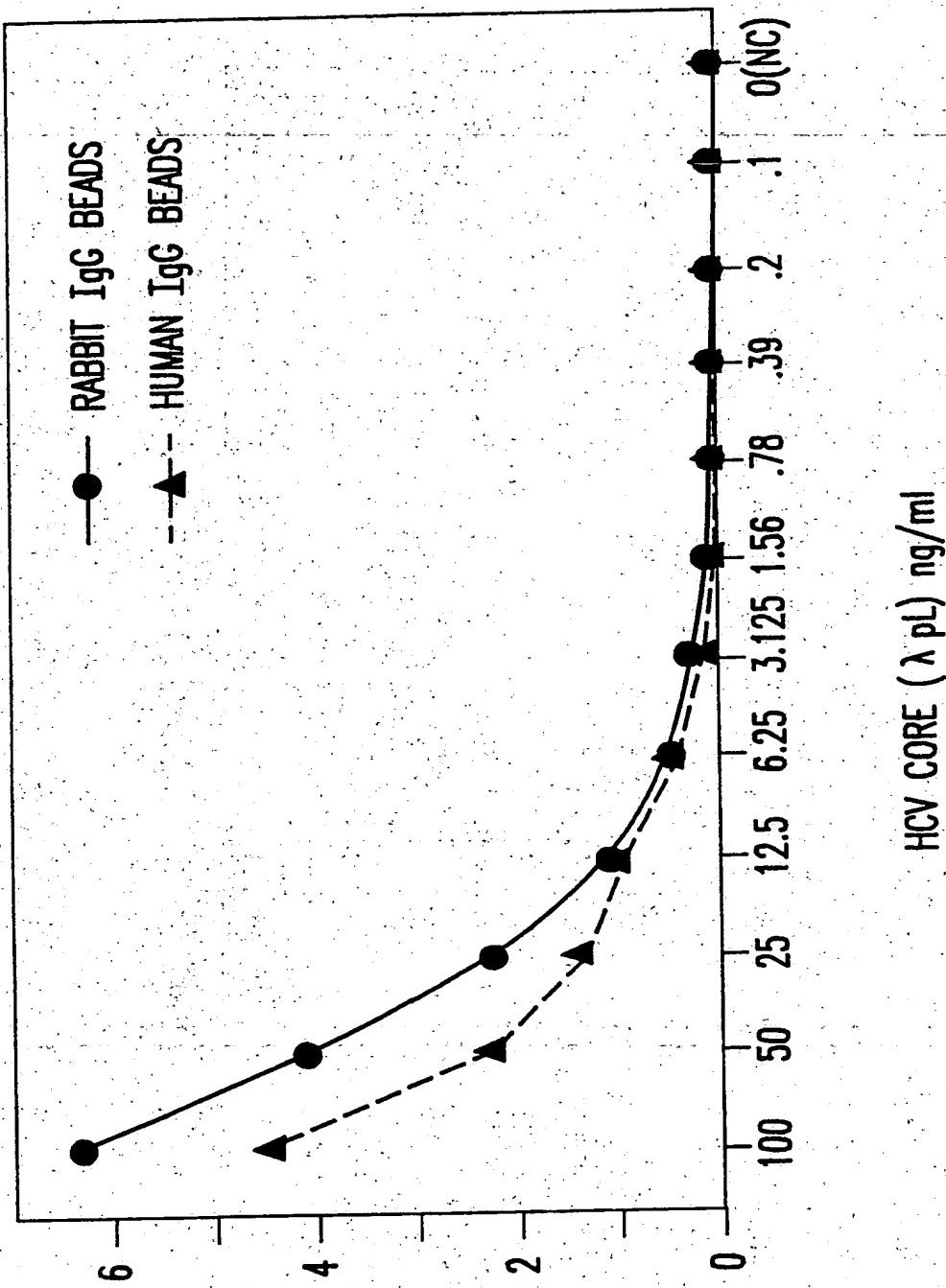


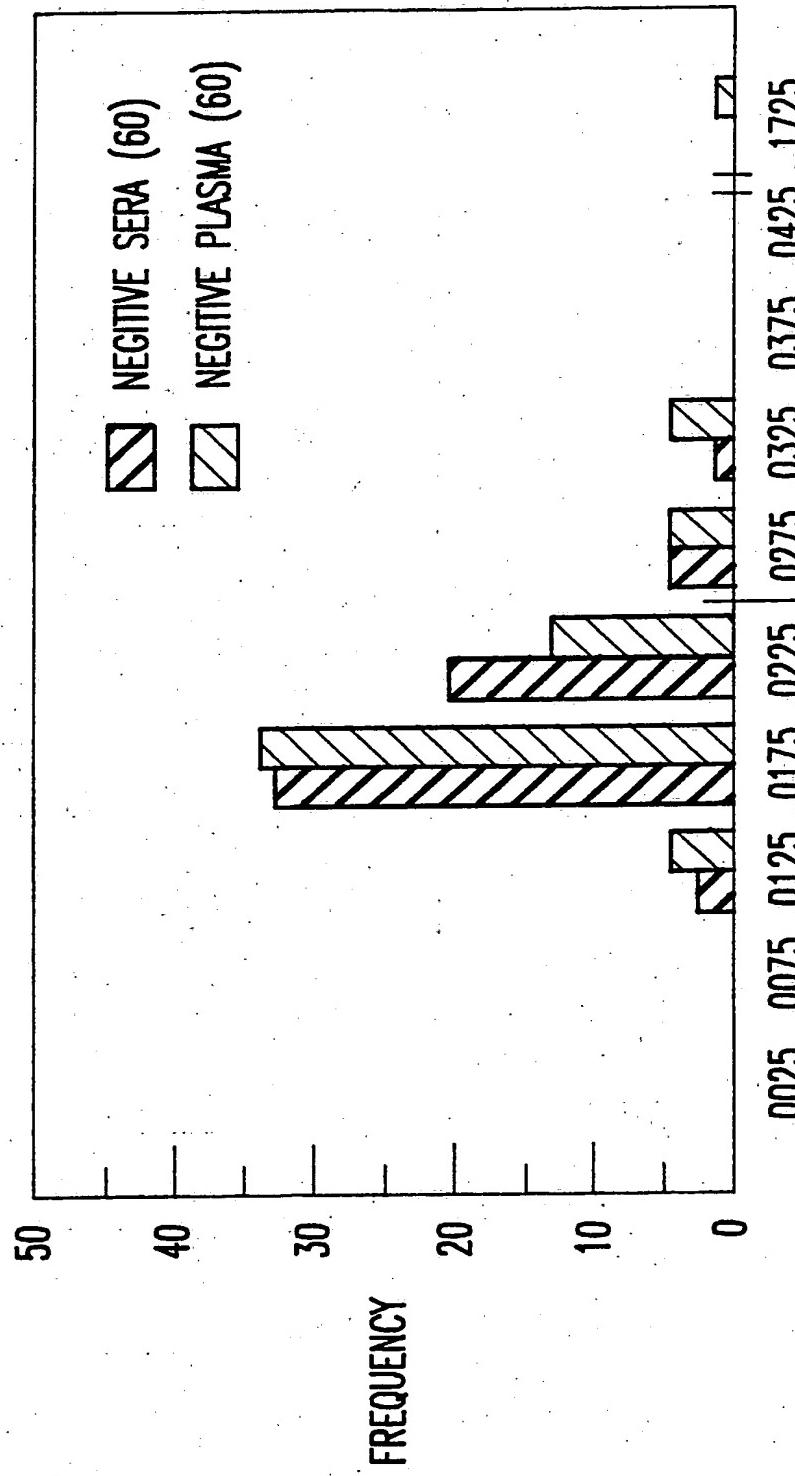
FIG. 4

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8/14

FIG.5



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9/14

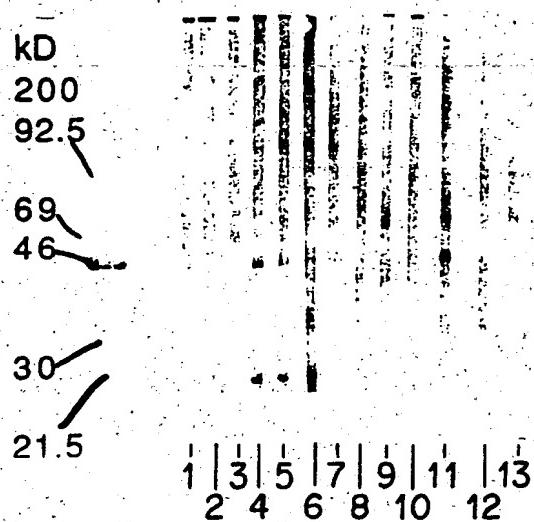


FIG. 6

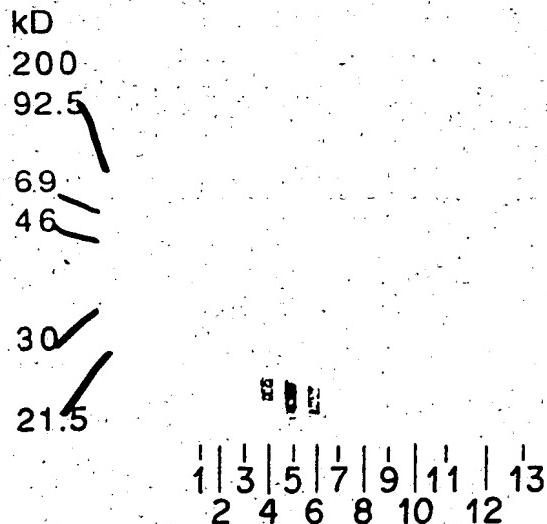


FIG. 7

10/14

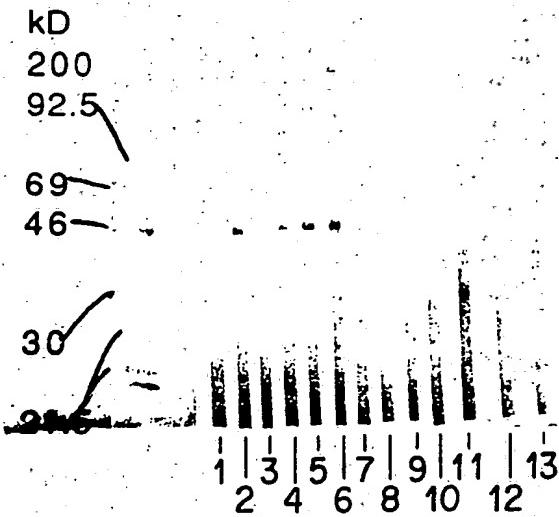


FIG. 8

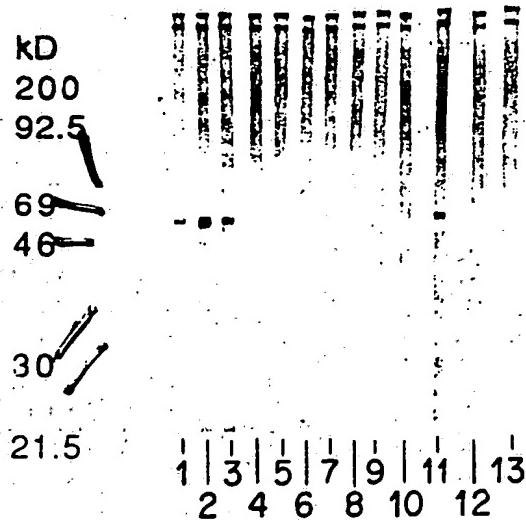


FIG. 9

11/14

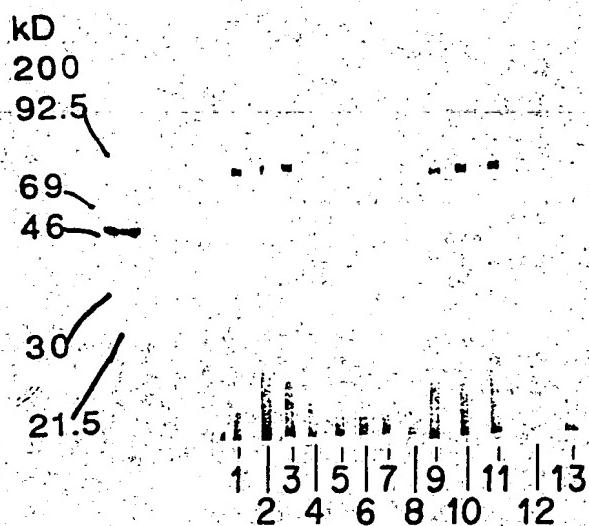


FIG. 10

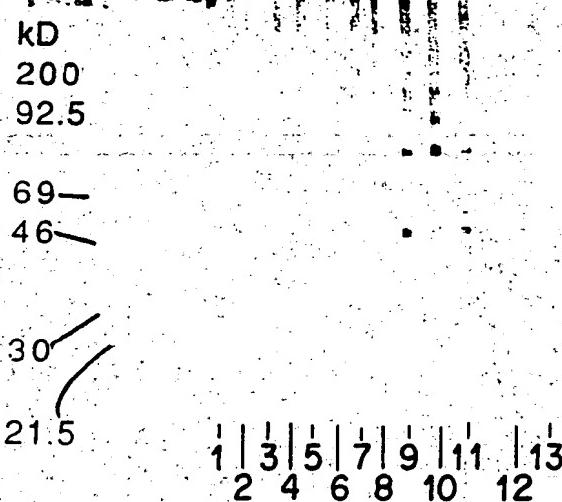


FIG. 11

12/14

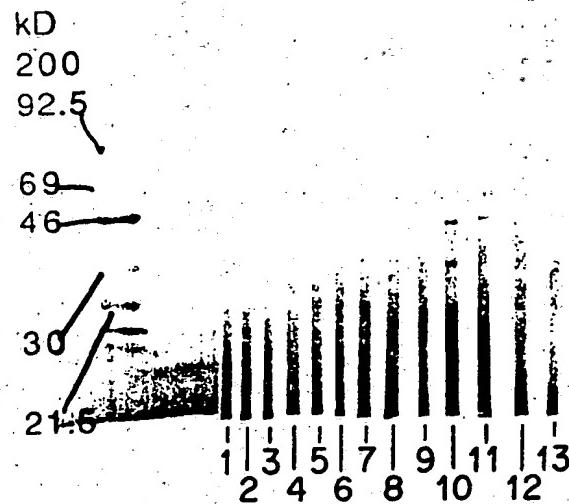


FIG. 12

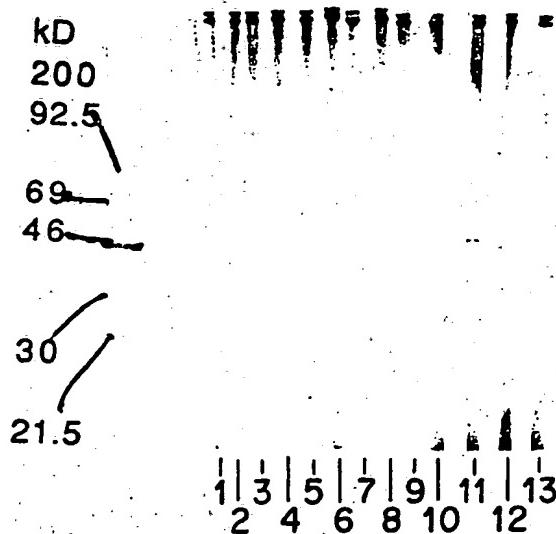


FIG. 13

13 / 14

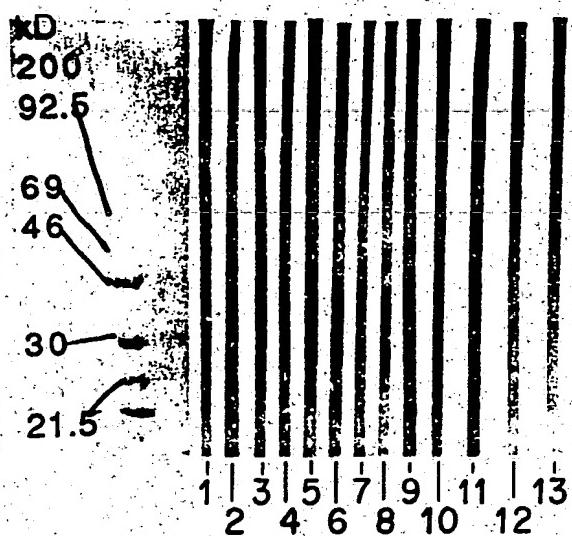


FIG. 14

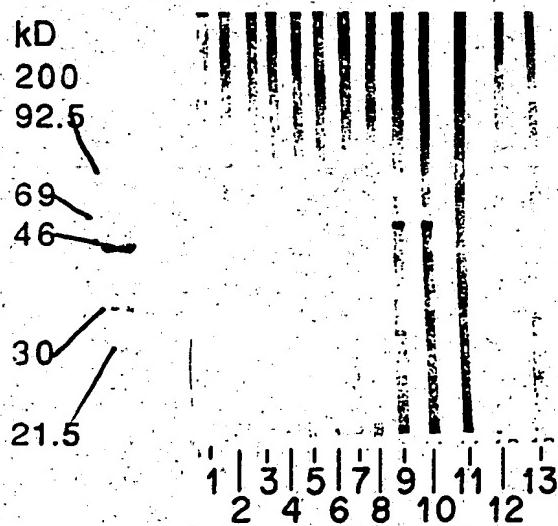


FIG. 15

14 / 14

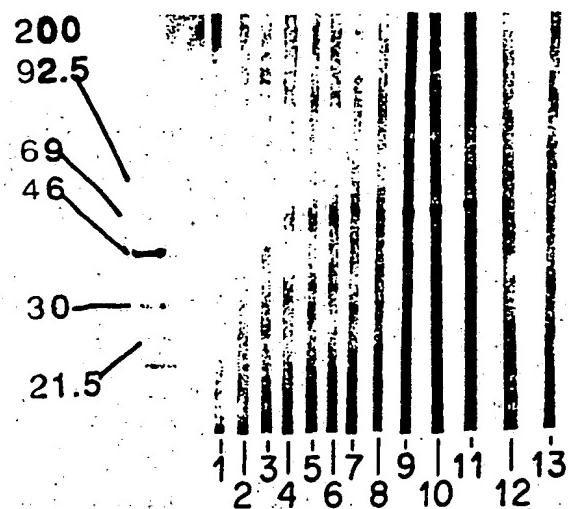


FIG. 16

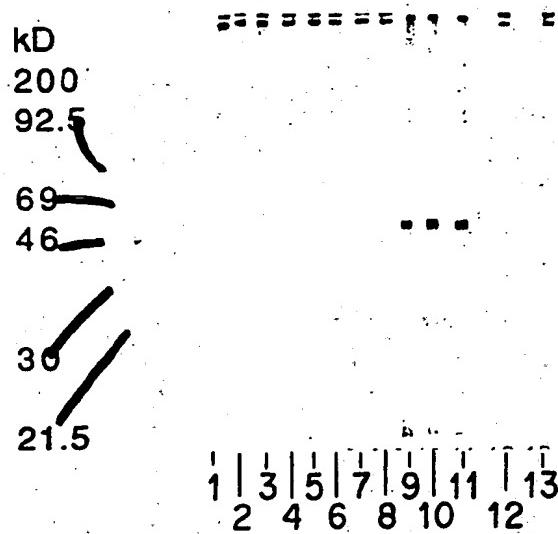


FIG. 17

SUBSTITUTE SHEET

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US91/08273

I. CLASSIFICATION & SUBJECT MATTER (if several classification symbols apply, indicate all) *

According to International Patent Classification (IPC) or to both National Classification and IPC

U.S.Cl.: 530/387; 435/5, 70.21, 240.27

IPC(5): C07K 15/00; C12Q 1/70; C12P 21/02; C12N 5/00

II. FIELDS SEARCHED

Minimum Documentation Searched ?

Classification System	Classification Symbols
U.S.Cl.	530/387; 435/5, 70.21, 240.27; 436/548

Documentation Searched other than Minimum Documentation
to the Extent that such Documents are Included in the Fields Searched *

APS
Dialog

III. DOCUMENTS CONSIDERED TO BE RELEVANT *

Category *	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
X	EP,A, 0,318,216 (Houghton et al.), 31 May 1989, see Claims 1-32	1,7,13-20,22 2-6,8-11,21,23-25
Y	US,A, 4,486,530 (David et al.) 04 December 1984, see entire document.	13, 15-23

* Special categories of cited documents: ¹⁰

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search

04 February 1992

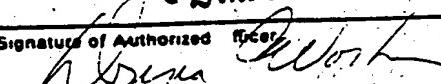
International Searching Authority

ISA/US

Date of Mailing of this International Search Report

02 MAR 1992

Signature of Authorized Officer


Donna C. Wirtman

F02

FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

V. OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE:

This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:

1. Claim numbers _____, because they relate to subject matter¹² not required to be searched by this Authority, namely:

2. Claim numbers _____, because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out¹³, specifically:

3. Claim numbers _____, because they are dependent claims not drafted in accordance with the second and third sentences of PCT Rule 6.4(a).

VI. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING:

This International Searching Authority found multiple inventions in this international application as follows:

- I. Claims 1, 2, 7, 8, 13-24, drawn to a monoclonal antibody and method of use, classified in class 530, subclass 387, and class 435, subclass 5. See attached sheet.

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.

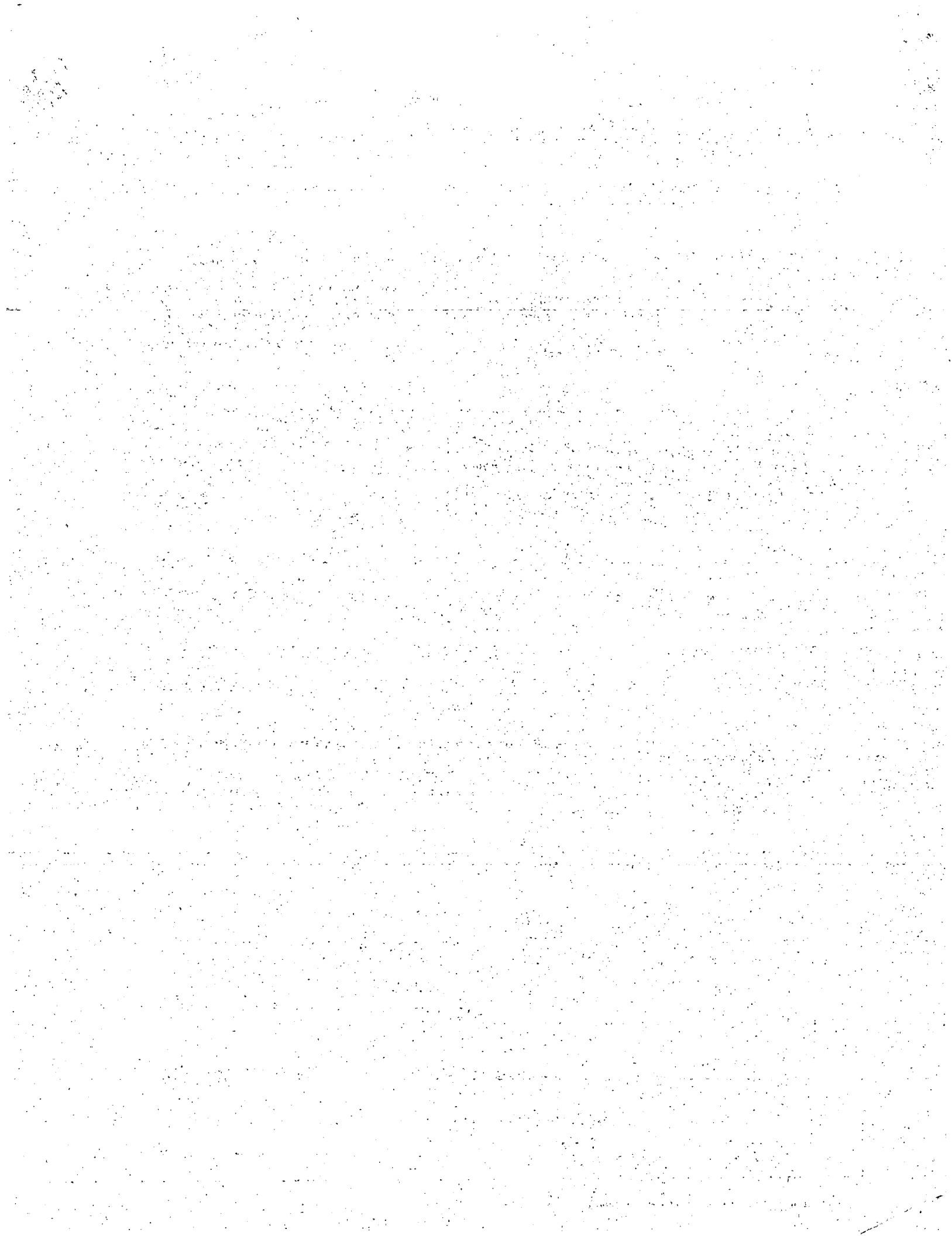
2. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:

3. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:

4. As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not invite payment of any additional fee.

Remarks on Protest:

- The additional search fees were accompanied by applicant's protest.
 No protest accompanied the payment of additional search fees.



**CORRECTED
VERSION ***

PCT

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International Bureau



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 5 : C07K 15/00, C12Q 1/70 C12P 21/02, C12N 5/00		A1	(11) International Publication Number: WO 92/08738 (43) International Publication Date: 29 May 1992 (29.05.92)
(21) International Application Number: PCT/US91/08273		(74) Agents: GORMAN, Edward, H., Jr. et al.; Abbott Laboratories, CHAD 0377/AP6D, One Abbott Park Road, Abbott Park, IL 60064-3500 (US).	
(22) International Filing Date: 7 November 1991 (07.11.91)		(81) Designated States: AT (European patent), AU, BE (European patent), CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FR (European patent), GB (European patent), GR (European patent), IT (European patent), JP, KR, LU (European patent), NL (European patent), SE (European patent).	
(30) Priority data: 610,175 7 November 1990 (07.11.90) US 648,473 31 January 1991 (31.01.91) US 648,477 31 January 1991 (31.01.91) US		Published <i>With international search report.</i>	
(71) Applicant: ABBOTT LABORATORIES [US/US]; Chad 0377/AP6D, One Abbott Park Road, Abbott Park, IL 60064 (US).			
(72) Inventors: MEHTA, Smriti, U. ; 1124 Kristin Drive, Libertyville, IL 60048 (US). JOHNSON-PAEPKE, Jill ; 4726 16th Avenue, Kenosha, WI 53140 (US). GEORGE, Sheila, M. ; 1324 Pimlico Parkway, Libertyville, IL 60048 (US). DESAI, Suresh, M. ; 1408 Amy Lane, Libertyville, IL 60048 (US). MIMMS, Larry, T. ; 8 Shoshoni Trail, Lake Villa, IL 60046 (US). DEVARE, Sushil, G. ; 2492 Farnsworth Lane, Northbrook, IL 60062 (US). TYNER, Joan, D. ; 37835 N. Orchard Road, Beach Park, IL 60087 (US). GIBADLO, Mary, S. ; 945 E. Kenilworth, Palatine, IL 60067 (US).			

(54) Title: MONOCLONAL ANTIBODIES TO HEPATITIS C VIRUS AND METHOD FOR USING SAME

(57) Abstract

Monoclonal antibodies which specifically bind to either Hepatitis C Virus C-100 protein, Hepatitis C Virus 33C protein and Hepatitis C Virus CORE protein, and hybridomas which produce these monoclonal antibodies. Also provided are methods for using these monoclonal antibodies and assay kits containing these antibodies.

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+ Any designation of "SU" has effect in the Russian Federation. It is not yet known whether any such designation has effect in other States of the former Soviet Union.

MONOCLONAL ANTIBODIES TO HEPATITIS C VIRUS AND METHOD FOR USING SAME5 BACKGROUND OF THE INVENTION

This invention relates generally to antibodies which specifically bind to Hepatitis C Virus (HCV), and more specifically, relates to a panel of novel hybridoma cells lines which secrete monoclonal antibodies to HCV proteins C-100, 33C and CORE, and methods for using these monoclonal antibodies.

10

Descriptions of Hepatitis diseases causing jaundice and icterus have been known to man since antiquity. Viral hepatitis is now known to include a group of viral agents with distinctive viral organization protein structure and mode of replication, causing hepatitis with different degrees of severity of hepatic damage through different routes of transmission. Acute viral hepatitis is clinically diagnosed by well-defined patient symptoms including jaundice, hepatic tenderness and an elevated level of liver transaminases such as Aspartate Transaminase and Alanine Transaminase.

Serological assays currently are employed to further distinguish between Hepatitis-A and Hepatitis-B. Non-A Non-B Hepatitis (NANBH) is a term first used in 1975 that described cases of post-transfusion hepatitis not caused by either Hepatitis A Virus or Hepatitis B Virus. Feinstone et al., New Engl. J. Med. 292:454-457 (1975). The diagnosis of NANBH has been made primarily by means of exclusion on the basis of serological analysis for the presence of Hepatitis A and Hepatitis B. NANBH is responsible for about 90% of the cases of post-transfusion hepatitis. Hollinger et al. in N. R. Rose et al., eds., Manual of Clinical Immunology, American Society for Microbiology, Washington, D. C., 558-572 (1986).

Attempts to identify the NANBH virus by virtue of genomic similarity to one of the known hepatitis viruses have failed thus far, suggesting that NANBH has a distinctive genomic organization and structure. Fowler et al., J. Med. Virol. 12:205-213 (1983); and Weiner et al., J. Med. Virol. 21:239-247 (1987). Progress in developing assays to detect antibodies specific for NANBH has been hampered by difficulties encountered in identifying antigens associated with the virus. Wards et al., U. S. Patent No. 4,870,076; Wards et al., Proc. Natl. Acad. Sci. 83:6608-6612 (1986); Ohori et al., J. Med. Virol. 12:161-178 (1983); Bradly et al., Proc. Natl. Acad. Sci. 84:6277-6281 (1987); Akatsuka et al., J. Med. Virol. 20:43-56 (1986).

In May of 1988, a collaborative effort of Chiron Corporation with the Centers for Disease Control resulted in the identification of a putative NANB agent, Hepatitis C Virus (HCV). M. Houghton et al. cloned and expressed in E. coli a NANB agent obtained from the infectious plasma of a chimp. Cuo et al., Science 244:359-361 (1989); Choo et al., Science 244:362-364 (1989). CDNA sequences from HCV were identified which encode antigens that react immunologically with antibodies present in a majority of the patients clinically diagnosed with NANBH. Based on the information available and on the molecular structure of HCV, the genetic makeup of the virus consists of single stranded linear RNA (positive strand) of molecular weight approximately 9.5 kb, and possessing one continuous translational open reading frame. J. A. Cuthbert, Amer. J. Med. Sci. 299:346-355 (1990). It is a small enveloped virus resembling the Flaviviruses. Investigators have made attempts to identify the NANB agent by ultrastructural changes in hepatocytes in infected individuals. H. Gupta, Liver 8:111-115 (1988); D.W. Bradley J. Virol. Methods 10:307-319 (1985). Similar ultrastructural changes in hepatocytes as well as PCR amplified HCV RNA sequences have been detected in NANBH patients as well as in chimps experimentally infected with infectious HCV plasma. T. Shimizu et al., Proc. Natl. Acad. Sci. 87:6441-6444 (1990).

Considerable serological evidence has been found to implicate HCV as the etiological agent for post-transfusion NANBH. H. Alter et al., N. Eng. J. Med. 321:1494, 1500 (1989); Estaben et al., The Lancet: Aug. 5:294-296 (1989); C. Van Der Poel et al., The Lancet Aug. 5:297-298 (1989); G. Sbollì, J. Med. Virol. 30:230-232 (1990); M. Makris et al., The Lancet 335:1117-1119 (1990). Although the detection of HCV antibodies eliminates 70 to 80% of NANBH infected blood from the blood supply system, the antibodies apparently are readily detected during the chronic state of the disease, while only 60% of the samples from the acute NANBH stage are HCV antibody positive. H. Alter et al., New Eng. J. Med. 321:1994-1500 (1989). The prolonged interval between exposure to HCV and antibody detection, and the lack of adequate information regarding the profile of immune response to various structural and non-structural proteins raises questions regarding the infectious state of the patient in the latent and antibody negative phase during NANBH infection. Therefore, there is a need for the development of assay systems to identify acute infection and viremia which may be present. Tools are needed to distinguish between acute and persistent infection and to define the prognostic course of NANBH infection, in order to develop preventive strategies.

SUMMARY OF THE INVENTION

The present invention provides a panel of highly specific and novel monoclonal antibodies that can be employed for the detection of Hepatitis C viral proteins. The 5 monoclonal antibodies specifically bind to either C-100, 33C or CORE antigens, and do not significantly bind to 33C and CORE, C-100 and CORE, and C-100 and 33C, respectively. The hybridomas which produce (secrete) these monoclonal antibodies are identified as follows: H81C17 (A.T.C.C. deposit No. HB 10588, producing monoclonal antibody H81C17), H35C54 (A.T.C.C. deposit No. HB 10592, producing monoclonal 10 antibody H35C54), H28C110 (A.T.C.C. deposit No. HB 10587, producing monoclonal antibody H28C110), H4C20 (A.T.C.C. deposit No. HB 10593, producing monoclonal antibody H4C20), H11C130 (A.T.C.C. deposit No. HB 10589, producing monoclonal antibody H11C130), H1C46 (A.T.C.C. deposit No. HB 10594, producing monoclonal antibody H1C46), 13-975-157 (A.T.C.C. deposit No. HB 10608, producing monoclonal 15 antibody 13-975-157), 14-153-234 (A.T.C.C. deposit No. 10604, producing monoclonal antibody 14-153-234), 14-1350-210 (A.T.C.C. deposit No. HB 10602, producing monoclonal antibody 14-1350-210), 6-296-534 (A.T.C.C. deposit No. HB 10607, secreting monoclonal antibody 6-296-534), and 6-914-518 (A.T.C.C. deposit 20 No. HB 10600, secreting monoclonal antibody 6-914-518). The specificity of these monoclonal antibodies enables advantageous differentiation studies, as well as prognostic and diagnostic applications in the diagnosis and evaluation of NANBH.

In a preferred assay format, a test sample which may contain HCV antigens is contacted with a solid phase to which a polyclonal or a monoclonal anti-HCV antibody or a 25 fragment thereof has been bound, to form a mixture. This mixture is incubated for a time and under conditions sufficient for antigen/antibody complexes to form. The so-formed complexes then are contacted with an indicator reagent comprising a monoclonal or polyclonal antibody or a fragment thereof, specific for the HCV antigen attached to a signal generating compound to form a second mixture. This second mixture is reacted for 30 a time and under conditions sufficient to form antibody/antigen/antibody complexes. The presence of HCV antigen is determined by detecting the measurable signal generated. The amount of HCV present in the test sample, thus the amount of HCV antigen captured on the solid phase, is proportional to the amount of signal generated.

35 Alternatively, an indicator reagent comprising a monoclonal or polyclonal antibody, or fragment thereof, specific for HCV and a signal generating compound is added

to a polyclonal or monoclonal anti-HCV antibody or fragment thereof coated on a solid phase and the test sample to form a mixture. This mixture is incubated for a time and under conditions sufficient to form antibody/antigen/antibody complexes. The presence and amount of HCV present in the test sample, and thus the amount of HCV antigen captured on the solid phase, is determined by detecting the measurable signal. The amount of HCV present in the test sample is proportional to the amount of signal generated.

In another alternate assay format, one or a combination of more than one monoclonal antibody of the invention can be employed as a competitive probe for the detection of antibodies to HCV antigen. For example, HCV CORE antigens, either alone or in combination, can be coated on a solid phase. A test sample suspected of containing antibody to HCV CORE antigen then is incubated with an indicator reagent comprising a signal generating compound and a monoclonal antibody of the invention for a time and under conditions sufficient to form antigen/antibody complexes of either the test sample and indicator reagent to the solid phase or the indicator reagent to the solid phase. The reduction in binding of the monoclonal antibody to the solid phase can be quantitatively measured. A measurable reduction in the signal compared to the signal generated from a confirmed negative NANBH test sample would indicate the presence of anti-HCV CORE antibody in the test sample.

In yet another assay format, a test sample is contacted with a solid phase to which Hepatitis C Virus proteins are attached and an indicator reagent comprising a monoclonal antibody or fragment thereof specific for Hepatitis C Virus attached to a signal generating compound, to form a mixture. The mixture is incubated for a time and under conditions sufficient for antibody/antigen complexes to form. The presence of anti-Hepatitis C virus present in the test sample is determined by detecting the measurable signal generated, and comparing the signal to the measured signal generated from a known negative sample. A measurable reduction of signal of the test sample, compared to the known negative sample's signal, is indicative of the presence of anti-HCV antibodies. Competitive assays for the detection of anti-HCV antibody using antigens free in solution also can be performed.

The presence of Hepatitis C Virus can be detected in a tissue sample by contacting the tissue sample with an indicator reagent comprising a signal generating compound attached to a monoclonal antibody selected from the group consisting of an anti-HCV

C100 antibody or fragment thereof, a monoclonal anti-HCV 33C antibody or fragment thereof, and an anti-HCV CORE antibody or fragment thereof; to form a mixture. This mixture is incubated for a time and under conditions sufficient for antigen/antibody complex to form. The presence of Hepatitis C Virus present in the tissue sample is determined by detecting the signal generated.

Also provided are kits for using the monoclonal antibodies of the invention.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1A is a drawing of the location of the location of the recombinant proteins on the HCV genome, employed as immunogens for generation of the hybridomas of the invention as well as that of subfragments employed for epitope mapping of the monoclonal antibodies of the invention.

FIG. 1B is a map of the HCV GENOME representing the non-structural (NS) genes and the structural genes, core (C) and envelope (E).

FIG. 2A is a photograph of Western blot analysis illustrating the specific binding and epitope mapping of monoclonal antibody H11C130 wherein lane 1 is pHCV29 (a.a. 1192-1457), lane 2 is 33Ca·CKS (a.a. 1192-1331), lane 3 is 33Cb·CKS (a.a. 1330-1457), lane 4 is 33C (λ pL) (a.a. 1192-1457) and lane 5 is CKS;

FIG. 2B is a photograph of Western blot analysis illustrating the specific binding and epitope mapping of monoclonal antibody H1C46 wherein lane 1 is pHCV29 (a.a. 1192-1457), lane 2 is 33Ca·CKS (a.a. 1192-1331), lane 3 is 33Cb·CKS (a.a. 1330-1457), lane 4 is 33C (λ pL) (a.a. 1192-1457) and lane 5 is CKS;

FIG. 2C is a photograph of Western blot analysis illustrating the specific binding of monoclonal antibody H81C17 wherein lane 1 is pHCV34 (a.a. 1-150), lane 2 is HCVCORE (λ pL) and lane 3 is CKS;

FIG. 2D is a photograph of Western blot analysis illustrating the specific binding of monoclonal antibody H35C54 wherein lane 1 is pHCV34 (a.a. 1-150), lane 2 is HCVCORE (λ pL) and lane 3 is CKS;

FIG. 2E is a photograph of Western blot analysis illustrating specific binding and epitope mapping of monoclonal antibody H28C110 wherein lane 1 is C-100B·CKS (a.a. 1676-1790), lane 2 is C-100 C·CKS (a.a. 1789-1863), lane 3 is C-100D·CKS (a.a. 1861-1931), lane 4 is C-100 (λ pL, a.a. 1676-1790) and lane 5 is CKS;

FIG. 2F is a photograph of Western blot analysis illustrating specific binding and epitope mapping of monoclonal antibody H4C20 wherein lane 1 is C-100B·CKS (a.a.

1676-1790), lane 2 is C-100-C-CKS (a.a. 1789-1863), lane 3 is C-100D-CKS (a.a. 1861-1931), lane 4 is C-100 (λ PL, a.a. 1676-1790) and lane 5 is CKS.

FIG. 3 is a profile of PEPSCAN analysis with overlapping hexamer peptides (a.a. 1694-1735) of monoclonal antibody H28C110 illustrating the epitope specificity and amino acid sequence of the HCV genome recognized by monoclonal antibody H28C110.

FIG. 4 is a graph illustrating the efficiency of anti-HCV rabbit and human polyclonal capture antibodies for the detection of HCV core protein in the antigen capture assay of the invention employing monoclonal antibody H81C17.

FIG. 5 is a graph of a negative sera and plasma population distribution tested by an antigen capture EIA wherein frequency is plotted against A492.

FIGS. 6 to 17 are photographs of Western blots showing the reactivity of the monoclonal antibodies of the invention, where

lanes 1 to 3 contain monoclonal antibodies against HCV 33C protein (6-296-534 in lane 1, 6-914-518 in lane 2 and 6-1070-110 in lane 3);

lanes 4-6 contain monoclonal antibodies against HCV CORE (13-975-157 in

lane 4, 14-153-234 in lane 5 and 14-1350-210 in lane 6);

lanes 7 and 8 contain monoclonal antibodies against the putative HCV ENV region (16-407-209 in lane 7 and 16-803-174 in lane 8);

lanes 9-10 contain monoclonal antibodies against HCV C-100 (25-1518-105 in lane 9, 28-735-355 in lane 10);

lane 11 contains monoclonal antibody against CKS (29-121-236 in lane 11);

lane 12 contains a normal mouse serum control; and

lane 13 contains a negative control of antibody diluent.

FIG. 6 is an electroblot of these monoclonal antibodies run against CKS-CORE;

FIG. 7 is an electroblot of these monoclonal antibodies run against λ PL-CORE;

FIG. 8 is an electroblot of these monoclonal antibodies run against λ PL-33C-CORE;

FIG. 9 is an electroblot of these monoclonal antibodies run against CKS-33C;

FIG. 10 is an electroblot of these monoclonal antibodies run against CKS-33C-BCD;

FIG. 11 is an electroblot of these monoclonal antibodies run against CKS-BCD;

FIG. 12 is an electroblot of these monoclonal antibodies run against CKS-B;

- FIG. 13 is an electroblot of these monoclonal antibodies run against CKS-E;
FIG. 14 is an electroblot of these monoclonal antibodies run against CKS;
FIG. 15 is an electroblot of these monoclonal antibodies run against SOD-100;
FIG. 16 is an electroblot of these monoclonal antibodies run against CKS-A'BCD;
5 and
FIG. 17 is an electroblot of these monoclonal antibodies run against CKS-A"BCD.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides novel monoclonal antibodies to HCV proteins C-
10 100, 33C and CORE, methods for using the monoclonal antibodies, and kits which contain
these monoclonal antibodies.

The monoclonal antibodies of the present invention can be employed in various
assay systems to determine the presence, if any, of any or all of HCV proteins C-100,
15 33C or CORE, or a combination thereof. Fragments of the monoclonal antibodies provided
also may be used. For example, in a first assay format, a polyclonal or monoclonal
anti-HCV-100, anti-33C or anti-CORE antibody or a fragment thereof, or a
combination thereof, which has been coated on a solid phase, is contacted with a test
sample which may contain any or all of these HCV proteins or a combination of them, to
20 form a mixture. This mixture is incubated for a time and under conditions sufficient to
form antigen/antibody complexes. Then, an indicator reagent comprising a monoclonal
or a polyclonal antibody or a fragment thereof, which specifically binds to either HCV C-
100 protein, HCV 33C protein or CORE protein, or a combination of these antibodies, to
which a signal generating compound has been attached, is contacted with the
25 antigen/antibody complexes to form a second mixture. This second mixture then is
incubated for a time and under conditions sufficient to form antibody/antigen/antibody
complexes. The presence of HCV protein present in the test sample and captured on the
solid phase, if any, is determined by detecting the measurable signal generated by the
signal generating compound. The amount of HCV protein present in the test sample is
30 proportional to the signal generated.

Alternatively, a polyclonal or monoclonal anti-HCV C-100, anti-HCV 33C or
anti-HCV CORE antibody or fragment thereof, or a combination of these antibodies which
is bound to a solid support, the test sample and an indicator reagent comprising a
35 monoclonal or polyclonal antibody or fragments thereof, which specifically binds to
either HCV C-100, HCV C33, or HCV CORE protein, or a combination of these antibodies

to which a signal generating compound is attached, are contacted to form a mixture. This mixture is incubated for a time and under conditions sufficient to form antibody/antigen/antibody complexes. The presence, if any, of HCV protein(s) present in the test sample and captured on the solid phase is determined by detecting the 5 measurable signal generated by the signal generating compound. The amount of HCV protein(s) present in the test sample is proportional to the signal generated.

In another alternate assay format, one or a combination of more than one monoclonal antibody of the invention can be employed as a competitive probe for the 10 detection of antibodies to HCV protein C-100, 33C or CORE. For example, HCV proteins, either alone or in combination, can be coated on a solid phase. A test sample suspected of containing antibody to Hepatitis C Virus then is incubated with an indicator reagent comprising a signal generating compound and a monoclonal antibody of the invention for a time and under conditions sufficient to form antigen/antibody complexes of either the 15 test sample and indicator reagent to the solid phase or the indicator reagent to the solid phase. The reduction in binding of the monoclonal antibody to the solid phase can be quantitatively measured. A measurable reduction in the signal compared to the signal generated from a confirmed negative NANBH test sample would indicate the presence of anti-HCV antibody in the test sample.

20 In yet another detection method, each of the monoclonal antibodies of the present invention can be employed in the detection of HCV antigens in fixed tissue sections, as well as fixed cells by immunohistochemical analysis.

25 In addition, these monoclonal antibodies can be bound to matrices similar to CNBr-activated sepharose and used for the affinity purification of specific HCV proteins from cell cultures, or biological tissues such as blood and liver.

30 The monoclonal antibodies of the invention can also be used for the generation of chimeric antibodies for therapeutic use, or other similar applications.

35 The monoclonal antibodies or fragments thereof can be provided individually to detect either HCV C-100, HCV 33C or HCV CORE protein. Combinations of the monoclonal antibodies (and fragments thereof) provided herein also may be used together as components in a mixture or "cocktail" of anti-HCV protein antibodies, each having different binding specificities. Thus, this cocktail can include the monoclonal

antibodies of the invention directed to different antigenic determinants of the HCV genome, along with monoclonal antibodies directed to putative HCV ENV region.

The polyclonal antibody or fragment thereof which can be used in the assay

- 5 formats should specifically bind to either HCV C-100 protein, HCV 33C protein or HCV CORE protein. The polyclonal antibody used preferably is of mammalian origin; human, goat, rabbit or sheep anti-HCV polyclonal antibody can be used. Most preferably, the polyclonal antibody is rabbit polyclonal anti-HCV antibody. The polyclonal antibodies used in the assays can be used either alone or as a cocktail of polyclonal antibodies. Since
10 the cocktails used in the assay formats are comprised of either monoclonal antibodies or polyclonal antibodies having different HCV specificity, they would be useful for diagnosis, evaluation and prognosis of HCV infection, as well as for studying HCV protein differentiation and specificity.
- 15 Test samples which can be tested by the methods of the present invention described herein include human and animal body fluids such as whole blood, serum, plasma, cerebrospinal fluid, urine, biological fluids such as cell culture supernatants, fixed tissue specimens and fixed cell specimens. Solid supports are known to those in the art and include the walls of wells of a reaction tray, test tubes, polystyrene beads,
20 magnetic beads, nitrocellulose strips, membranes, microparticles such as latex particles, and others.

The indicator reagent comprises a signal generating compound (label) which is capable of generating a measurable signal detectable by external means conjugated

- 25 (attached) to a specific binding member for HCV. "Specific binding member" as used herein means a member of a specific binding pair. That is, two different molecules where one of the molecules through chemical or physical means specifically binds to the second molecule. In addition to being an antibody member of a specific binding pair for HCV, the indicator reagent also can be a member of any specific binding pair, including
30 either hapten-anti-hapten systems such as biotin or anti-biotin, avidin or biotin, a carbohydrate or a lectin, a complementary nucleotide sequence, an effector or a receptor molecule, an enzyme cofactor and an enzyme, an enzyme inhibitor or an enzyme, and the like. An immunoreactive specific binding member can be an antibody, an antigen, or an antibody/antigen complex that is capable of binding either to HCV as in a sandwich assay,
35 to the capture reagent as in a competitive assay, or to the ancillary specific binding member as in an indirect assay.

The various signal generating compounds (labels) contemplated include chromogens, catalysts such as enzymes, luminescent compounds such as fluorescein and rhodamine, chemiluminescent compounds, radioactive elements, and direct visual labels.

5 Examples of enzymes include alkaline phosphatase, horseradish peroxidase, beta-galactosidase, and the like. The selection of a particular label is not critical, but it will be capable of producing a signal either by itself or in conjunction with one or more additional substances.

10 It is contemplated that the reagent employed for the assay can be provided in the form of a kit with one or more containers such as vials or bottles, with each container containing a separate reagent such as a monoclonal antibody, or a cocktail of monoclonal antibodies, employed in the assay.

15 The following examples demonstrate the advantages and utility of this invention for serodiagnosis of Hepatitis C virus by describing methods for the development, characterization, epitope mapping and clinical utility of these monoclonal antibodies. The methods used for monoclonal antibody development follow procedures known in the art and detailed in Kohler and Milstein, Nature 256:494 (1975), and reviewed in
20 J.G.R. Hurrel, ed., Monoclonal Hybridoma Antibodies: Techniques and Applications, CRC Press, Inc., Boca Raton, FL (1982). Another method of monoclonal antibody development which is based on the Kohler and Milstein method is that of L.T. Mimms et al., Virology 176:604-619 (1990), which is incorporated herein by reference. These examples are meant to illustrate, but not to limit, the spirit and scope of the invention.

25 Examples 1-8 exemplify the production and uses of cell lines 81C17, H35C54, H28C110, H4C20, H11C130 and H1C46. Examples 9-13 exemplify the production and uses of cell lines 13-975-257, 14-153-234 and 14-1350-210. Examples 14-17 exemplify the production and uses of cell lines 6-296-534, 6-914-518 and 6-1070-
30 110.

EXAMPLES

EXAMPLE 1

Immunization of Mice

35 E. coli derived recombinant antigens encoded by HCV sequences, designated as pHCV23 (HCV C-100, a.a. 1677-1931), pHCV29 (HCV 33C, a.a. 1192-1457) and

pHCV34 (HCV-CORE, a.a. 1-150) were employed as immunogens for the generation of murine monoclonal antibodies specific for HCV C-100, 33C and CORE, respectively. Detailed information on the synthesis, cloning and expression of these recombinant proteins is disclosed in U.S. Patent Application Serial No. 07/572,822, which enjoys common ownership and is incorporated herein by reference. These proteins were prepared for immunization with appropriate adjuvants after purification with protein purification methods known to those skilled in the art. FIG. 1A shows the location of recombinant HCV proteins and their subfragments on the genome.

10 Immunization with pHCV23

On day one, BALB/c mice received 15 µg of purified pHCV23 in 200 µl of Freund's complete adjuvant injected intraperitoneally (i.p.). A second immunization was done after 14 days with 15 µg of pHCV23 in incomplete Freund's adjuvant. Mice were bled on day 21 and the immune response to pHCV was assessed by enzyme linked immunoassay (EIA) and Western blot analysis. Fusion was performed after allowing the mice to rest for at least eight weeks.

15 Immunization with pHCV29

On day one, BALB/c mice received 15 µg of purified pHCV29 in 100 µl of Freund's complete adjuvant injected i.p. Subsequent immunizations were done after 14 and 28 days with 15 µg of pHCV23 in incomplete Freund's adjuvant. Mice were bled on day 21 and the immune response to pHCV23 was assessed as described above.

20 Immunization with pHCV34

BALB/c mice were immunized following a schedule similar to that of pHCV29 immunization described above, using RIBI adjuvant system (RIBI Immunochem Research, Hamilton, Montana). On day one, mice received 15 µg of purified pHCV34 with 15 µg each of Trehalose dimycolate (TDM) and *M. phlei* in a buffer emulsion prepared according to the manufacturer's instructions. Subsequent immunizations were performed on day 14, 28 and 42. Mice were bled on days 21 and 49 and the immune response was assessed as described hereinbelow.

25 Enzyme-Linked Immunoassay (EIA)

The immune response to the immunizing antigen was assessed by microtiter EIA and Western blot analysis. Wells of microtiter plates were coated with 100 µl of purified antigen in 0.1M bicarbonate buffer at pH 9.5. After washing with Phosphate

Buffered Saline (PBS) which also contained 0.01% sodium dodecyl sulfate (SDS) and 0.05% Tween-20® (available from Biorad Laboratories, Richmond, CA), free sites were overcoated with 1% BSA in bicarbonate buffer at pH 9.5. Plates were stored at 4°C following a final wash. Sera from native or immunized mice were serially diluted in 5 100 µl of dilution buffer which contained 20 mM sodium phosphate, pH 7.4, 0.15M NaCl, 20% normal goat serum, 10% fetal calf serum, 5 mM EDTA, 10 mM EGTA, 50 mM Tris, 0.2% Tween-20®, with sodium azide as a preservative (at pH 6.8). The diluted sera were reacted with the antigen for three (3) hours at 37°C. The plates were washed and 100 µl of appropriately diluted goat anti-mouse IgG (heavy (H) and light (L) chain) 10 Horseradish Peroxidase (HRPO)-conjugated antibody (Jackson Immunochemicals, West Grove, PA) was added. The plates were incubated at 37°C for two (2) hours. After a final wash, 100 µl of o-phenylenediamine:2HCl (OPD) color reagent was added. The reaction was carried out at room temperature for 10 to 30 minutes, and then stopped by the addition of 1 ml of 1N H₂SO₄. The absorbance at 492/600 nm was recorded, which 15 was found to be directly proportional to the amount of specific antibody bound to the respective antigen.

Western Blot Analysis

Approximately 300 µg of purified rHCV protein were treated with SDS and 2-mercaptoethanol at 95°C, and electrophoresed in a 12% polyacrylamide-SDS gel 20 (Laemmli et al., *Nature* 227:680-685 (1970)). Proteins were transferred overnight from the gel to nitrocellulose by electrophoresis at 100 mamp, or transferred in 1-2 hours at 1.0 amp, in a standard transfer buffer which comprised 25 mM Tris [(Hydroxymethyl) Aminomethane], 192 mM glycine, and 2.0% methanol, pH 8.3. 25 (Towbin et al., *Proc. Natl. Acad. Sci.* 73:4350-4354 [1979]). After transferring the proteins and blocking the nitrocellulose with 5% dry milk in PBS, the nitrocellulose was cut into strips (each strip containing approximately 5 µg of recombinant protein) which then were used to determine the presence of anti-HCV antibody in test sera (or other samples). Reaction mixtures consisted of a nitrocellulose strip incubated with an 30 appropriate amount of test sample in 2.0 ml of buffer (20 mM Tris, 1 mM EDTA, 0.2 M NaCl, 0.3% Triton X-100® and 2 mg/ml bovine serum albumin (BSA), pH 7.5; 5% *E. coli* lysate and 3% CKS lysate overnight at 4°C. The strips were washed with buffered detergent (10 mM phosphate buffered saline (PBS) pH 7.5, containing 0.1% SDS and 0.5% Triton X-100®), followed by addition of goat anti-mouse IgG antibody conjugated 35 to HRPO. The strips were incubated for one to two hours at room temperature, followed by washing with buffered detergent. Finally, antibody bound to the protein was

visualized by addition of freshly prepared HRP color reagent (Biorad Laboratories, Richmond, CA) (120 mg dissolved in 40 ml ice-cold methanol, then diluted into 200 ml Tris buffered saline, pH 7.8, containing 120 µl of 30% hydrogen peroxide). This assay demonstrated the presence of antibody to the respective proteins with which the mice had
5 been immunized.

EXAMPLE 2

Cell Fusion

Upon demonstration of specific anti-HCV antibody present at reasonable titers in
10 sera of immunized mice, the mice were allowed to rest for at least eight weeks prior to a pre-fusion boost of antigen. The pre-fusion antigen boost then was performed by intravenous (IV) tail vein injection of approximately 40 µg of respective purified recombinant HCV protein. Three days later the mice were sacrificed, and their spleens which contained anti-HCV antibody-producing cells were disrupted into single cells.
15 These single cell suspensions were treated with 0.83% NH₄Cl to remove red blood cells, and then these suspensions were mixed with SP2/0 cells at a 10:1 (SP2/0:spleen cells) ratio. The mixed cells were centrifuged, washed once with serum-free medium, and again centrifuged. The fusogen polyethylene glycol (PEG) was used to form hybrids of the immune donor spleen cells with the myeloma cell line SP2/0 (HPRT neg.) Kohler and Milstein, Nature 256:494 (1975), and reviewed in J.G.R. Hurrel, ed., Monoclonal Hybridoma Antibodies: Techniques and Applications, CRC Press, Inc., Boca Raton, FL (1982). Briefly, fusion of the spleen and SP2/0 cells was accomplished by exposing the pellet to 40% PEG (ATTC, MW 1300-1600) in serum-free Iscoe's Modified Dulbecco's Medium (IMDM) for two minutes. The PEG and cell suspension was diluted
20 slowly by the addition of 20 ml of serum-free IMDM over a period of five minutes, followed by collection of the cells by centrifugation. The supernatant was decanted and replaced with 30 ml IMDI containing 20% fetal bovine serum (FBS) (Hyclone Laboratoreis, Logan, Utah) with HAT (hypoxanthene, aminopterin and thymidine) media in order to select for hybridomas. Spleen cells from one non-immune BALB/c mouse also
25 were added as a feeder layer. The cells were plated at 0.1 ml/well in three 96-well tissue culture plates. An additional 0.1 ml of HAT media was added to each well three days later. At weekly intervals thereafter, one-half the media was replaced with IMDM containing 20% FBS with HT (hypoxanthene and thymidine), and hybrids were allowed to grow for an additional seven to fourteen days.
30

It was found that some of the hybrids were composed of spleen cells making antibody to HCV fused with SP2/0 cells. Briefly, the fusogen promoted fusion of spleen cell and SP2/0 cell membranes, which formed a heterokaryon containing nuclei of both cells. Eventually, the dissimilar nuclei fuse produced a single nucleus capable of 5 synchronous mitosis. As the fused cells divided, the hybrid stabilized by losing chromosomes of each nucleus. The fused cells were plated into multiple 96-well plates at 10^5 to 10^6 cells per well. The hybrid cells formed from SP2/0:spleen cell fusions were selectively propagated by culturing in HAT medium. All unused SP2/0 or SP2/0:SP2/0 fused cells were prevented from growing by aminopterin, and unfused 10 spleen cells or spleen:spleen fused cells died off in culture. Only SP2/0:spleen cell hybrids grew in the HAT selection medium.

EXAMPLE 3

Screening and Cloning of Monoclonal Antibodies

15 After 10 to 14 days, culture fluids from wells containing hybridoma cell growth were screened for the presence of a monospecific antibody as follows. Each of the hybridoma culture fluids was tested on a plate coated with the immunogen as well as on a plate coated with CKS protein (fusion partner used for HCV proteins) by the EIA procedure described in Example 1. Hybridoma culture fluids reacting specifically to the immunogen, i. e., HCV protein, and not the CKS fusion partner were selected for further analysis by Western blot analysis. EIA-positive hybridoma culture fluids were tested for their reactivity to the respective HCV proteins as well as CKS by Western blot analysis as described in Example 1. Hybrid samples reacting specifically with the HCV protein but not with the CKS protein by both EIA and Western blot were identified, and 20 selected for cloning by the limiting dilution method, using the guidelines outlined by J. W. Goding, Monoclonal Antibodies: Principles and Practices, Academic Press, New York 25 (1983). Culture supernatant of cloned samples were tested again by EIA with the immunogen and the CKS protein as described above in Example 1, for the confirmation of monospecific reactivity to HCV protein sequence. Clones with strongest reactivity to the 30 protein of choice were selected for expansion and further analysis.

EXAMPLE 4

Amplification of Antibody Yields by Ascites Method

In order to obtain greater amounts of monoclonal antibodies, 10 to 20 million 35 cloned cells of the desired hybridoma cell line were inoculated into a BALB/c mouse previously treated i.p. with 0.5 ml pristane (2,6,10,14-tetramethylpentadecane) by

the method outlined in J. G. R. Hurrell, ed., Monoclonal Hybridoma Antibodies: Techniques and Application, CRC Press, Boca Raton, FL (1982). Pristane treatment enhanced growth of mouse myeloma hybrids within the peritoneum of the mouse, and the ascites fluids which formed were rich in the monoclonal antibody secreted by the hybrid 5 cells. After formation of adequate ascites fluid (approximately seven days), the mice were sacrificed and the ascites were withdrawn from the peritoneum, clarified by centrifugation and stored at -20°C. Monoclonal antibodies from ascites fluid were purified using protein-A sepharose (according to J.G.R. Hurrell et al, supra). All characterization procedures described herein were performed with either culture 10 supernatants, ascites fluids or protein-A purified IgG.

EXAMPLE 5

Characterization of Monoclonal Antibodies

EIA

15 Enzyme-linked immunoassay as described in Example 1 was used to determine the specificity of each of the monoclonal antibodies. Briefly, clarified ascites fluids or protein-A purified IgG were reacted in serial dilutions in microtiter plates coated with either a) the immunogen (i.e., pHCV23, or pHCV39 or pHCV34), b) CKS protein (fusion partners used for cloning and expression of all three immunogens), and c) respective 20 proteins (i.e. C-100, 33C or CORE) expressed in E. coli under the control of bacteriophage λpL promoter (i.e. HCV proteins expressed without CKS fusion partner). Specificity of each of the monoclonal antibodies for the respective protein was confirmed by specific activity of the monoclonal antibody to the immunogen as well as the HCV protein expressed in E. coli λpL but not with CKS protein. TABLE 1 illustrates the 25 representative data for monoclonal antibodies of the invention to C-100, 33C and CORE proteins of HCV.

Western blot analysis

The general protocol for Western blot analysis is as described in Example 1, 30 except that CKS lysate was excluded from the dilution buffer. Briefly, approximately 300 μg of either a) the immunogen (i.e. either pHCV 23, pHCV 29 or pHCV 34), or b) CKS protein (fusion partner for immunogen) or c) the respective proteins expressed in E. coli under the control of λpL promoter, were electrophoresed and transferred to the nitrocellulose. After blocking free sites on nitrocellulose, 2 mm side strips were cut. 35 Each of the monoclonal antibodies was tested for reactivity against all three antigens (i.e., the immunogens, CKS and the respective HCV proteins expressed in E. coli λpL).

Specificity of each of the monoclonal antibodies was confirmed as described for EIA analysis. Representative data are illustrated in FIGS. 2A to 2F. Referring to FIGS. 2A to 2F, the photographs show mono-specific binding of each monoclonal antibody of the invention to its specific protein.

5

Isotype

The isotype of each of the monoclonal antibodies was determined by using an isotyping kit (Amersham, Arlington Heights, IL) and following the instructions included with it. Briefly, the tissue culture supernatant of each monoclonal antibody and appropriate controls were reacted at a 1:5 dilution with strips coated with specific anti-isotype antibody, provided in the kit described above. Assay protocol was followed exactly according to the manufacturer's instructions. The isotype of each monoclonal antibody of the invention is provided in TABLE 1.

15 Competition With Immune Human Sera

In order to establish whether each of the monoclonal antibodies recognized an epitope that is immunogenic in humans, a competition assay was performed as follows. Each of the monoclonal antibodies was tested in an assay where the monoclonal antibody competed with a human sera seropositive for antibody to C-100, 33C and CORE for the binding to the respective antigen. Briefly, a human serum from an individual infected with NANBH and strongly seropositive for antibodies to C-100, 33C and CORE proteins of HCV was included in the reaction mixture with each of the monoclonal antibodies at a final concentration of 10%. Microtiter EIA was carried out as described in Example 1. A greater than 50% inhibition in the binding of the monoclonal antibody to the respective protein by the immune human sera was considered as competitive (data presented in TABLE 1).

EXAMPLE 6

Epitope Mapping

30 Monoclonal antibodies to HCV proteins C-100, 33C and CORE were mapped to the specific region of the protein by (a) Western blot reactivity of each of the monoclonal antibodies with subfragments of the respective HCV proteins and (b) reactivity with several synthetic peptides selected for respective protein sequences, by microtiterEIA. In addition to these two methods, monoclonal antibodies to C-100 also were mapped by 35 PEPSCAN analysis, in order to further define the epitope recognized by these antibodies.

Specific additional details for mapping will be detailed where applicable for an individual monoclonal antibody.

Reactivity of Monoclonals to Various Subfragments of Recombinant HCV proteins

5 Briefly, several individual oligonucleotides representing a.a. 1676-1931 of HCV genome were ligated and cloned as three separate EcoRI-BamHI subfragments into the CKS fusion vector pJ0200. These three subfragments were designated as CKS-B (a.a. 1676-1790), CKS-C (a.a. 1789-1863) and CKS-D (a.a. 1861-1931) as described in FIG.1.. The Western blots of selected monoclonal antibodies of the invention are shown in
10 FIGS. 2-2F. The detailed methods for cloning and expression of the CKS-fusion proteins are as disclosed in U. S. Patent Application Serial No. 07/572,822, which enjoys common ownership and is incorporated herein by reference. Cell lysates of these clones were used as antigens on Western blot analysis for preliminary epitope mapping of anti-C-100 monoclonal antibodies. Similarly, two subfragments from HCV 22C regions
15 designated as 33C A-CKS (a.a. 1192-1331) and 33CB-CKS (a.a. 1330-1457) also were cloned and expressed in E. coli as described above for FIG. 1. These lysates were used as antigens for epitope mapping of anti-33C monoclonal antibodies.

20 Western blot analysis of each of the monoclonal antibodies with appropriate sets of subfragments of control proteins (full length proteins as well as CKS fusion partner) was carried out as described in Example 1, except that CKS lysate was excluded from the dilution buffer. Data for epitope mapping with these recombinant subfragments are illustrated in FIGS. 2A to 2F. Monoclonal antibody H28C110 showed reactivity with C-100B, pHCV23 and C-100 (λ pL construct) but did not react with C-
25 100C, C-100D or CKS protein, which indicated that H28C110 specifically recognizes or binds an epitope between a.a. 1676-1790 of the HCV genome. Similarly, monoclonal antibody H4C20 recognizes (specifically binds) an epitope between a.a. 1861-1931. Based on these data, monoclonal antibody H11C130 recognizes an epitope between a.a.
30 1192-1331, and monoclonal antibody H1C46 recognizes (specifically binds) an epitope between a.a. 1330-1457.

Reactivity with Synthetic Peptides

Several amino acid sequences were selected from different regions of HCV proteins C-100, 33C and CORE. A list of the peptides used for the epitope mapping of
35 these monoclonal antibodies is listed below in TABLE 2.

TABLE 2
Epitope Mapping With Synthetic Peptides

5	REGION OF HCV GENOME	MONOCLONAL TESTED	PEPTIDE a.a.	REACTIVITY OF EACH WITH PEPTIDE
10	CORE	H81C17 H35C54	sp 1-75 sp 35-75	None None
15	33C	H1C46 H11C130	sp 1192-1240 sp 1223-1240 sp 1357-1407 sp 1418-1457	None None
20	C-100	H28C110	PEPSCAN analysis (a.a. 1694-1735) sp 1694-1750 sp 1684-1735 sp 1696-1708 sp 1866-1930	sp 1694-1750 and sp 1684-1735; a.a. 1702-1709 (PEPSCAN)
25		H4C20	sp 1899-1930	sp 1899-1930

Each of these peptides were assembled on a resin support by a stepwise solid phase synthesis, starting with the carboxy terminal residue. A procedure was employed similar to that described in E. Gross and T. Heinehofer, eds., Barany and Merrifield, The Peptides 2:1284, Academic Press, New York, New York (1980), using a reaction vessel of an Applied Biosystems Synthesizer Model 430A. After cleavage of the peptide from the resin, the peptide was washed with diethyl ether and extracted in 40% acetic acid solution. Crude peptide obtained after lyophilization of the aqueous solution was employed as the antigen target for epitope mapping experiments. Briefly, each of the peptides tested was coated on microtiter wells at a concentration of 10 μ g/ml in bicarbonate buffer at pH 9.5. EIA was performed in the manner described in Example 1. Monoclonal antibody showing reactivity four times the negative control was considered positive.

In addition, monoclonal antibodies to HCV C-100 were also mapped with PEPSCAN analysis. Synthetic peptides were synthesized on polypropylene pins following the manufacturer's instructions (Cambridge Research Bioscience, Valley Stream, New York). An EIA was performed with each of the monoclonal antibodies to HCV C-100 by the procedure outlined by the manufacturer with 67 overlapping hexamer peptides.

encompassing a.a. 1684-1750 of the HCV genome. Representative data are illustrated in FIG. 4. Monoclonal antibody H28C110 specifically reacted with peptide sequence Tyr-Arg-Glu-Phe-Asp-Glu-Met-Glu (a.a. 1702-1709 of the HCV genome). Monoclonal antibody H4C20 did not show reactivity in PEPSCAN but did react in EIA with a larger peptide 1899-1930 a.a. This could be due to the fact that H4C20 requires a longer peptide to recognize a linear epitope.

EXAMPLE 7

EIA for the Detection of HCV Proteins in Biological Samples

10 For the purposes of simplification, detailed methods are described for the CORE proteins of HCV. Methods described for the preparation of rabbit polyclonal antibodies to HCV proteins 33C and C-100 were identical to that detailed hereinbelow for HCV CORE protein.

15 Preparation of Rabbit Polyclonal Antibodies Against HCV CORE Protein

Young rabbits (3-4 months old and weighing 2-3 kg) were obtained from Hazelton Labs, Denver, PA. Primary immunization was carried out with 100-150 µg of highly purified HCV CORE protein (expressed in *E. coli* under λpL promoter) in Freund's complete adjuvant by intra-muscular (I.M.) injection at four different sites. 20 Subsequently, two immunizations were carried out at two week intervals in similar fashion in Freund's incomplete adjuvant. Immune response of the rabbits was monitored by EIA and Western blot analysis as described in Example 1. Rabbits were bled when acceptable immune response to the protein was achieved. IgG from the immune rabbit sera was purified by Protein-A sepharose affinity chromatography, by methods known to those in the art. 25

Bead Coating

In the presently most preferred assay format, rabbit IgG prepared as hereinabove was coated on polystyrene beads as the solid support for capture of CORE 30 antigens in the test sample. The polystyrene beads were washed with distilled water and incubated at 40°C for two (2) hours with 5-10 µg/ml of purified HCV CORE rabbit IgG in a buffer solution (0.1M Tris, 0.5M NaCl, 0.0022% Triton X-100®, pH 8.5). The beads were washed once with PBS and then soaked in 0.1% Triton X-100® in PBS for approximately one (1) hour at 40°C. After washing twice with PBS, the beads were 35 overcoated with 3% bovine serum albumin (BSA) in PBS for approximately one (1) hour at 40°C. Finally, the beads were overcoated with a 5% sucrose solution in PBS and

dried under nitrogen. Anti-HCV human polyclonal IgG, purified from sera of individuals seropositive for HCV antibodies to C-100, 33C and CORE also was coated in similar fashion.

5 EIA

Several monoclonal antibodies specific for either HCV C-100, HCV 33C or HCV CORE were screened for use as the probe for detection of HCV proteins in a test sample by EIA. Briefly, each of the monoclonal antibodies was incubated with the respective antigen in the presence of polystyrene beads coated with anti-HCV rabbit polyclonal IgG.

10 The detailed protocol for EIA was similar to that described hereinbelow. Data illustrating the reactivity of the monoclonal antibodies which showed the best reactivity as a probe for the antigen detection assay are shown in TABLE 3.

15 TABLE 3

	Monoclonal Antibody	Antigen	Absorbance at A492 at antigen conc.	
			NEG CONTROL 0 ng/ml	POS. CONTROL 100 ng/ml
20	H81C17	HCV CORE	0.246	>6.62
	H11C130	HCV 33C	0.194	0.832
	H28C110	HCV C-100	0.623	2.53

25 In the EIA procedure, 200 µl of specimen which was suspected of containing antigen to HCV CORE protein was incubated in a reaction tray with 50 µl of monoclonal antibody H81C17 (at a final protein concentration of 5-10 µg/ml diluted in a buffer containing 20 mM Tris, 0.1 mM NaCl, 1 mM EDTA, 3.0% BSA, 0.3% Tween-20® and 10% FBS at pH 7.5), and a bead coated with HCV rabbit IgG (prepared as described hereinabove). After overnight incubation at ambient room temperature, the beads were washed with distilled water and 200 µl of appropriately diluted Horseradish peroxidase labeled goat anti-mouse IgG (H + L) (Jackson Immunoresearch, West Grove, PA) was added. Incubation with the labeled probe was carried out at about 40°C for approximately two (2) hours. Beads were washed and transferred to reaction tubes containing 300 µl of O-phenylenediamine:2HCl (OPD) color reagent. The reaction was carried out at ambient room temperature in the dark for 30 minutes, and then it was stopped by the addition of 1 ml of 1N H₂SO₄. Absorbance was recorded at 492/600 nm. A negative control which was previously screened and confirmed to be negative for

NANBH infection was included in the experiment. The positive control consisted of a solution of recombinant HCV CORE protein (pHCV 34) in the buffer solution described hereinabove. Triplicates of both positive and negative control were included with each set of experiments.

5

In order to determine the efficiency of the antigen capture assay for the detection of HCV CORE protein in a sample, various concentrations of recombinant HCV CORE protein (expressed in *E. coli* under λ pL promoter), ranging from 100 ng protein/ml to 100 pg protein/ml were diluted in the buffer mentioned above. The EIA procedure was performed with each of diluted the diluted panel members following the procedure described herein. For the purpose of comparison; each of the panel members was tested with (a) anti-HCV rabbit polyclonal antibody on the solid phase and (b) anti-HCV human polyclonal antibody on the solid phase. As illustrated in FIG. 4 and TABLE 4, the antigen capture assay of the present invention detected as low as 200 pg HCV CORE protein in 200 μ l of the test sample. Although there was no significant difference in the sensitivity of detection with either rabbit or human capture antibody, anti-HCV rabbit antibody showed overall superior performance in the assay and was chosen as the preferred polyclonal antibody for the antigen capture.

20

TABLE 4.
Detection of HCV CORE Protein by Monoclonal Antibody H81C17

	CONC. OF ANTIGEN pL CORE* ng/ml	REACTIVITY IN EIA	
		A492 Rabbit Bead	A492 Human Bead
25	100	6.39	4.45
	50	4.06	2.19
30	25	2.19	1.25
	12.5	1.02	0.887
	6.25	0.339	0.382
	3.125	0.230	0.121
35	1.56	0.084	0.043
	0.78	0.050	0.032
	0.39	0.038	0.014
	0.2	0.028	0.015
	0.1	0.020	0.014
40	0	0.014	0.014

* Recombinant HCV CORE expressed in *E. coli* under λ pL promoter system used as the antigen. Concentrations represent ng of protein/ml based on A280 values. The assay

system has 200 µl/well for testing, so the actual amount of antigen per well is one-fifth of the value given here.

EXAMPLE 8

5

Testing of Negative Serum and Plasma Samples

Sixty sera and sixty plasma samples negative for NANBH were tested by the EIA procedure detailed in Example 7. The results of these assays are presented in FIG. 5. It can be seen by the data that the majority of negative plasma and sera fell within a tight range of O.D. (Optical Density) values close to the negative control value. One plasma 10 sample with an O.D. of 0.1725 was found to be repeat reactive. Confirmation of the presence of antigen could not be performed on this sample due to the insufficient volume of the sample..

EXAMPLE 9

15

Production and Use of Cell Lines 1 3-975-157, 14-153-234 and 14-1350-210

A. Production of recombinant HCV antigens and immunogens

Synthetic peptides corresponding to regions within the putative CORE domains of the HCV genome were made by automated peptide synthesizer. The following peptides 20 were constructed utilizing methods known in the art:

CORE 1-75

35-75

35-61

These peptides are described in pending United States patent application Serial 25 No. 07/610,180, entitled HEPATITIS C ASSAY, which enjoys common ownership and is incorporated herein by reference.

Recombinant antigens were made as fusion proteins with CMP-XDO synthetase in E. coli according to methods known in the art, or as non fused protein with λPL promoter 30 system. The following proteins were cloned and purified:

λPL-CORE (1-150)

CKS-CORE (1-150)

CKS-33c (1191-1457)

λPL-33c-CORE (fusion of 1191-1457 and 1-150)

35 CKS-BCD (156 -1930)

CKS-E (1931-2189) (NS4/NS5 reaction)

CKS-B (1676-1790)

See FIG. 1B for map of HCV genome and approximate locations of HCV regions.

Recombinant protein C-100 (1569-1930) was obtained from Chiron as a fusion

5 protein with superoxide dismutase (SOD). All recombinant proteins were greater than 90% pure by SDS-PAGE.

B. Immunization of Mice

BALB/c mice (Charles River Laboratories, Charles River, NY), 6-8 weeks old, 10 were initially immunized subcutaneously and intraperitoneally with 50 µg of λPL-CORE in 100 µl of Freund's complete adjuvant (Difco, Detroit, MI). On day 15, 50 µg of the immunogen was diluted into 100 µl of phosphate buffered saline (PBS), pH 7.2, and injected intravenously into the tail vein (J. Goding, Monoclonal Antibodies: Principles and Practice [New York; Academic Press, 1986]). Sera titers were not evaluated.

15 **C. Fusion**

On day 18, mice were sacrificed and splenocytes were fused in a 1:1 ratio with the SP2/0 myeloma line according to known conventional methods (G. Kohler and C. Milstein, Nature (1975) 256:495-497; J. Goding, *supra*). The cell fusion pellet was dispersed with 1 ml 50% polyethylene glycol (PEG) (American Type Culture Collection, MW 1450) and centrifuged in Iscove's Modified Dulbecco's Medium (IMDM) (Gibco, Grand Island, NY). The cells were resuspended in HAT (hypoxanthine-aminopterin-thymidine)-selective IMDM with 10% fetal bovine serum (FBS) (Hyclone Laboratories, Logan, UT) and plated at 3×10^5 cells per 96-well tissue culture plates.

20 25 Growth promoters included in the HAT media were 0.5% STM (RIBI Immunochem Research, Inc., Hamilton, MT) and 1% Origen Hybridoma Cloning Factor (Igen, Rockville, MD). Growth medium was replaced in culture wells post-fusion on day 5 and 7 using HT (hypoxanthine-thymidine) supplemented IMDM with 10% FBS.

30 **D. Enzyme Immunoassay (EIA)**

Culture supernates were EIA screened 10 days post-fusion against the immunizing antigen to detect hybrids secreting HCV specific antibody and a non-specific protein to eliminate any false positives (Langone & Van Yunakis, eds., Methods in Enzymology, 92:168-174, Academic Press [1983]). Polystyrene 96-well microtiter

35 plates were coated overnight at room temperature with 50 µl per well of a 1 µg/ml HCV antigen solution in PBS. Any remaining binding sites on the polystyrene wells were

blocked with 3% bovine serum albumin (BSA) (Intergen, Purchase, NY) in PBS for 30 minutes at room temperature. Plates were washed three times with distilled water. Fifty microliters of hybridoma tissue culture supernatants were incubated for 1 hour at room temperature in the wells, and the wells were washed three times with distilled water. Antibody binding to antigen was detected using goat anti-mouse IgG+M-horseradish peroxidase (HRPO) (Kirkegaard-Perry Laboratories [KPL], Gaithersburg, MD) diluted at a concentration of 1:1000 in the block solution and incubated 30 minutes at room temperature. The plates were washed with distilled water and o-phenylenediamine substrate (OPD; Abbott Laboratories, Abbott Park, IL) was used as the chromogen. Plates were read at 492 nm. Hybrid cultures were regarded as potential HCV antibody-positive when the optical density (OD) was 3 times the negative control (NC) and significant preferential to the HCV antigen plate was observed compared to antibody binding of the irrelevant antigen coated plate, i.e.: >0.2 OD difference and <0.2 OD signal on the latter.

15

E. Western Blot

Hybrid antibody specificity was confirmed with Western blot analysis (Towbin & Gordon, *J. Immunol. Methods*, 72:313-340 [1984]). HCV recombinant proteins and irrelevant proteins were electrophoresed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and then transferred to nitrocellulose, according to the manufacturer's instructions (Schleicher & Schuell, Keene, NH; Bio-Rad, Richmond, CA). The nitrocellulose strips were blocked with 1% bovine hemoglobin (Sigma Chemical Co., St. Louis, MO) and 0.5% Tween-20 (Fisher Scientific, Pittsburgh, PA) in PBS for 30 minutes at room temperature, then the strips were incubated with hybrid tissue culture supernatant. The strips were then washed in PBS and goat anti-mouse IgG+M-HRPO (KPL) added for 30 minutes. Antibody binding to the HCV antigen was visualized with 4-chloro-1-naphthol (Sigma) as the chromogenic substrate. Hybrid cultures were cloned and placed in cryostorage if HCV antibody specificity was demonstrated.

30

F. Establishment of Clones

HCV specific hybrids were cloned by limiting dilution (Goding, *Monoclonal Antibodies: Principles and Practices*, 2nd ed, Academic Press, New York [1986]). Modifications included plating of the cultures in log₁₀ dilution series and selecting positive clones for expansion from plates which exhibit <20% growth per 96 well tissue culture plate. Culture supernates were tested after 10 days using the EIA and

Western blot procedures described above. The selected clones were expanded for further evaluation and cryostoraged in 80% IMDM with 10% FBS and 10% DMSO (Sigma).

5 G. Monoclonal Antibody Isotype

Monoclonal antibody isotype was determined with the SBA Clonotyping System III kit (Southern Biotechnology Associates, Inc., Birmingham, AL) with modifications. EIA 96-well microtiter plates were coated overnight at room temperature with 100 µl/well of a 1:1000 dilution of goat anti-mouse IgG+M (H+L) (KPL). Plates were 10 blocked for 30 minutes with 3% BSA in PBS and washed with water. Culture samples were added to the wells, incubated for 1 hour, and washed with water. The kit's goat anti-mouse subtype specific conjugates were added for a 30 minute incubation period. Following a water wash, color was identified with OPD substrate. The goat anti-mouse isotype specific conjugate that bound to the mouse immunoglobulin and displayed a >0.1 OD 15 at 492 nm signaled the subtype.

H. Monoclonal Antibody Production

Clones selected for further evaluation were scaled up in tissue culture T-flasks and 10⁶ cells were injected into the peritoneal cavity of pre-pristaned BALB/c mice 20 (Charles River Biotechnical Services, Inc., Wilmington, MA) (see Hurrell, supra). The resulting ascites fluid was harvested 7-10 days after injection, centrifuged, and stored at -20°C. The IgG antibody was affinity purified on Protein A (Pharmacia-LKB Biotechnologies, Piscataway, NJ) utilizing the automated OROS purification system Model 100 (see Goding, supra, for basic principles). The IgM antibodies were purified 25 by molecular sizing on a S-300 column (Pharmacia-LKB).

All the following characterization information was performed with purified monoclonal antibody.

30 I. Isoelectric Focusing (IEF)

A cell line quality control to ensure consistency of frozen lots included measuring the antibody pI point on an IEF gel apparatus (Bio-Rad) which separates proteins based on net charge. Briefly, a bis-acrylamide-riboflavin solution was applied to an acrylamide gel, exposed to fluorescent lighting for 1 hour, then stored overnight at 4°C. 35 A 1 µg sample of monoclonal antibody and standards were laid upon the gel and electrophoresed over a 1-2 hour period. Following a series of fixatives and washes, the

gel was silver stained (Bio-Rad). The pl value of the monoclonal antibody was calculated by migratory distance through the gel and was directly compared to the protein standards' migratory distance of known pl values. The distinctive finger print banding pattern reflected the pl microheterogeneity between independently produced lots of antibody (Hamilton, R.G., Reimer, C.B., Rodkey, L.S. (1987) Quality control of murine monoclonal antibodies using isoelectric focusing affinity immunoblot analysis. Hybridoma 6:205-217).

J. EIA and Western Blot Specificity of Monoclonal Antibodies

10 All monoclonal antibodies noted herein were screened on an assortment of available recombinant HCV antigens as disclosed in U. S. Patent Application Serial No. 07/572,822 entitled HEPATITIS C ASSAY UTILIZING RECOMBINANT PROTEINS, which enjoys common ownership and is incorporated herein by reference. The procedures are outlined above. The multiple antigen screening technique confirmed the HCV specificity
15 and to exclude the HCV non-specific CKS, λ PL, or linker-arm reactivity of the monoclonal antibodies.

K. EIA Epitope Competition Studies

To investigate specificity and antigen binding distinctions, epitope grouping experiments were performed utilizing biotin labeled and unlabeled monoclonal antibodies (Langone & Van Vunakis, Methods in Enzymology, 92:242-253, Academic Press [1983]). Briefly, the antibodies were labeled with NHS-LC-biotin (Pierce Chemical Co., Rockford, IL) according to the manufacturer's instructions. Microtiter wells were coated with the immunogen as previously described. First, log₂ dilutions of
25 the unlabeled antibody were pre-incubated in the wells for 15 minutes, followed by the addition of a fixed amount of biotinylated antibody (the dilution in a direct EIA of the biotinylated antibody alone which gave a value of 50% of the maximum absorbance value) and incubated for 20 minutes. Plates were washed three times with water.
Diluted streptavidin-HRPO (Zymed, South San Francisco, CA) was added to the wells and
30 incubated for 30 minutes. The plates were washed again and OPD color developed as previously described. The absorbance was read at 492 nm. Antibodies of the same or related epitope had signal blocked or inhibited by >50%. No inhibition was observed with antibodies of distinct specificity. This was performed reciprocally for antibodies produced within HCV core region.

L. RIA Reciprocal Competition

Beads coated with the appropriate antigen or peptide were incubated with 100 µl of unlabeled monoclonal antibody diluted into recalcified negative human plasma (NHP, testing negative for anti-HCV, anti-HIV and HBsAg) at monoclonal antibody

5 concentrations of 1-20 µg/ml. 100 µl of radiolabeled antibody at 1 to 4 µCi/ml diluted into HTLV I kit specimen diluent (containing detergent, animal sera, buffer) was incubated with the bead for 2 hours at 45°C or 18-20 hours at 20-25°C. Beads were washed and counted for radioactivity.

10 M. EIA Reactivity with Synthetic Peptides

Beads coated with 3 mg/ml were incubated with 100 ml of monoclonal antibody at concentrations of 0.02 -1 mg/ml for 1 hour. The beads were washed and goat anti-mouse IgG+M-HRPO (KPL) was added for 30 minutes. The beads were washed again and OPD (Abbott Labs) was used as the chromagen for O.D. readings at 492 nm.

15

N. HCV Antigen Assays

Beads coated with one or a cocktail of anti-HCV monoclonal antibody were incubated with 200 µl of specimen for 2 hours at 40-45°C or 18-20 hours at 20-25°C. Beads were washed with distilled water and then incubated with 200 µl of 20 radiolabeled anti-HCV monoclonal antibody (one or more) for 2 hours at 45°C. Beads were washed and counted in a gamma counter.

O. Characterization of Monoclonal Antibody

Monoclonal antibody against the HCV CORE domain (1-75) fell into two distinct groups based on reciprocal competitive studies. All groups reacted with CKS-CORE (1-25 150) and IPL-core, IPL-33c-core and synthetic peptides corresponding to (1-75). Group 1 monoclonal antibody 13-975-157 strongly reacted with peptide (1-75), somewhat to (35-75), and not to (35-61). Group 2 monoclonal antibody 14-153-30 234 clearly reacted to peptides (1-75) and (35-75), but not to (35-61). Unlike the other groups, monoclonal antibody 14-1350-210, reacted strongly to all 3 of the HCV core synthetic peptides. These data are shown in Tables 5 and 6. Referring to FIGS. 6 to 35 17, the reactivities summarized below in Table 5 are shown in lanes 4, 5 and 6. Lanes 1 to 3 contains monoclonal antibodies against HCV 33C protein (6-296-534 in lane 1, 6-914-518 in lane 2 and 6-1070-110 in lane 3); lanes 4-6 contain monoclonal antibodies against HCV CORE (13-975-157 in lane 4, 14-153-234 in lane 5 and 14-1350-210 in lane 6); lanes 7 and 8 contain monoclonal antibodies against the putative

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TABLE 7
Competitive One step anti-CORE assay

		<u>Label: 14-153-234</u>					<u>Label: 14-1350-210</u>				
	Specimen	CPM	Avg	S/N	% Inhibition	Result	CPM	Avg	S/N	% Inhibition	Result
5	NC	48272	48542				36866	34521			
		47010					35655				
		50345					31041				
10	Elevated ALT										
	27	4845	5130	0.11	89.4	+	440	405	0.01	98.8	+
		5414					370				
	238-NC	40035	41964	0.86	13.6	-	28481	28294	0.82	18.0	-
		43892					28106				
15	135	620	558	0.01	98.9	+	114	117	0.00	99.7	+
		496					119				
	163	14540	15471	0.32	68.1	+	100	102	0.00	99.6	+
		16402					104				
20	173	5344	5531	0.11	88.6	+	114	126	0.00	99.6	+
		5717					137				
	220	1049	1012	0.02	97.6	+	124	130	0.00	99.6	+
		974					136				
	252	231	212	0.00	99.6	+	138	135	0.00	99.6	+
		192					132				
25	R13203	56075	55778	1.15	-14.9	-	3029	3479	0.10	89.9	+
		55480					3929				
	SAC190	472	417	0.01	99.1	+	102	194	0.01	99.4	+
		362					284				
30	7088396	41692	41963	0.86	13.6	-	102	122	0.00	99.6	+
		42233					142				
	EP10968	39418	40470	0.83	16.6	-	94	95	0.00	99.7	+
		41522					95				
	14-153	5580	5491	0.11	88.7	+	16216	14178	0.41	58.9	+
		5402					12140				
35	283-NC	45170	44219	0.91	8.9	-	24748	23908	0.69	30.7	-
		43268					23067				

40

EXAMPLE 11
Anti-HCV CORE Two-Step Blocking Assay

A two step blocking assay performed according to a competitive one-step assay described herein for "RIA Reciprocal Competition" for detection of anti-HCV CORE, using labeled group 3 monoclonal antibody (14-1350-210), is shown in Table 8. Sixteen specimens which were repeat reactive for anti-C100 (using the Ortho 1.0 gen kit, available from Ortho Diagnostics, Raritan, New Jersey) were tested. Six had been

shown to be false positive by an anti-C100 confirmatory assay and 10 were confirmed as positive. None of the six false positives gave greater than 20% inhibition in the assay compared to 48-99% inhibition observed for true anti-C100 positives in this anti-CORE assay.

5

Table 8
Two Step Blocking Anti-HCV-CORE Assay

Bead: 1.0 µg λPL-CORE; Label: 14-1350 in HTLV I diluent

	Specimen	CPM	Avg	S/N	% Inhibition	Result
	NC	23376	25969			
		26257				
		28275				
10	Sacramento Negative 1.0 Gen.					
	183	31956	1.23	-23.1		
	184	26525	1.02	-2.1		
	185	24714	0.95	4.8		
	192	20664	0.80	20.4		
	193	20934	0.81	19.4		
	194	23321	0.90	10.2		
15	Sacramento Positive 1.0 Gen.					
	188	13569	0.52	47.7	+	
	189	2243	0.09	91.4	+	
	191	2361	0.09	90.9	+	
	195	284	0.01	98.9	+	
	196	170	0.01	99.3	+	
	197	487	0.02	98.1	+	
20	206	3639	0.14	86.0	+	
	207	2713	0.10	89.6	+	
	208	145	0.01	99.4	+	
	214	10398	0.40	60.0	+	
25						
30						
35						

EXAMPLE 12

HCV CORE Ag Assay

Results from one type of CORE Ag assay is shown in Table 9. This assay was performed as a 2 step assay according to the procedure outlined hereinabove in "HCV Antigen Assay" and as follows: The first incubation with specimen (200 µl) was 18 hours at room temperature, followed by 2 hr. incubation at 45°C with radiolabeled group 2 monoclonal antibody 14-153. Since the bead was coated with a group 3 monoclonal antibody 14-1350 which does not compete with group 2, a one step sandwich assay was also possible.

Specimens giving S/N values greater than 3.0 were considered reactive for CORE Ag. Sensitivity of the assay with recombinant λPL-CORE was about 100 ng/ml. Two of

13 specimens from patients with elevated ALT and anti-C-100 reactivity were considered reactive for CORE antigen.

Table 9

HCV CORE Ag Assay

(14-1350 bead/14-153 label)

	Specimen NC	CPM	Average	S/N	Result
10		114	128		
		137			
		133			
15	λPL-CORE	110 µg/ml	1846	1822	14.23 +
			1798		
			1488	1447	11.30 +
			1405		
		1.1 µg/ml	1220	1246	9.73 +
			1271		
		110 ng/ml	514	472	3.69 +
			430		
20		11 ng/ml	182	175	1.36 -
			167		
Elevated ALT	10	206	212	1.65 -	
		217			
25		27	160	174	1.36 -
			187		
		54	710	628	4.91 +
			546		
		77	454	458	3.57 +
			461		
30		135	160	189	1.48 -
			218		
		163	323	328	2.56 -
			332		
35		173	346	342	2.67 -
			337		
		220	274	275	2.14 -
			275		
		238	157	171	1.34 -
			185		
40		252	227	216	1.68 -
			204		
		283	194	175	1.36 -
			155		
45		28	258	226	1.77 -
			194		
		290	162	175	1.36 -
			187		

EXAMPLE 13HCV Antibody Test Employing Cocktails of Monoclonal Antibodies

Also performed was a similar CORE Antigen format using a cocktail of anti-CORE monoclonal antibody on the solid phase and in label (14-1350-210, 14-153-234, and 14-726). In twenty five anti-C-100 repeat reactive specimens, one specimen (SAC 5 161) was significantly elevated in reactivity. A frequency histogram of a negative anti-HCV population from the Interstate Blood Bank was prepared (not shown). A cutoff set at S/N=2.0 gave 5 standard deviations from the negative population mean. No negative specimen gave S/N > 1.6.

10

EXAMPLE 14Production and use of cell lines 16-296-534, 6-914-518 and 6-1070-110A. Production of recombinant HCV antigens and immunogens

Recombinant antigens were made as fusion proteins with CMP-XDO synthetase in 15 E. coli according to methods known to those in the art, or as non fused protein with λ PL promoter system. The following proteins were cloned and purified:

16 λ PL-CORE (1-150)

CKS-CORE (1-150)

CKS-33c (1191-1457)

20 λ PL-33c-CORE (fusion of 1191-1457 and 1-150)

CKS-BCD (156 -1930)

CKS-E (1931-2189) (NS4/NS5 reaction)

CKS-B (1676-1790)

25 See FIG. 1B for map of HCV genome and approximate locations of HCV regions.

Recombinant protein C-100 (1569-1930) was obtained from Chiron as a fusion protein with superoxide dismutase (SOD). All recombinant proteins were greater than 90% pure by SDS-PAGE.

30 B. Immunization of Mice

BALB/c mice (Charles River Laboratories, Charles River, NY), 6-8 weeks old, were initially immunized subcutaneously and intraperitoneally with 10-100 μ g of the HCV CKS-33C in 100 μ l of Freund's complete adjuvant (Difco, Detroit, MI). On day 14, a second similar boost was administered with the immunogen emulsified in incomplete 35 Freund's adjuvant (Difco). On day 25, 10-100 μ g of the immunogen was diluted into 100 μ l of phosphate buffered saline (PBS), pH 7.2, and injected intravenously into the

tail vein (J. Goding, Monoclonal Antibodies: Principles and Practice [New York; Academic Press, 1986]). Sera titers were not evaluated.

C. Fusion

5 On day 28, mice were sacrificed and splenocytes were fused in a 1:1 ratio with the SP2/0 myeloma line according to known conventional methods (G. Kohler and C. Milstein, Nature (1975) 256:495-497; J. Goding, *supra*). The cell fusion pellet was dispersed with 1 ml 50% polyethylene glycol (PEG) (American Type Culture Collection, MW 1450) and centrifuged in Iscove's Modified Dulbecco's Medium (IMDM) (Gibco, 10 Grand Island, NY). The cells were resuspended in HAT (hypoxanthine-aminopterin-thymidine)-selective IMDM with 10% fetal bovine serum (FBS) (Hyclone Laboratories, Logan, UT) and plated at 3×10^5 cells per 96-well tissue culture plates. Growth promoters included in the HAT media were 0.5% STM (RIBI Immunochem Research, Inc., Hamilton, MT) and 1% Origen Hybridoma Cloning Factor (Igen, 15 Rockville, MD). Growth medium was replaced in culture wells post-fusion on day 5 and 7 using HT (hypoxanthine-thymidine) supplemented IMDM with 10% FBS.

D. Enzyme Immunoassay (EIA)

Culture supernates were EIA screened 10 days post-fusion against the 20 immunizing antigen to detect hybrids secreting HCV specific antibody and a non-specific protein to eliminate any false positives (Langone & Van Vunakis, eds., Methods in Enzymology, 92:168-174, Academic Press [1983]). Polystyrene 96-well microtiter plates were coated overnight at room temperature with 100 μ l per well of a 1 μ g/ml HCV antigen solution in PBS. Any remaining binding sites on the polystyrene wells were 25 blocked with 3% bovine serum albumin (BSA) (Intergen, Purchase, NY) in PBS for 30 minutes at room temperature. Plates were washed three times with distilled water. Fifty microliters of hybridoma tissue culture supernatants were incubated for 1 hour at room temperature in the wells, and the wells were washed three times with distilled water. Antibody binding to antigen was detected using goat anti-mouse IgG+M- 30 horseradish peroxidase (HRPO) (Kirkegaard-Perry Laboratories [KPL], Gaithersburg, MD) diluted at a concentration of 1:1000 in the block solution and incubated 30 minutes at room temperature. The plates were washed with distilled water and o-phenylenediamine substrate (OPD; Abbott Laboratories, Abbott Park, IL) was used as the chromogen. Plates were read at 492 nm. Hybrid cultures were regarded as 35 potential HCV antibody-positive when the optical density (OD) was 3 times the negative control (NC) and significant preferential to the HCV antigen plate was observed.

compared to antibody binding of the irrelevant antigen coated plate, i.e.: >0.2 OD difference and <0.2 OD signal on the latter.

E. Western Blot

5 Hybrid antibody specificity was confirmed with Western blot analysis (Towbin & Gordon, *J. Immunol. Methods*, 72:313-340 [1984]). HCV recombinant proteins and irrelevant proteins were electrophoresed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and then transferred to nitrocellulose, according to the manufacturer's instructions (Schleicher & Schuell, Keene, NH; Bio-Rad, Richmond, CA). The nitrocellulose strips were blocked with 1% bovine hemoglobin (Sigma Chemical Co., St. Louis, MO) and 0.5% Tween-20 (Fisher Scientific, Pittsburgh, PA) in PBS for 30 minutes at room temperature, then the strips were incubated with hybrid tissue culture supernatant. The strips were then washed in PBS and goat anti-mouse IgG+M-HRPO (KPL) added for 30 minutes. Antibody binding to the HCV antigen was visualized with 4-chloro-1-naphthol (Sigma) as the chromogenic substrate. Hybrid cultures were cloned and placed in cryostorage if HCV antibody specificity was demonstrated.

F. Establishment of Clones

20 HCV specific hybrids were cloned by limiting dilution (Goding, *Monoclonal Antibodies: Principles and Practices*, 2nd ed, Academic Press, New York [1986]). Modifications included plating of the cultures in log₁₀ dilution series and selecting positive clones for expansion from plates which exhibit <20% growth per 96 well tissue culture plate. Culture supernates were tested after 10 days using the EIA and Western blot procedures described above. The selected clones were expanded for further evaluation and cryostoraged in 80% IMDM with 10% FBS and 10% DMSO (Sigma).

G. Monoclonal Antibody Isotype

3.0 Monoclonal antibody isotype was determined with the SBA Clonotyping System III kit (Southern Biotechnology Associates, Inc., Birmingham, AL) with modifications. EIA 96-well microtiter plates were coated overnight at room temperature with 100 µl/well of a 1:1000 dilution of goat anti-mouse IgG+M (H+L) (KPL). Plates were blocked for 30 minutes with 3% BSA in PBS and washed with water. Culture samples were added to the wells, incubated for 1 hour, and washed with water. The kit's goat anti-mouse subtype specific conjugates were added for a 30 minute incubation period.

Following a water wash, color was identified with OPD substrate. The goat anti-mouse isotype specific conjugate that bound to the mouse immunoglobulin and displayed a >0.1 OD at 492 nm signaled the subtype.

5 H. Monoclonal Antibody Production

Clones selected for further evaluation were scaled up in tissue culture T-flasks and 10^6 cells were injected into the peritoneal cavity of pre-pristane BALB/c mice (Charles River Biotechnical Services, Inc., Wilmington, MA) (see Hurrell, supra). The resulting ascites fluid was harvested 7-10 days after injection, centrifuged, and stored 10 at -20°C. The IgG antibody was affinity purified on Protein A (Pharmacia-LKB Biotechnologies, Piscataway, NJ) utilizing the automated OROS purification system Model 100 (see Goding, supra, for basic principles). The IgM antibodies were purified by molecular sizing on a S-300 column (Pharmacia-LKB).

15 All the following characterization information was performed with purified monoclonal antibody.

I. Isoelectric Focusing (IEF)

A cell line quality control to ensure consistency of frozen lots included measuring 20 the antibody pI point on an IEF gel apparatus (Bio-Rad) which separates proteins based on net charge. Briefly, a bis-acrylamide-riboflavin solution was applied to an acrylamide gel, exposed to fluorescent lighting for 1 hour, then stored overnight at 4°C. A 1 µg sample of monoclonal antibody and standards were laid upon the gel and electrophoresed over a 1-2 hour period. Following a series of fixatives and washes, the 25 gel was silver stained (Bio-Rad). The pI value of the monoclonal antibody was calculated by migratory distance through the gel and was directly compared to the protein standards' migratory distance of known pI values. The distinctive finger print banding pattern reflected the pI microheterogeneity between independently produced lots of antibody (Hamilton, R.G., Reimer, C.B., Rodkey, L.S. (1987) Quality control of murine 30 monoclonal antibodies using isoelectric focusing affinity immunoblot analysis. Hybridoma 6:205-217).

J. EIA and Western Blot Specificity of Monoclonal Antibodies

All monoclonal antibodies noted herein were screened on an assortment of 35 available recombinant HCV antigens as disclosed in U. S. Patent Application Serial No. 07/572,822 entitled HEPATITIS C ASSAY UTILIZING RECOMBINANT PROTEINS, which

enjoys common ownership and is incorporated herein by reference. The procedures are outlined above. The multiple antigen screening technique confirmed the HCV specificity and to exclude the HCV non-specific CKS, IPL, or linker-arm reactivity of the monoclonal antibodies.

5

K. EIA Epitope Competition Studies

To investigate specificity and antigen binding distinctions, epitope grouping experiments were performed utilizing biotin labeled and unlabeled monoclonal antibodies (Langone & Van Yunakis, *Methods in Enzymology*, 92:242-253, Academic

10 Press [1983]). Briefly, the antibodies were labeled with NHS-LC-biotin (Pierce Chemical Co., Rockford, IL) according to the manufacturer's instructions. Microtiter wells were coated with the immunogen as previously described. First, \log_2 dilutions of the unlabeled antibody were pre-incubated in the wells for 15 minutes, followed by the addition of a fixed amount of biotinylated antibody (the dilution in a direct EIA of the 15 biotinylated antibody alone which gave a value of 50% of the maximum absorbance value) and incubated for 20 minutes. Plates were washed three times with water.

Diluted streptavidin-HRPO (Zymed, South San Francisco, CA) was added to the wells and incubated for 30 minutes. The plates were washed again and OPD color developed as previously described. The absorbance was read at 492 nm. Antibodies of the same or 20 related epitope had signal blocked or inhibited by >50%. No inhibition was observe with antibodies of distinct specificity. This was performed reciprocally for antibodies produced within HCV core region.

L. RIA Reciprocal Competition

25 Beads coated with the appropriate antigen or peptide were incubated with 100 μ l of unlabeled monoclonal antibody diluted into recalcified negative human plasma (NHP, testing negative for anti-HCV, anti-HIV and HBsAg) at monoclonal antibody concentrations of 1-20 μ g/ml. 100 μ l of radiolabeled antibody at 1 to 4 μ Ci/ml diluted into HTLV 1 kit specimen diluent (containing detergent, animal sera, buffer, available 30 from Abbott Laboratories, Abbott Park, IL) was incubated with the bead for 2 hours at 45°C or 18-20 hours at 20-25°C. Beads were washed and counted for radioactivity.

M. HCV Antigen Assays

Beads coated with one or a cocktail of anti-HCV monoclonal antibody were 35 incubated with 200 μ l of specimen for 2 hours at 40-45°C or 18-20 hours at 20-25°C. Beads were washed with distilled water and then incubated with 200 μ l of

radiolabeled anti-HCV monoclonal antibody (one or more) for 2 hours at 45°C. Beads were washed and counted in a gamma counter.

N. Characterization of Monoclonal Antibody

5 Two monoclonal antibody competition groups against the HCV 33c region (1191-1457) are found in Table 10. The groups reacted with HCV CKS-33c, CKS-33c-CORE, and λ PL-33c-CORE fusion proteins. Referring to FIGS. 6 TO 17, the reactivities summarized below in Table 10 are shown in lanes 1 and 2. Lanes 1 to 3 contains monoclonal antibodies against HCV 33C protein (6-296-534 in lane 1, 6-914-518 in 10 lane 2 and 6-1070-110 in lane 3); lanes 4-6 contain monoclonal antibodies against HCV CORE (13-975-157 in lane 4, 14-153-234 in lane 5 and 14-1350-210 in lane 6); lanes 7 and 8 contain monoclonal antibodies against the putative HCV ENV region (16-407-209 in lane 7 and 16-803-174 in lane 8); lanes 9-11 contain monoclonal 15 antibodies against HCV C-100 (25-1518-105 in lane 9, 28-735-355 in lane 10; lane 11 contains CKS control monoclonal antibody (29-121-236); lane 12 contains a normal mouse serum control; and lane 13 contains a negative control.

TABLE 10
Anti-HCV-33c Monoclonal Antibodies
EIA and Western Blot Reactivity

20	Group #	Cell Line	pI	Isotype	Recombinant HCV Antigen Reactivity						
					CKS-		λ PL-		33c-		CKS-
					core	core	core	core	33c	33c	BCD
25	1	6-296-534	6.0	IgG1 k	-	-	-	-	+	+	+
	2	6-914-518		IgG2b k	-	-	-	-	+	+	+

EXAMPLE 15

Anti-HCV-33c Competitive Assay

30 A one step competitive assay was developed using CKS-33c coated beads (0.1 μ g/ml) coating) and radiolabeled group 1 (6-296-534) and group 2 (6-914-518) monoclonal antibody for detection. Seven anti-C100 false positive sera samples and ten anti-C100 true positive sera samples obtained from the Interstate Blood Bank were tested using the two anti-33c monoclonal antibodies of the invention (Table 2). The false positive specimens gave no more than 25% inhibition for any of the monoclonal antibodies. Binding of monoclonal antibody 6-296-534 was almost completely inhibited by the anti-C100 positive specimen (84-100%). Interestingly, these

specimens were not able to effectively compete group 2 monoclonal antibody for binding to 33c (0-24% inhibition).

Subclones of Table 11 hybrids have been established demonstrating comparable reactivity by EIA and RIA competitive binding studies.

5

TABLE 11
Competitive anti-HCV 33c Assays

<u>Label: 6-296 in HTLV I diluent</u>						
	Specimen	CPM	Avg	S/N	% Inhib.	Result
1.0	NC	72473	70641			
		67993				
		71456				
	Sacramento negative specimens					
1.5	145	55325	0.78	21.7		
	146	65197	0.92	7.7		
	148	53705	0.76	24.0		
	149	57741	0.82	18.3		
2.0	151	89301	1.26	-26.4		
	152	88711	1.26	-25.6		
	154	67721	0.96	4.1		
	Sacramento positive specimens					
2.5	147	5366	0.08	92.4	+	
	150	3351	0.05	95.3	+	
	153	240	0.00	99.7	+	
	156	128	0.00	99.8	+	
	159	3045	0.04	95.7	+	
	160	47	0.00	99.9	+	
3.0	161	11276	0.16	84.0	+	
	163	42	0.00	99.9	+	
	165	1076	0.02	98.5	+	
	167	66	0.00	99.9	+	
<u>Label: 6-914 in HTLV I diluent</u>						
	Specimen	CPM	Avg	S/N	% Inhib.	Result
3.5	NC	32156	34010			
		34431				
		35444				
	Sacramento negative specimens					
4.0	145	38613	1.14	-13.5		
	146	36134	1.06	-6.2		
	148	35130	1.03	-3.3		
	149	30137	0.89	11.4		
4.5	151	36794	1.08	-8.2		
	152	30860	0.91	9.3		
	154	34493	1.01	-1.4		
	Sacramento positive specimens					
	Specimen	CPM	Avg	S/N	% Inhib.	Result
5.0	147	29573	0.87	13.0		
	150	34219	1.01	-0.6		

	153	28002	0.82	17.7	
	156	31866	0.94	6.3	
	159	32633	0.96	4.0	
5	160	30984	0.91	8.9	
	161	34569	1.02	-1.6	
	163	27479	0.81	19.2	
	165	25865	0.76	23.9	

10

EXAMPLE 16HCV-33c Antigen Assay

Results from a two step 33c antigen RIA assay are shown in Table 12. Assay specimens which gave S/N values greater than 4.0 were considered reactive. The assay was able to detect CKS-33c and IPL-33c-CORE at 300 ng/ml and 2.0 µg/ml respectively. Two of seven specimens from patients with elevated ALT and anti-C-100 were reactive for 33c antigen.

TABLE 12

HCV-33c Ag Assay

			(6-914-518 bead/6-296-534.6-1070-110 label)			
	Specimen		CPM	Average	S/N	Result
20		NC	327	312		
			287			
			322			
25	CKS-33c	36 µg/ml	17439	18125	58.09	+
			18810			
		3.6 µg/ml	5256	5133	16.45	+
			5010			
30		360 ng/ml	1401	1340	4.29	+
			1279			
		36 ng/ml	408	412	1.32	-
			415			
		3.6 ng/ml	305	321	1.03	-
			336			
35	IPL-33c-CORE	248 µg/ml	5832	5873	18.82	+
			5913			
		24.8 µg/ml	3607	3545	11.36	+
			3483			
40		2.48 g/ml	1261	1369	4.39	+
			1476			
		248 mg/ml	455	471	1.51	-
			487			
		24.8 ng/ml	310	331	1.06	-
			352			
45	Elevated ALT	27	2239	1999	6.41	+
			1759			
		238	717	516	1.65	-
			315			

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recognize serum Ag both free and bound with human antibody. Furthermore, "cryptic" or hidden antigens or antigenic determinants may be uncovered by treatment of specimen with detergent or reducing agent or both. For example, CORE antigen may exist in a capsid form covered by the virus envelope. Stripping the envelope with detergent should expose CORE antigen. Monoclonal antibodies may also offer pragmatic advantages over high titer polyclonal antibody in giving greater sensitivity in assay or allowing shorter incubation times.

Further, antibody immunoassays, one or two step competitive assays, were developed in which anti-HCV competed with labeled anti-HCV monoclonal antibody for binding to a limited number of antigenic sites. A more sensitive competitive assay may be developed in which human anti-HCV binds to HCV Ag in solution blocking or inhibiting the HCV Ag binding in HCV Ag sandwich assay. Competitive assays using monoclonal antibodies allow a more precise mapping of human antibody epitopes and may be useful for determining virus neutralizing antibody epitopes. Some monoclonal antibodies may have virus neutralizing activity. Finally, monoclonal antibodies should be useful in immunoaffinity purification of native viral and recombinant HCV antigens and proteins.

The hybridomas which produce the monoclonal antibodies of the invention are identified as hybridoma H81C17 producing monoclonal antibody H81C17, hybridoma H35C54 producing monoclonal antibody H35C54, hybridoma H28C110 producing monoclonal antibody H28C110, hybridoma H4C20 producing monoclonal antibody H4C20, hybridoma H11C130 producing monoclonal antibody H11C130 and hybridoma H1C46 producing monoclonal antibody H1C46. Hybridomas H28C110, H81C17 and H11C130 were deposited at the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852 as of October 30, 1990, and have been accorded the following deposit numbers: H28C110 was accorded A.T.C.C. deposit No. HB 10587; H81C17 was accorded A.T.C.C. deposit No. HB10588 and H11C130 was accorded A.T.C.C. deposit No. HB 10589. Hybridomas H35C54, H4C20 and H1C46 were deposited at the American Type Culture Collection, 10231 Parklawn Drive, Rockville, Maryland 20852 as of October 31, 1990, and were accorded the following deposit numbers: H35C54 was accorded A.T.C.C. deposit No. HB 10592; H4C20 was accorded A.T.C.C. deposit No. HB 10593 and H1C46 was accorded A.T.C.C. deposit No. HB 10594.

Also, the hybridoma cell lines which produce the monoclonal antibodies of the invention are identified as hybridoma cell line 13-975-157 (producing monoclonal

antibody 13-975-157), hybridoma cell line 14-153-234 (producing monoclonal antibody 14-153-234) and hybridoma cell line 14-1350-210 (producing monoclonal antibody 14-1350-210). These hybridoma cell lines were deposited at the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852 on

5 November 16, 1990 and were accorded the following deposit numbers: Hybridoma cell line 13-975-157 was accorded A.T.C.C. deposit No. HB 10608, hybridoma cell line 14-153-234 was accorded A.T.C.C. deposit No. HB 10604, and hybridoma cell line 14-1350-210 was accorded A.T.C.C. deposit No. HB 10602.

10 Further, the hybridoma cell lines which secrete the monoclonal antibodies of the invention are identified as hybridoma cell line 6-296-534 (secreting monoclonal antibody 6-296-534) and hybridoma cell line 6-914-518 (secreting hybridoma cell line 6-914-518). These hybridoma cell lines were deposited at the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852 on November
15 16, 1990 and were accorded the following deposit numbers: Hybridoma cell line 6-296-534 was accorded A.T.C.C. deposit No. HB 10607, and hybridoma cell line 6-914-518 was accorded A.T.C.C. deposit No. HB 10600.

Other variations of applications of the use of the unique monoclonal antibodies provided herein include the detection of HCV antigen in immune complexes, or latent and/or cryptic antigens, and/or associated with viral nucleic acid for detection of the nucleic acid by PCR, LCR, or by direct hybridization. Still other variations and modifications of the specific embodiments of the invention as set forth herein will be apparent to those skilled in the art. Accordingly, the invention is intended to be limited
25 only in accordance with the appended claims.

TABLE 1
CHARACTERIZATION OF MONOCLONAL ANTIBODIES TO HCV PROTEINS

MAB ID	Immunogen	Western Blot Reactivity ¹	EIA Titer With Purified IgG ²	Isotype	Competition w/ Immune Human Sera ³	Epitope Specificity a.a. of HCV genome ⁴
H81C17	pHCV34 (HCV-CORE)	+++	+++	16	3.9	IgG1, k
H35C54	pHCV34	+++	++	31	7.8	IgG1, k
H28C110	pHCV23 (HCV C-100)	+++	+++	0.5	0.4	IgG1, k
H4C20	pHCV23 (HCV C-100)	+++	+++	125	30	IgG1, k
H11C130	pHCV29 (HCV 33C)	+++	+++	0.5	0.5	IgG1, k
H1C46	pHCV29 (HCV 33C)	+++	+	0.5	63	IgG1, k

¹ Data reflects the reactivity of one each monoclonal antibody with the respective immunogen expressed as (a) CKS fusion protein and (b) under APL promoter without any fusion protein in *E. coli*. +++ indicates strong reactivity; + indicates weak reactivity.

² Data reflects the reactivity of each monoclonal with the immunogen as described in 1. The EIA Titer is defined as the concentration of the monoclonal IgG protein in ng/ml which will give the absorbance of 4 times the negative control at 492 nm.

³ +++ indicates strong competition (>80%); + indicates weak competition (approximately 50%).

⁴ Epitope specificity was determined based on several experiments as described in Examples 5 and 6.

WE CLAIM:

1. A monoclonal antibody which specifically binds to HCV protein C-100 and does not significantly bind to HCV proteins 33C and CORE.
2. The monoclonal antibody of claim 1 secreted by a hybridoma selected from the group consisting of A.T.C.C. deposit Nos. HB 10593 and HB 10587.
3. A monoclonal antibody which specifically binds to HCV protein 33C and does not significantly bind to HCV proteins C-100 and CORE.
4. The monoclonal antibody of claim 3 secreted by a hybridoma selected from the group consisting of A.T.C.C. deposit Nos. HB 10594, HB 10589, HB 10607 and HB 10600.
5. A monoclonal antibody which specifically binds to HCV protein CORE and does not significantly bind to HCV proteins C-100 and 33C.
6. The monoclonal antibody of claim 5 secreted by a hybridoma selected from the group consisting of A.T.C.C. deposit Nos. HB 10592, HB 10588, HB 10608, HB 10604 and HB 10602.
7. A hybridoma which produces a monoclonal antibody which specifically binds to HCV protein C-100 and does not significantly bind to HCV 33C and HCV CORE proteins.
8. The hybridoma of claim 7 selected from the group consisting of A.T.C.C. deposit Nos. HB 10593 and HB 10587.
9. A hybridoma which produces a monoclonal antibody which specifically binds to HCV protein 33C and does not significantly bind to HCV C-100 and HCV CORE proteins.
10. The hybridoma of claim 9 selected from the group consisting of A.T.C.C. deposit Nos. HB 10594, HB 10589, HB 10607 and HB 10600.

11. A hybridoma which produces a monoclonal antibody which specifically binds to HCV protein CORE and does not significantly bind to HCV C-100 and HCV 33C proteins.

12. The hybridoma of claim 11 selected from the group consisting of A.T.C.C. deposit Nos. HB 10592, HB 10588, HB 10608, HB 10604 and HB 10602.

13. A method for determining the presence of Hepatitis C Virus HCV C-100 protein, or HCV 33C protein or HCV CORE protein in a test sample which may contain the Hepatitis C Virus, comprising:

- a. contacting the test sample with an anti-HCV antibody which specifically binds to HCV C-100 protein, or HCV 33C protein or HCV CORE protein attached to a solid phase which antibody specifically binds to HCV protein, to form a mixture;
- b. incubating said mixture for a time and under conditions sufficient to form antigen/antibody complexes;
- c. contacting said complexes with an indicator reagent comprising a signal generating compound capable of generating a measurable detectable signal attached to an anti-HCV antibody which specifically binds to HCV C-100 protein, or HCV 33C protein or HCV CORE protein to form a second mixture;
- d. incubating said second mixture for a time and under conditions sufficient to form antibody/antigen/antibody complexes; and
- e. determining the presence of Hepatitis C Virus in the test sample by detecting the measurable signal generated, wherein the amount of HCV C-100 protein, or HCV 33C protein or HCV CORE protein present in the test sample is proportional to said measurable signal.

14. A competitive assay method for determining the presence and amount of HCV antibody which may be present in a test sample, comprising:

- a. contacting a test sample suspected of containing HCV antibodies with a solid phase coated with HCV 33C, C-100 or HCV CORE proteins and an indicator reagent comprising a signal generating compound and a monoclonal antibody which specifically binds to said proteins, for a time and under conditions sufficient to form antigen/antibody complexes of the test sample and solid phase and/or indicator reagent and solid phase;

b. determining the presence of HCV antibody present in the test sample by detecting the reduction in binding of the indicator reagent to the solid phase as compared to the signal generated from a negative test sample to indicate the presence of HCV antibody in the test sample.

15. The method of claim 13 wherein step (a) and (c) are performed simultaneously.

16. The method of claim 13 wherein the anti-HCV antibody attached to the solid phase is a polyclonal antibody.

17. The method of claim 13 or 14 wherein the signal generating compound is selected from the group consisting of a luminescent compound, a chemiluminescent compound, an enzyme and a radioactive element.

18. The method of claim 13 wherein said anti-HCV antibody attached to the solid phase is a monoclonal antibody.

19. The method of claim 13 wherein the indicator reagent comprises a signal generating compound attached to a polyclonal antibody.

20. The method of claim 13 wherein said indicator reagent comprises a signal generating compound attached to a monoclonal antibody.

21. The method of either one of claims 14, 18 or 20 wherein said monoclonal antibody is the monoclonal antibody secreted by a hybridoma selected from the group consisting of A.T.C.C. deposit Nos. HB 10593, HB 10587, HB 10594, HB 10589, HB 10592, HB 10588, HB 10607, HB 10600, HB 10608, HB 10604 and HB 10602.

22. An assay kit for detecting the presence of HCV in a test sample comprising: a container containing a monoclonal antibody which specifically binds either HCV C-100 protein, HCV 33C protein or HCV CORE protein.

23. The assay kit of claim 22 wherein said monoclonal antibody is secreted by a hybridoma selected from the group consisting of A.T.C.C. deposit Nos. HB 10593, HB

10587, HB 10594, HB 10589, HB 10592, HB 10588, HB 10607, HB 10600, HB 10608, HB 10604 and HB 10602, or a combination thereof.

24. A hybridoma cell line selected from the group consisting of A.T.C.C. deposit Nos. HB 10593, HB 10587, HB 10594, HB 10589, HB 10592, HB 10588, HB 10607, HB 10600, HB 10608, HB 10604 and HB 10602.

25. A monoclonal antibody selected from the group consisting of A.T.C.C. deposit Nos. HB 10593, HB 10587, HB 10594, HB 10589, HB 10592, HB 10588, HB 10607, HB 10600, HB 10608, HB 10604 and HB 10602.

1/14

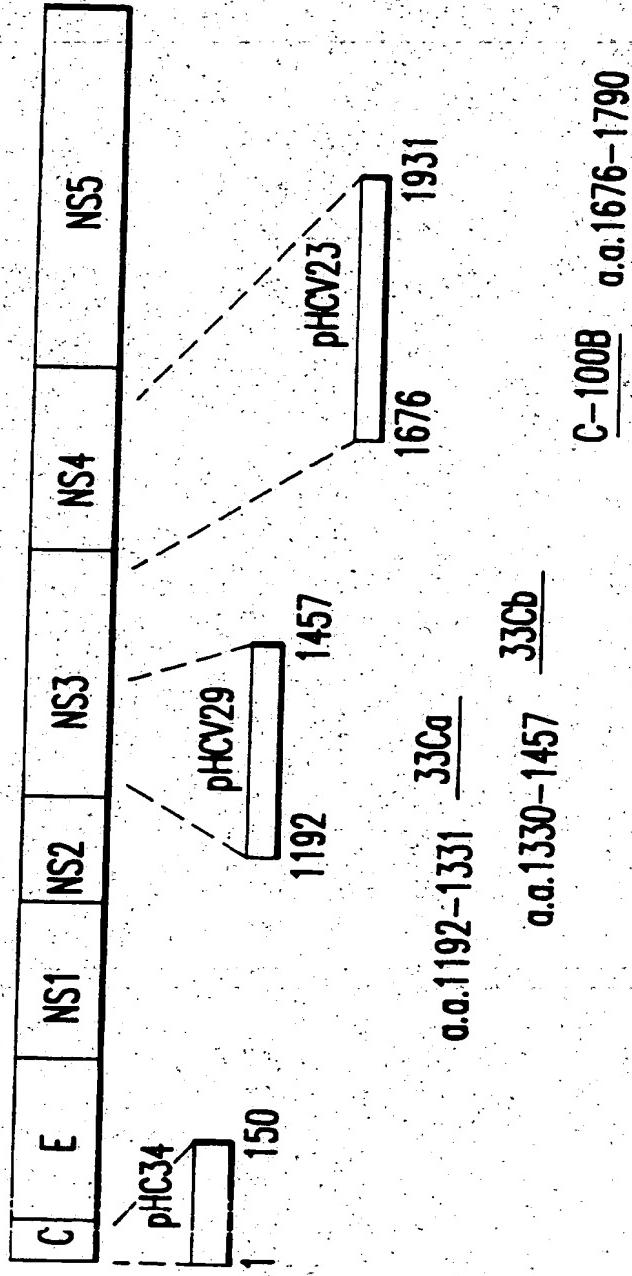
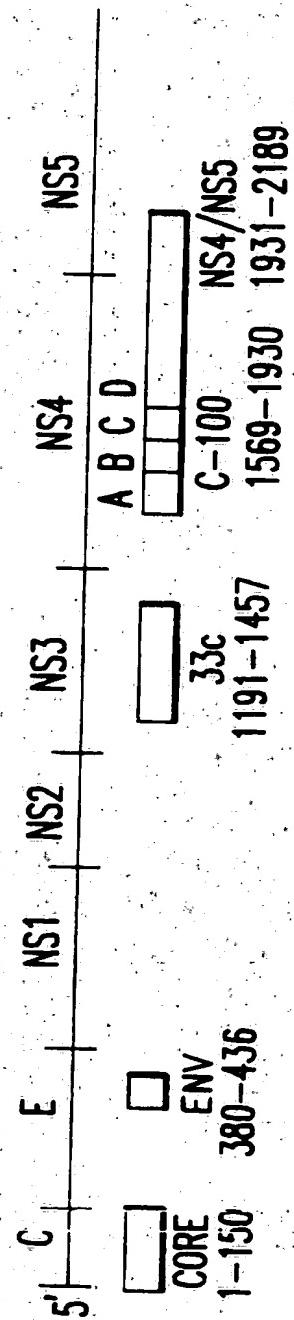


FIG. 1A

2/14

FIG. 1B



3/14

1 2 3 4 5

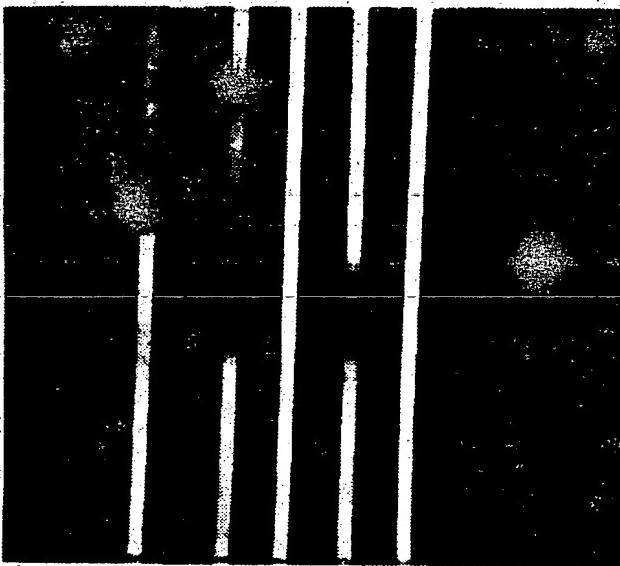


FIG. 2A

1 2 3 4 5

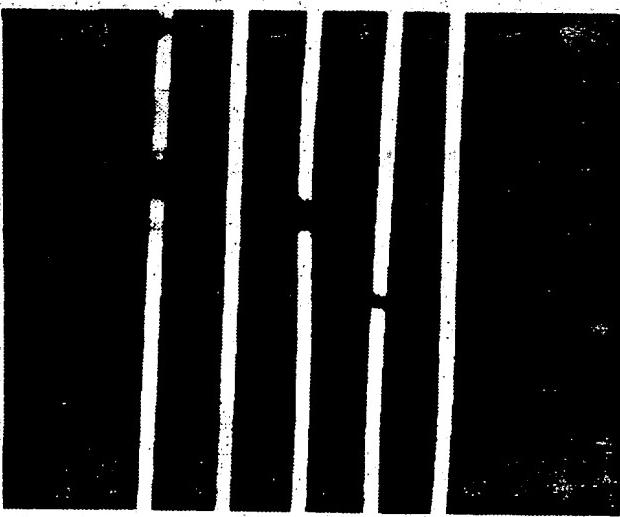


FIG. 2B

4/14

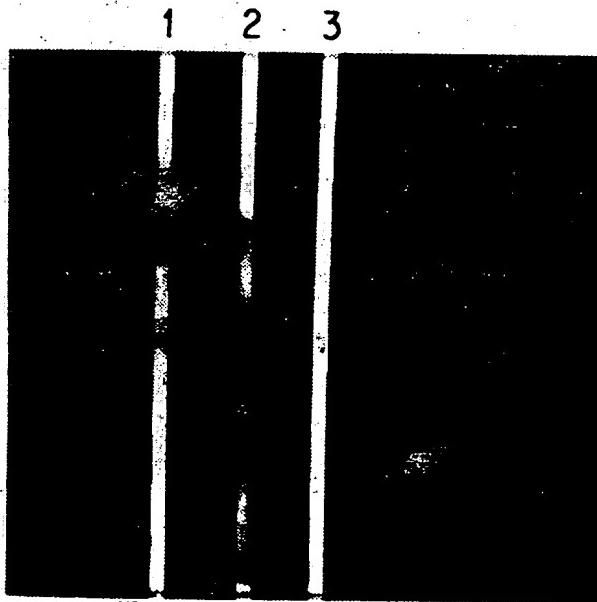


FIG. 2C

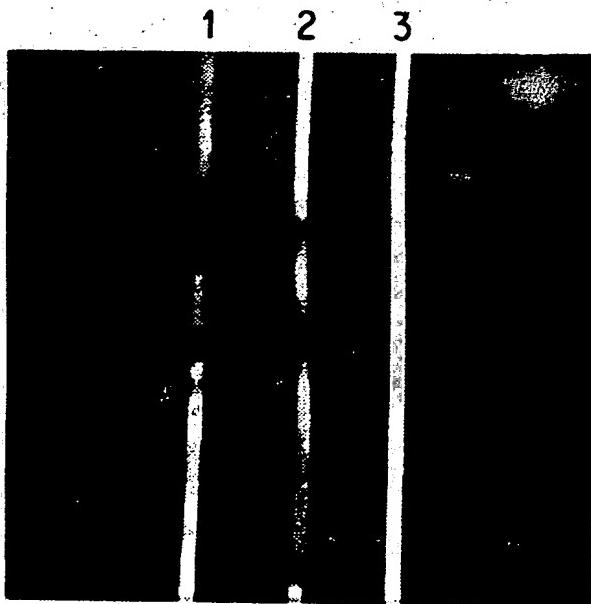


FIG. 2D

5/14

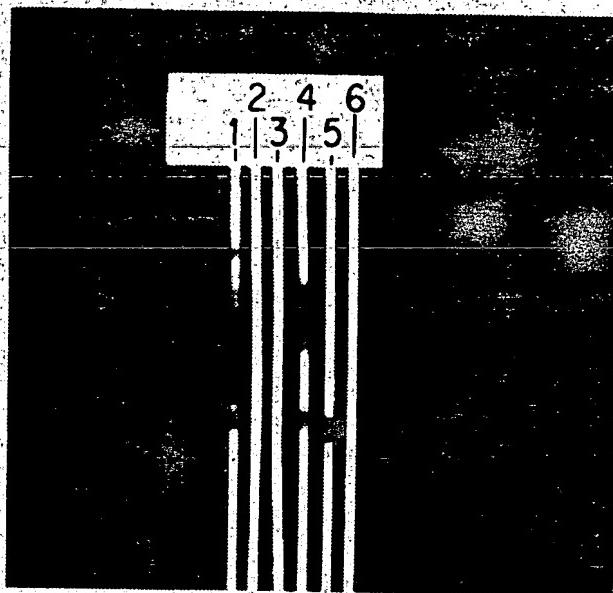


FIG. 2E

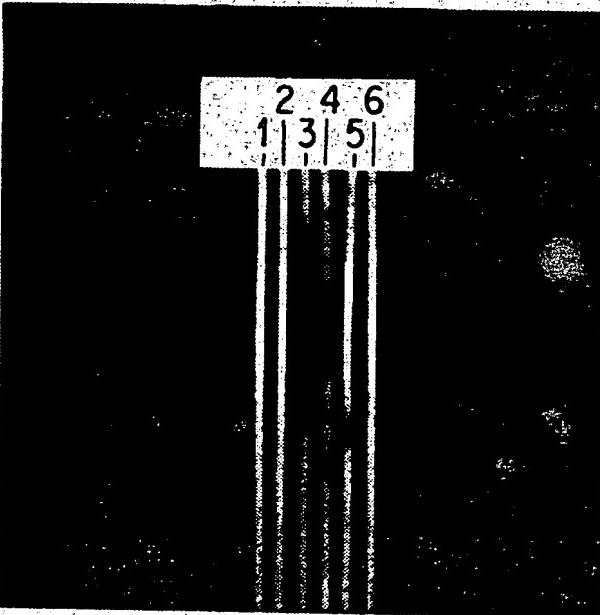


FIG. 2F

6/14

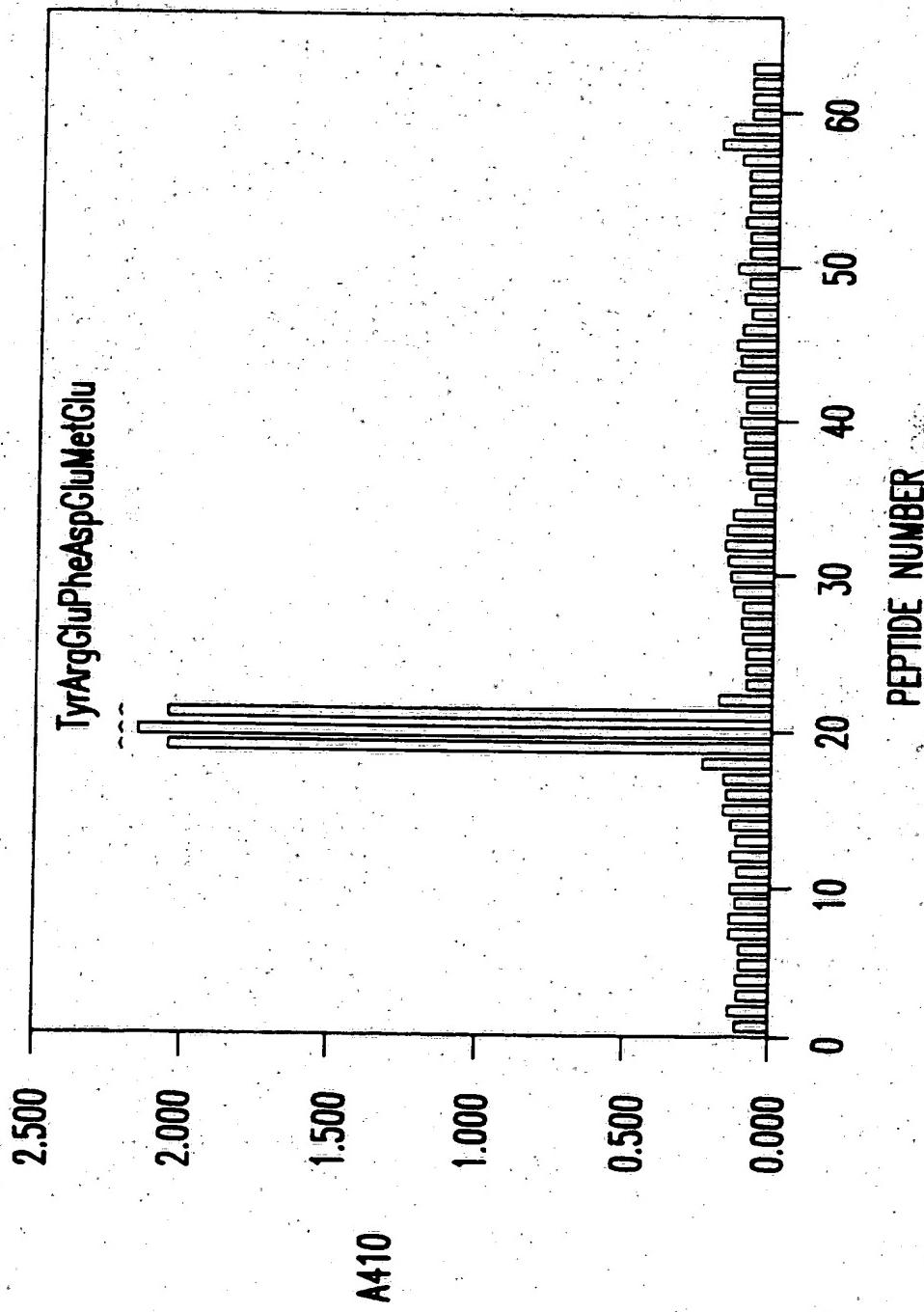


FIG.3

7/14

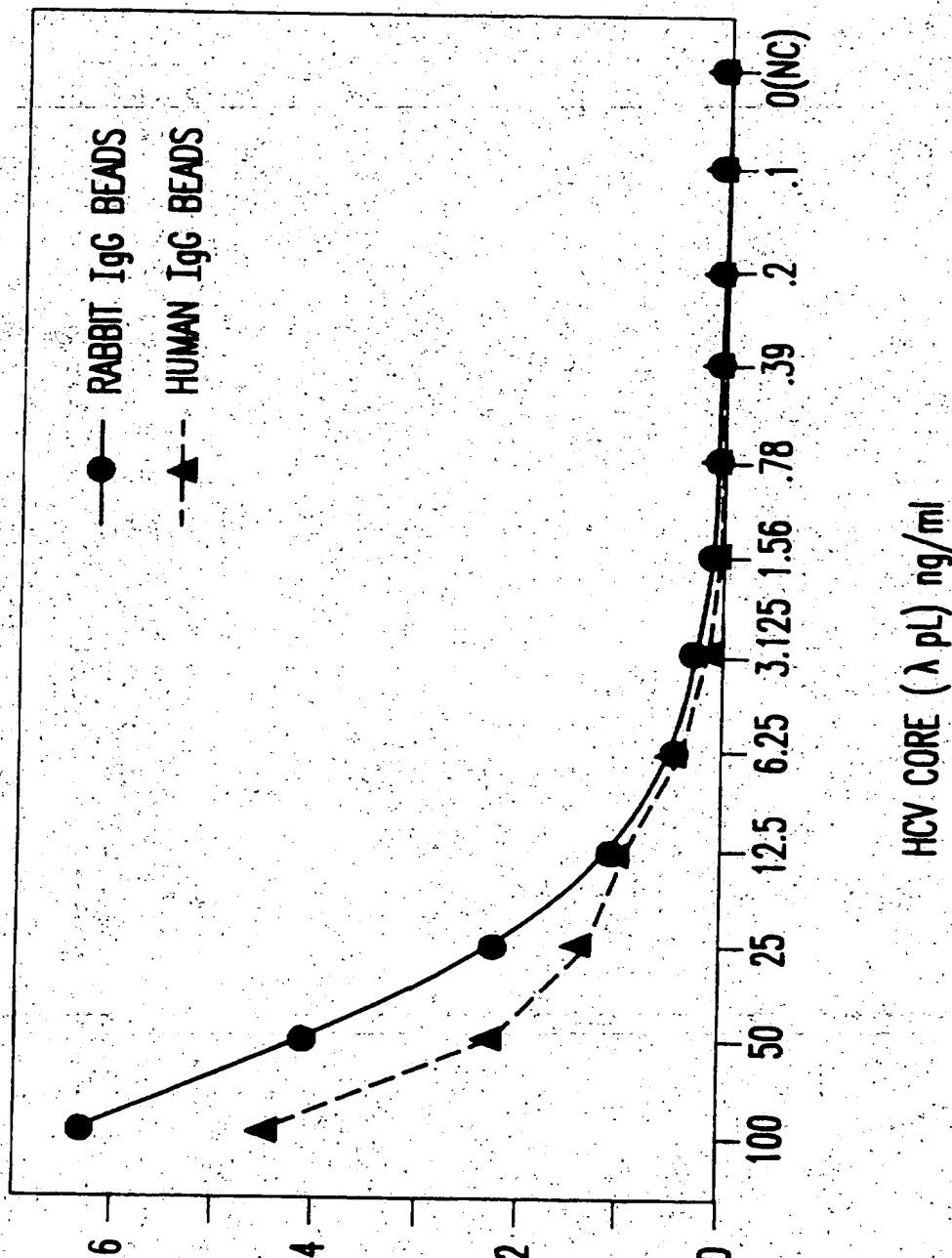
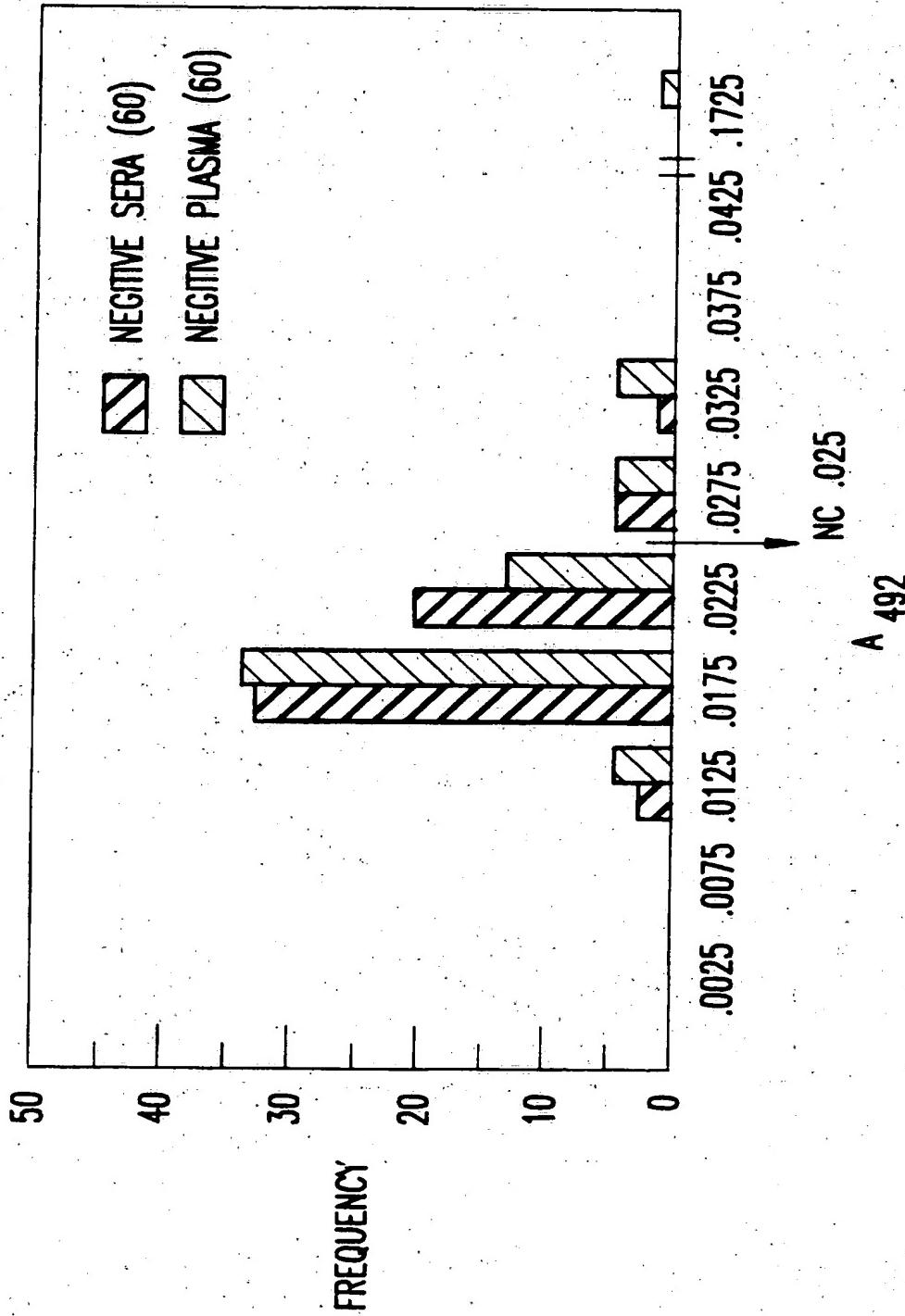


FIG. 4

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8/14

FIG. 5



9/14.

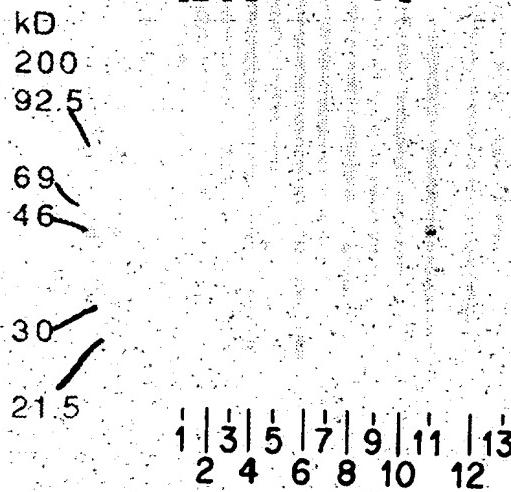


FIG. 6

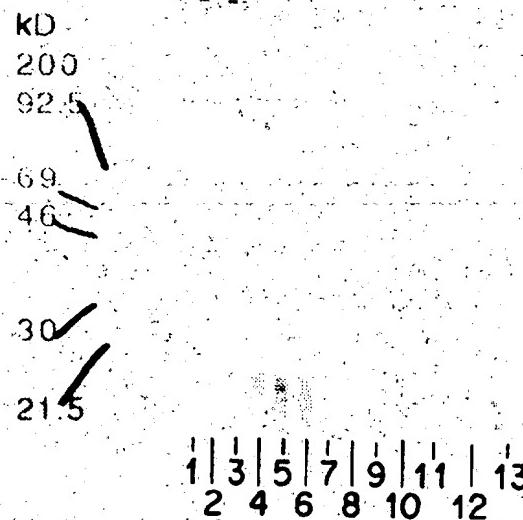
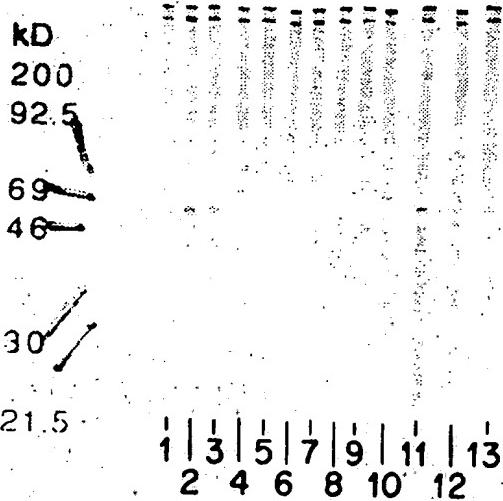
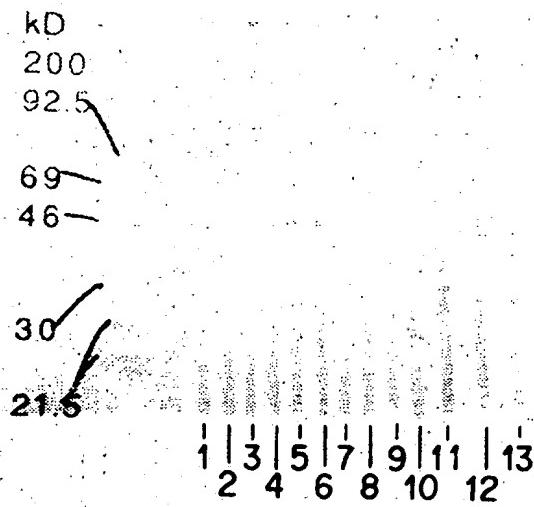


FIG. 7

10/14



11/14

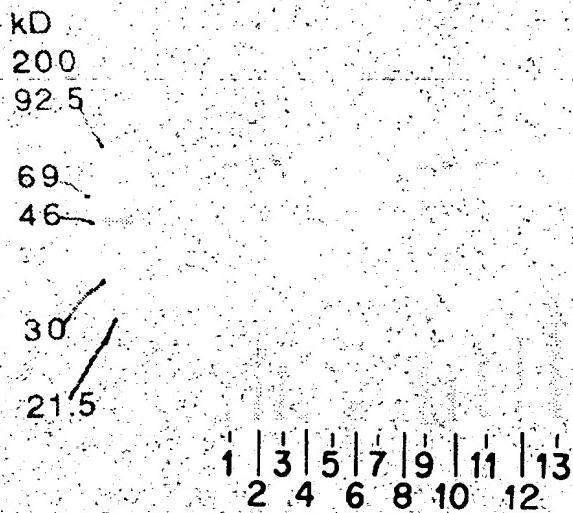


FIG. 10

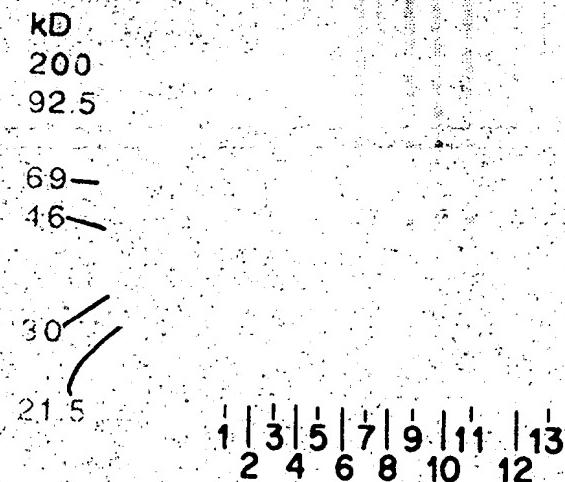


FIG. 11

12/14

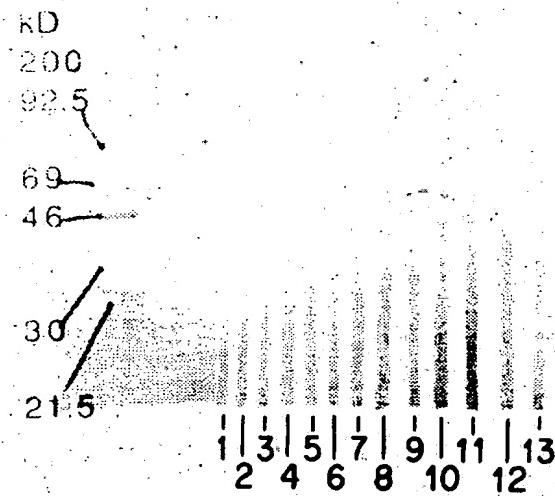


FIG. 12

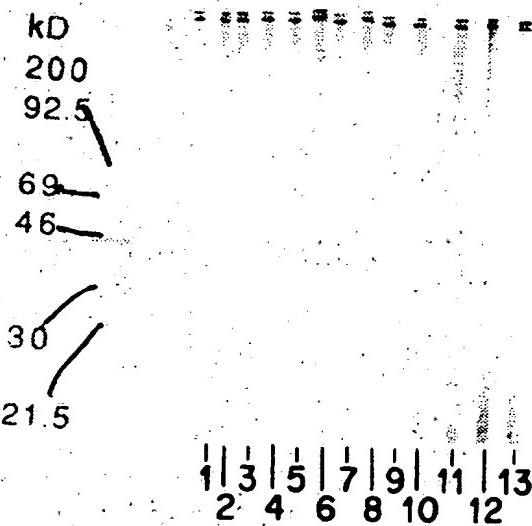


FIG. 13

13/14

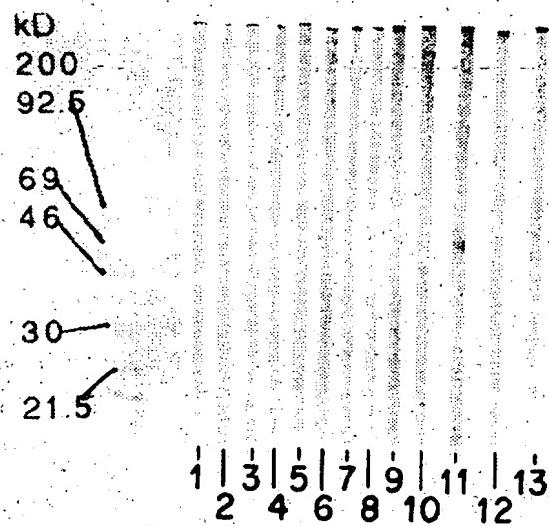


FIG. 14

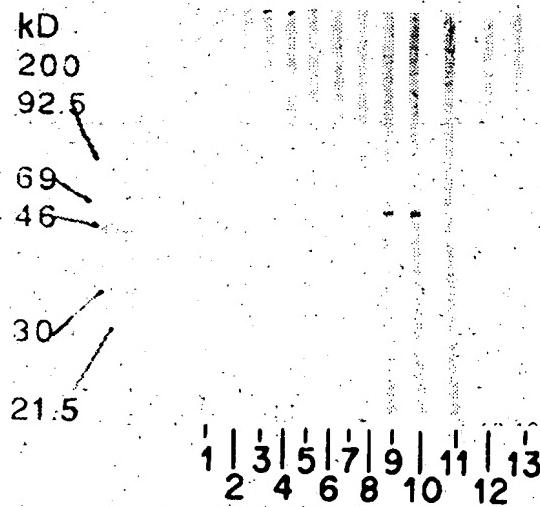


FIG. 15

14 / 14

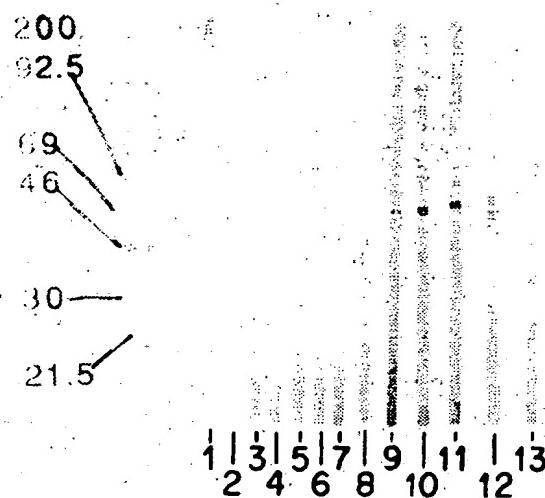


FIG. 16

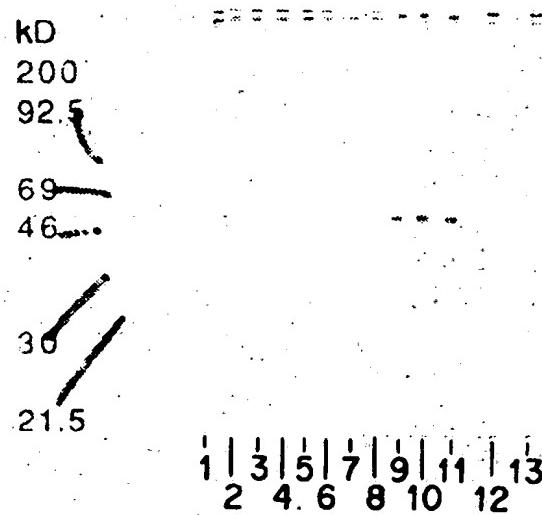


FIG. 17

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US91/08273

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) *

According to International Patent Classification (IPC) or to both National Classification and IPC

U.S.CI.: 530/387; 435/5, 70.21, 240.27

IPC(5): C07K 15/00; C12Q 1/70; C12P 21/02; C12N 5/00

II. FIELDS SEARCHED

Minimum Documentation Searched ?

Classification System	Classification Symbols
U.S.CI.	530/387; 435/5, 70.21, 240.27; 436/548

Documentation Searched other than Minimum Documentation
to the Extent that such Documents are Included in the Fields Searched *

APS
Dialog

III. DOCUMENTS CONSIDERED TO BE RELEVANT *

Category *	Citation of Document, ** with indication, where appropriate, of the relevant passages ***	Relevant to Claim No. ****
X	EP.A. 0,318,216 (Houghton et al.), 31 May 1989, see Claims 1-32	1,7,13-20,22 2-6,8-11,21,23-25
Y	US.A. 4,486,530 (David et al.) 04 December 1984, see entire document.	13, 15-23

* Special categories of cited documents: ¹⁰

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"Z" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search

04 February 1992

International Searching Authority

ISA/US

Date of Mailing of this International Search Report

02 MAR 1992

Signature of Authorized Officer

J. Wortsman
Donna C. Wortsman

F02

FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

V. OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE:

This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:

1. Claim numbers _____ because they relate to subject matter¹² not required to be searched by this Authority, namely:

2. Claim numbers _____ because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out¹³, specifically:

3. Claim numbers _____, because they are dependent claims not drafted in accordance with the second and third sentences of PCT Rule 6.4(a).

VI. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING:

The International Searching Authority found multiple inventions in this international application as follows:

- I. Claims 1, 2, 7, 8, 13-24, drawn to a monoclonal antibody and method of use, classified in class 530, subclass 387, and class 435, subclass 5. See attached sheet.

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.

2. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:

3. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:

4. As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not invite payment of any additional fee.

Remarks on Protest:

- The additional search fees were accompanied by applicant's protest.
 No protest accompanied the payment of additional search fees.

